

CSEP 590 B

Markov Models and Hidden
Markov Models



http://upload.wikimedia.org/wikipedia/commons/b/ba/Calico_cat

Dosage Compensation and X-Inactivation

2 copies (mom/dad) of each chromosome 1-23

Mostly, both copies of each gene are expressed

E.g., A B O blood group defined by 2 alleles of 1 gene

Women (XX) get double dose of X genes (vs XY)?

So, early in embryogenesis:

- One X randomly inactivated in each cell
 - Choice maintained in daughter cells
- } How?

Calico: major coat color gene is on X

Reminder: Proteins “Read” DNA

E.g.:

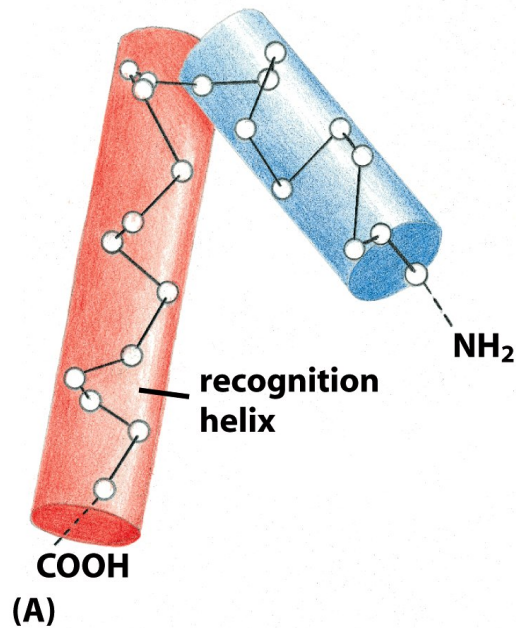


Figure 7-10 Molecular Biology of the Cell 5/e (© Garland Science 2008)

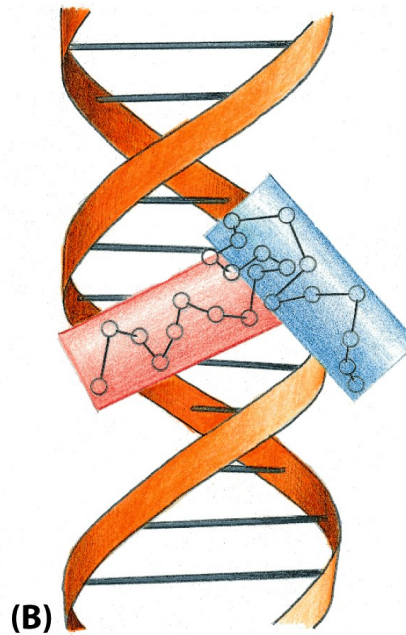


Figure 7-19 Molecular Biology of the Cell 5/e (© Garland Science 2008)



Down in the Groove

Different patterns of hydrophobic methyls, potential H bonds, etc. at edges of different base pairs. They're accessible, esp. in major groove

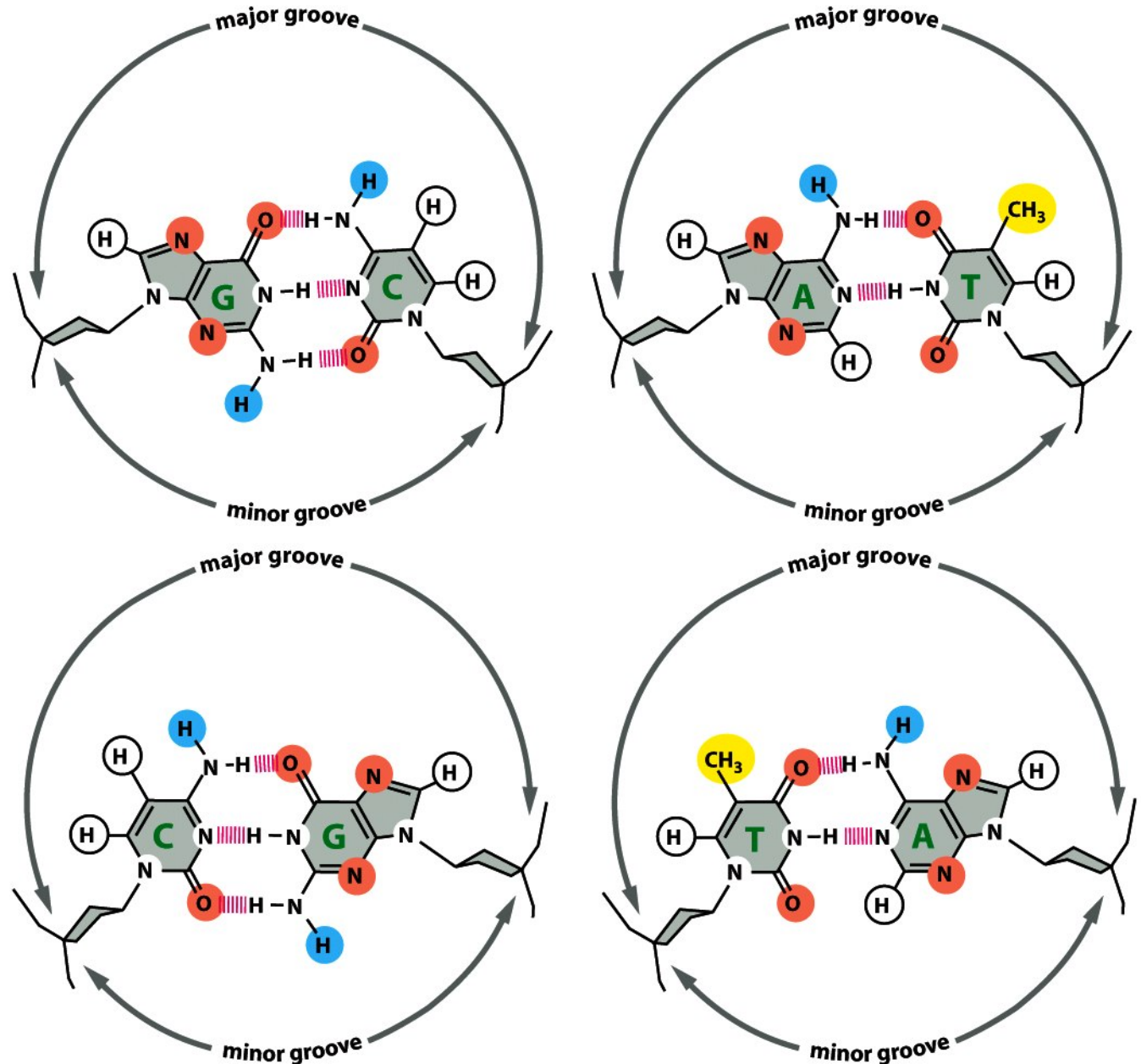
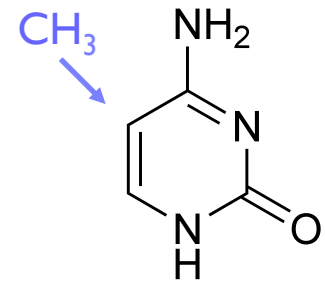


Figure 7-7 Molecular Biology of the Cell 5/e (© Garland Science 2008)

DNA Methylation

CpG - 2 adjacent nts, same strand
(not Watson-Crick pair; “p” mnemonic for the phosphodiester bond of the DNA backbone)

C of CpG is often (70-80%) methylated in mammals i.e., CH₃ group added (both strands)



cytosine

Same Pairing

Methyl-C
alters major
groove
profile (\therefore TF
binding), but
not base-
pairing,
transcription
or replication

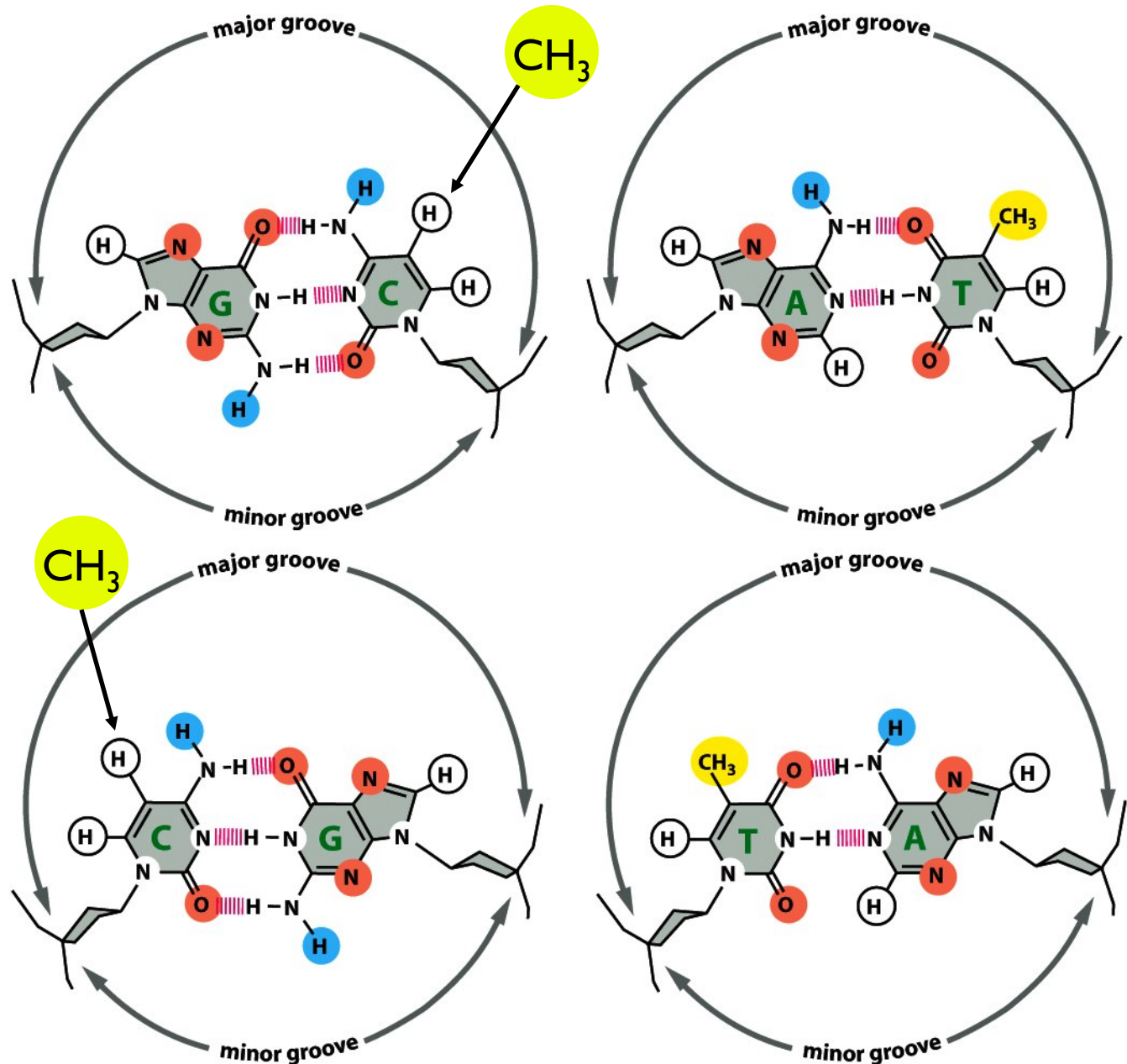


Figure 7-7 Molecular Biology of the Cell 5/e (© Garland Science 2008)

DNA Methylation—Why

In vertebrates, it generally silences transcription
(Epigenetics) X-inactivation, imprinting, repression of mobile elements, cancers, aging, and developmental differentiation

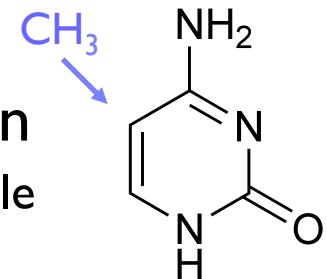
E.g., if a stem cell divides, one daughter fated to be liver, other kidney, need to

- (a) turn off liver genes in kidney & vice versa,
- (b) remember that through subsequent divisions

How? One way:

- (a) Methylate genes, esp. promoters, to silence them
- (b) after ÷, DNA methyltransferases convert hemi- to fully-methylated (& deletion of methyltransferase is embryonic-lethal in mice)

Major exception: promoters of housekeeping genes



cytosine

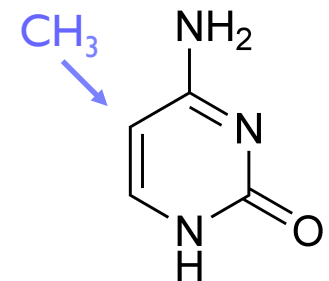
“CpG Islands”

Methyl-C mutates to T relatively easily

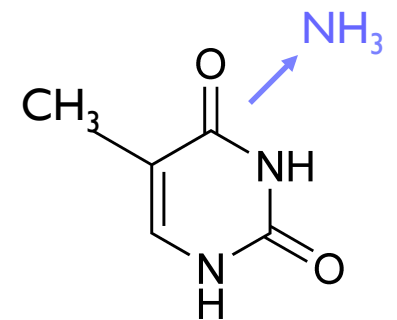
Net: CpG is less common than
expected genome-wide:

$$f(\text{CpG}) < f(\text{C}) * f(\text{G})$$

BUT in some regions (e.g. active
promoters), CpG remain
unmethylated, so CpG → TpG less
likely there: makes “CpG Islands”;
often mark gene-rich regions



cytosine



thymine

CpG Islands

CpG Islands

More CpG than elsewhere (say, $\text{CpG}/\text{GpC} > 50\%$)

More C & G than elsewhere, too (say, $\text{C} + \text{G} > 50\%$)

Typical length: few 100 to few 1000 bp

Questions

Is a short sequence (say, 200 bp) a CpG island or not?

Given long sequence (say, 10-100kb), find CpG islands?

Markov & Hidden Markov Models

References (see also online reading page):

Eddy, "What is a hidden Markov model?" *Nature Biotechnology*, 22, #10 (2004) 1315-6.

Durbin, Eddy, Krogh and Mitchison, "Biological Sequence Analysis", Cambridge, 1998 (esp. chs 3, 5)

Rabiner, "A Tutorial on Hidden Markov Models and Selected Application in Speech Recognition," *Proceedings of the IEEE*, v 77 #2, Feb 1989, 257-286

Independence

A key issue: Previous models we've talked about assume *independence* of nucleotides in different positions - definitely unrealistic.

Markov Chains

A sequence x_1, x_2, \dots of random variables is a *k -th order Markov chain* if, for all i , i^{th} value is independent of all but the previous k values:

$$P(x_i \mid x_1, x_2, \dots, x_{i-1}) = P(x_i \mid x_{i-k}, x_{i-k+1}, \dots, x_{i-1})$$

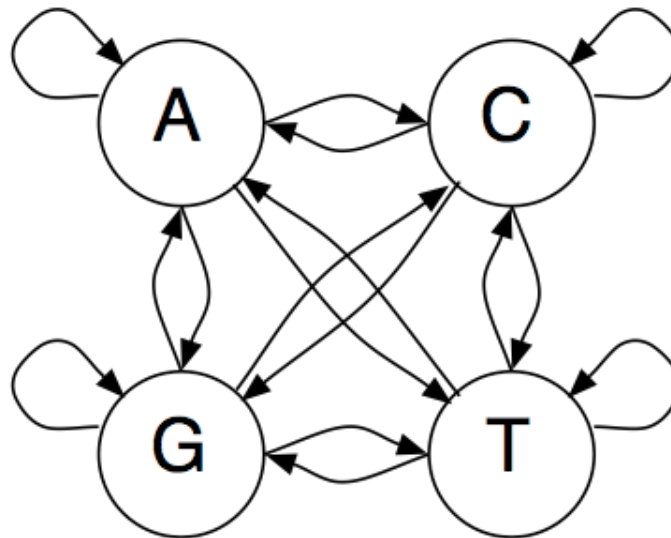
Example 1: Uniform random ACGT

Example 2: Weight matrix model

Example 3: ACGT, but \downarrow Pr(G following C)

} 0th
order
}
1st
order

A Markov Model (1st order)

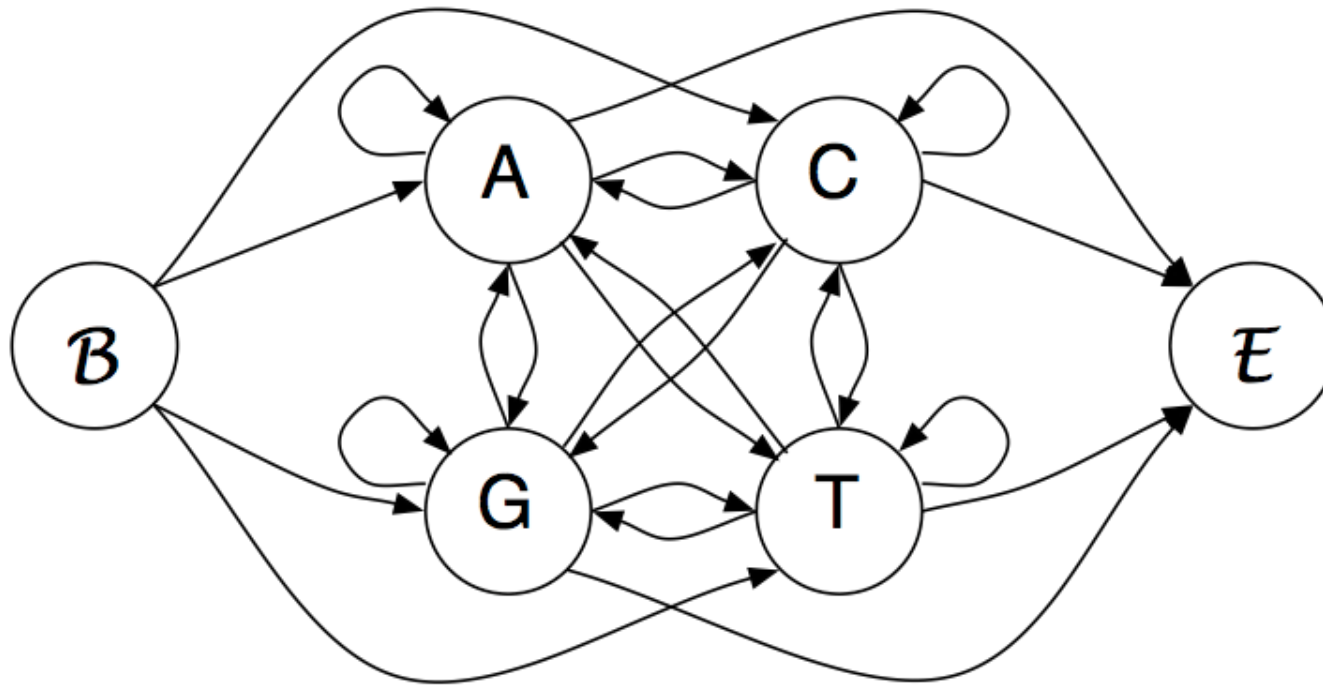


States: A,C,G,T

Emissions: corresponding letter

Transitions: $a_{st} = P(x_i = t \mid x_{i-1} = s)$ ← 1st order

A Markov Model (1st order)



States: A,C,G,T

Emissions: corresponding letter

Transitions: $a_{st} = P(x_i = t \mid x_{i-1} = s)$

Begin/End states

Pr of emitting sequence x

$$x = x_1 x_2 \dots x_n$$

$$P(x) = P(x_1, x_2, \dots, x_n)$$

laws of probability

$$= P(x_1) \cdot P(x_2 \mid x_1) \cdots P(x_n \mid x_{n-1}, \dots, x_1)$$

$$= P(x_1) \cdot P(x_2 \mid x_1) \cdots P(x_n \mid x_{n-1})$$

if 1st
order MC

$$= P(x_1) \prod_{i=1}^{n-1} a_{x_i, x_{i+1}}$$

$$= \prod_{i=0}^{n-1} a_{x_i, x_{i+1}} \quad (\text{with Begin state})$$

Training

Max likelihood estimates for transition probabilities are just the frequencies of transitions when emitting the training sequences

E.g., from 48 CpG islands in 60k bp:

+	A	C	G	T	-	A	C	G	T
A	0.180	0.274	0.426	0.120	A	0.300	0.205	0.285	0.210
C	0.171	0.368	<u>0.274</u>	0.188	C	0.322	0.298	<u>0.078</u>	0.302
G	0.161	0.339	0.375	0.125	G	0.248	0.246	0.298	0.208
T	0.079	0.355	0.384	0.182	T	0.177	0.239	0.292	0.292

From DEKM

Discrimination/Classification

Log likelihood ratio of CpG model vs background model

$$S(x) = \log \frac{P(x|\text{model } +)}{P(x|\text{model } -)} = \sum_{i=1}^L \log \frac{a_{x_{i-1}x_i}^+}{a_{x_{i-1}x_i}^-} = \sum_{i=1}^L \beta_{x_{i-1}x_i}$$

β	A	C	G	T
A	−0.740	0.419	0.580	−0.803
C	−0.913	0.302	1.812	−0.685
G	−0.624	0.461	0.331	−0.730
T	−1.169	0.573	0.393	−0.679

CpG Island Scores

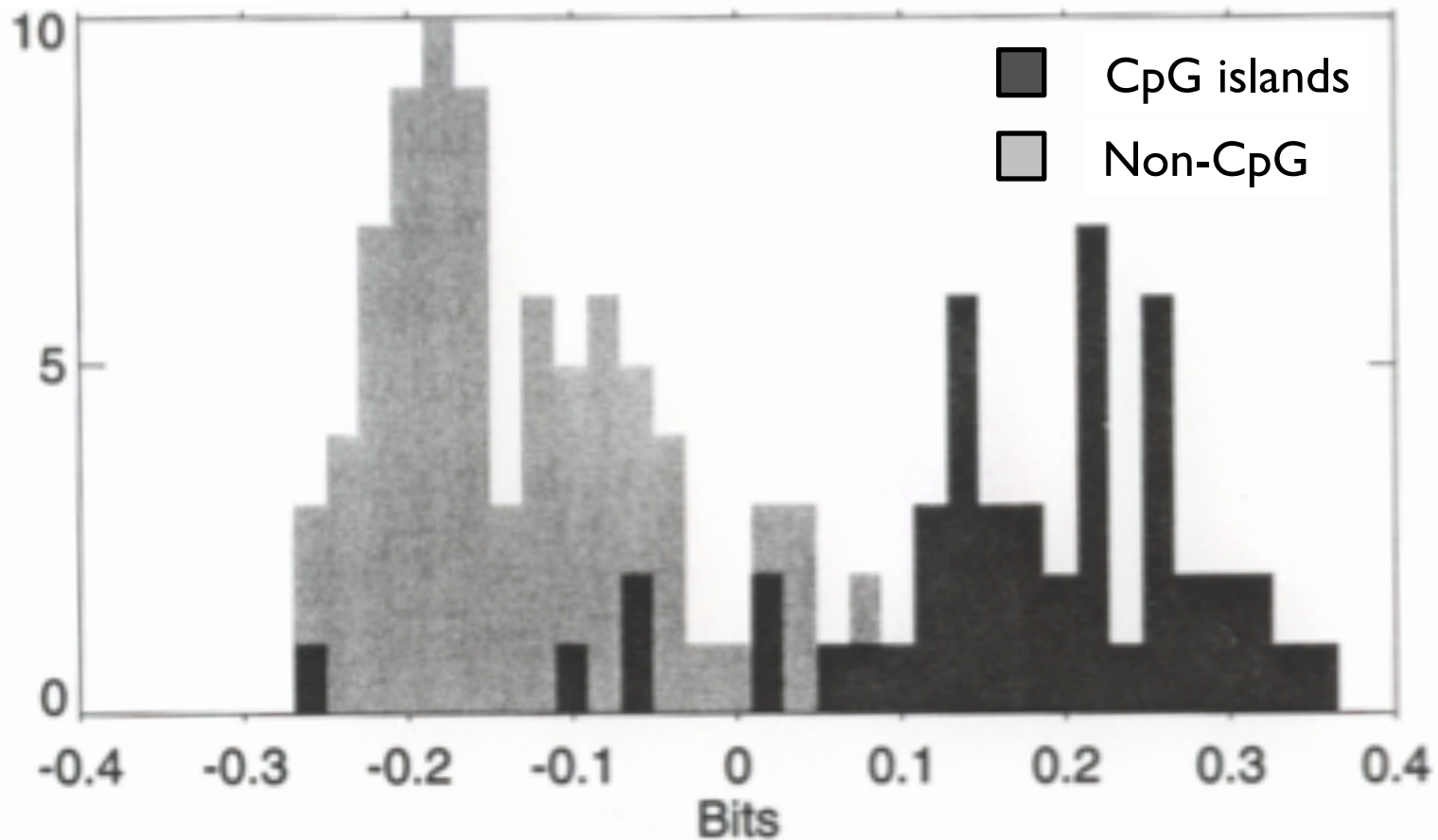


Figure 3.2 *Histogram of length-normalized scores.*

What does a 2nd order
Markov Model look like?

3rd order?

Questions

Q1: Given a *short* sequence, is it more likely from feature model or background model? *Above*

Q2: Given a *long* sequence, where are the features in it (if any)

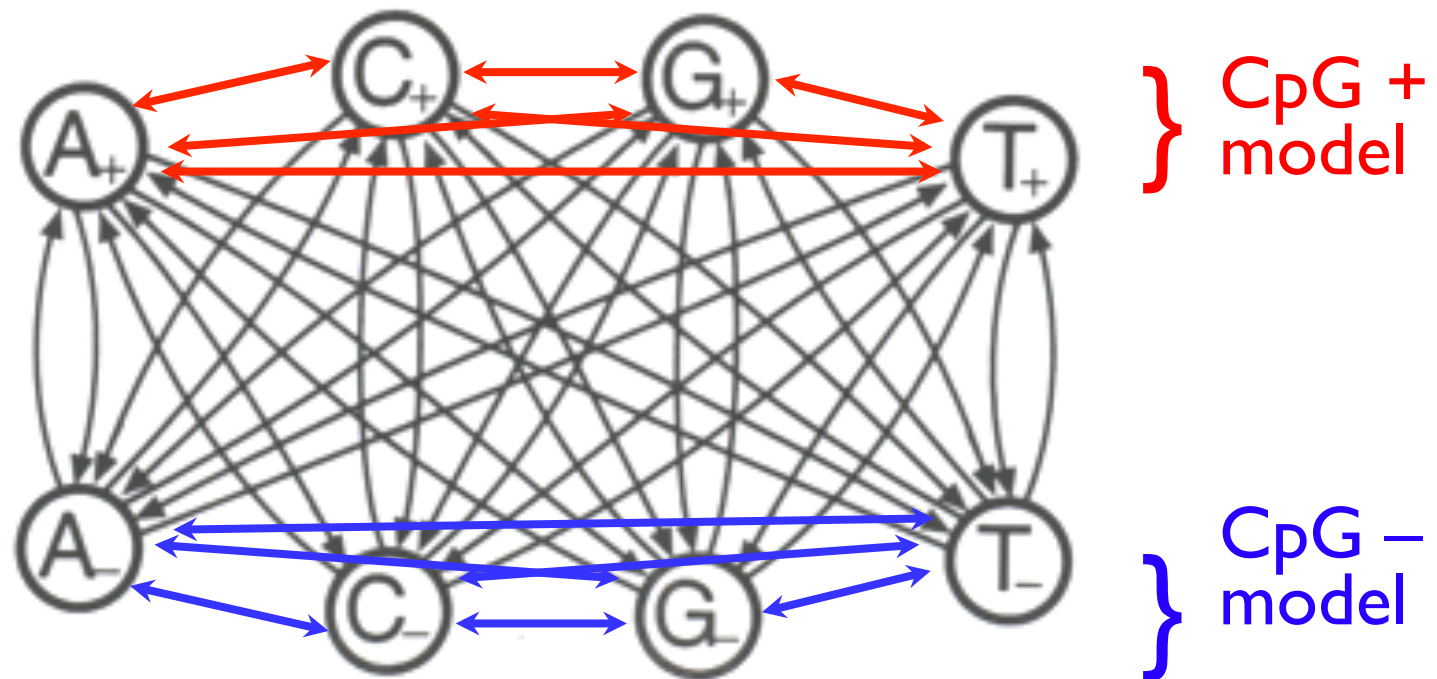
Approach 1: score 100 bp (e.g.) windows

Pro: simple

Con: arbitrary, fixed length, inflexible

Approach 2: combine +/- models.

Combined Model



Emphasis is “Which (hidden) state?” not “Which model?”

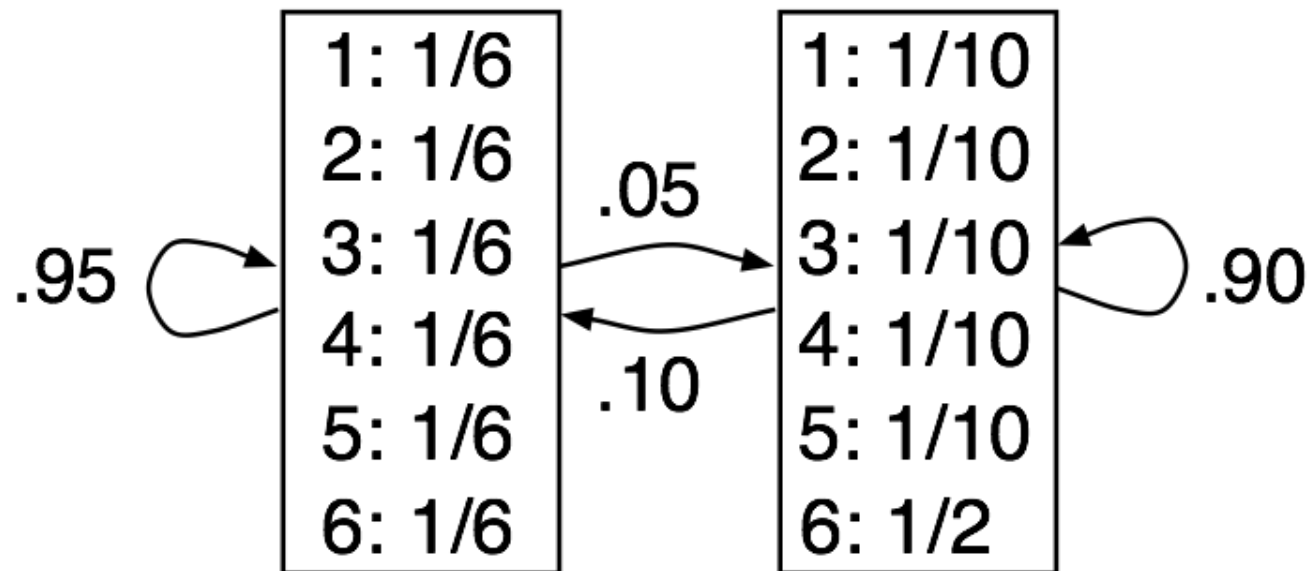
Hidden Markov Models

(HMMs; Claude Shannon, 1948)

States:	$1, 2, 3, \dots$
Paths:	sequences of states $\pi = (\pi_1, \pi_2, \dots)$
Transitions:	$a_{k,l} = P(\pi_i = l \mid \pi_{i-1} = k)$
Emissions:	$e_k(b) = P(x_i = b \mid \pi_i = k)$
Observed data:	emission sequence
Hidden data:	state/transition sequence

The Occasionally Dishonest Casino

1 fair die, 1 “loaded” die, occasionally swapped



Inferring hidden stuff

Joint probability of a given path π & emission sequence x :

$$P(x, \pi) = a_{0, \pi_1} \prod_{i=1}^n e_{\pi_i}(x_i) \cdot a_{\pi_i, \pi_{i+1}}$$

But π is hidden; what to do? Some alternatives:

Most probable single path

$$\pi^* = \arg \max_{\pi} P(x, \pi)$$

Sequence of most probable states

$$\hat{\pi}_i = \arg \max_k P(\pi_i = k \mid x)$$

The Viterbi Algorithm: The most probable path

Viterbi finds: $\pi^* = \arg \max_{\pi} P(x, \pi)$

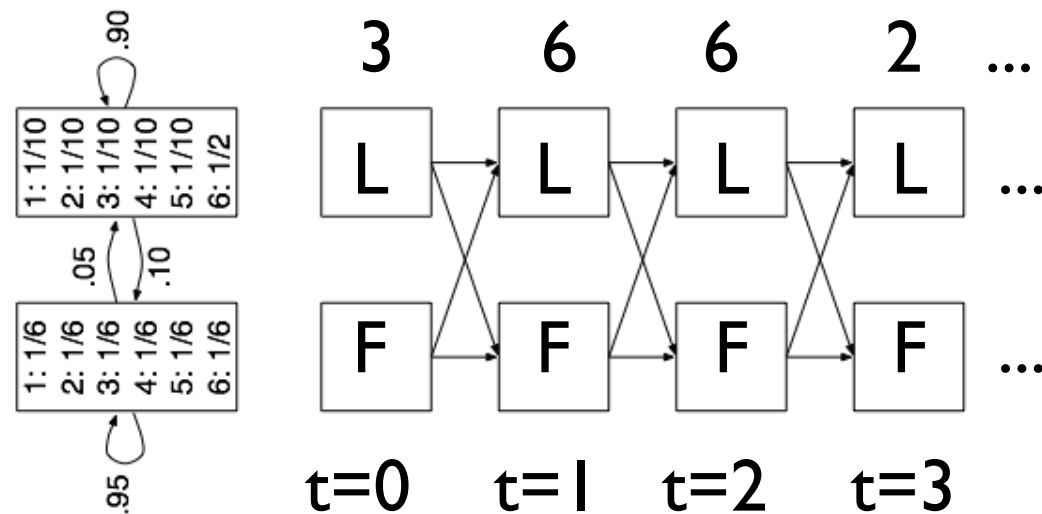
Possibly there are 10^{99} paths of prob 10^{-99}

More commonly, one path (+ slight variants)
dominate others.

(If not, other approaches may be preferable.)

Key problem: exponentially many paths π

Unrolling an HMM



Conceptually, sometimes convenient

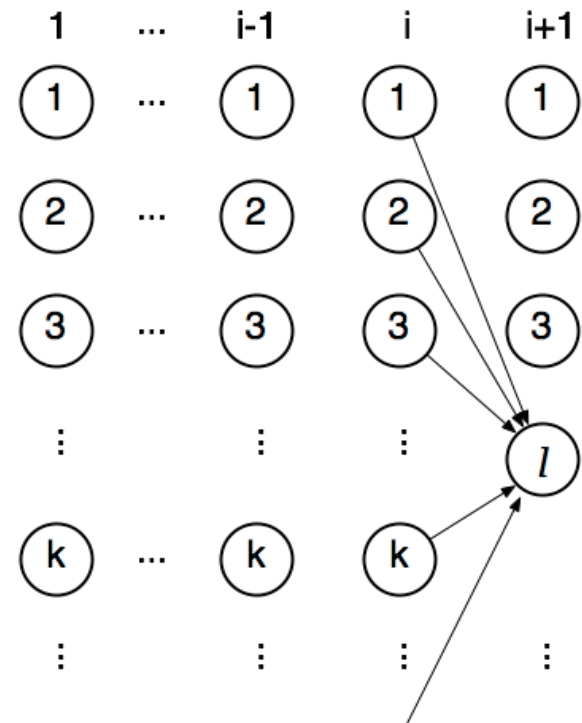
Note exponentially many paths

Viterbi

$v_l(i)$ = probability of the most probable path
emitting x_1, x_2, \dots, x_i and ending in state l

Initialize:

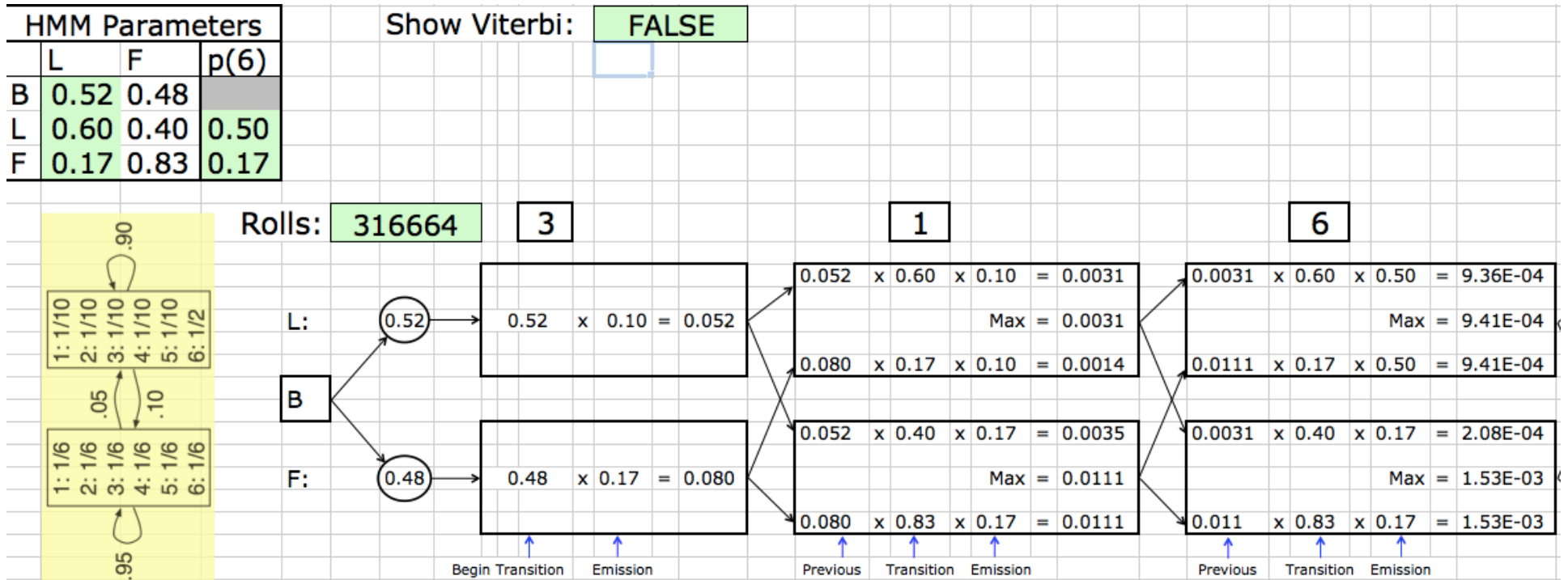
$$v_l(0) = \begin{cases} 1 & \text{if } l = \text{Begin state} \\ 0 & \text{otherwise} \end{cases} \longrightarrow$$



General case:

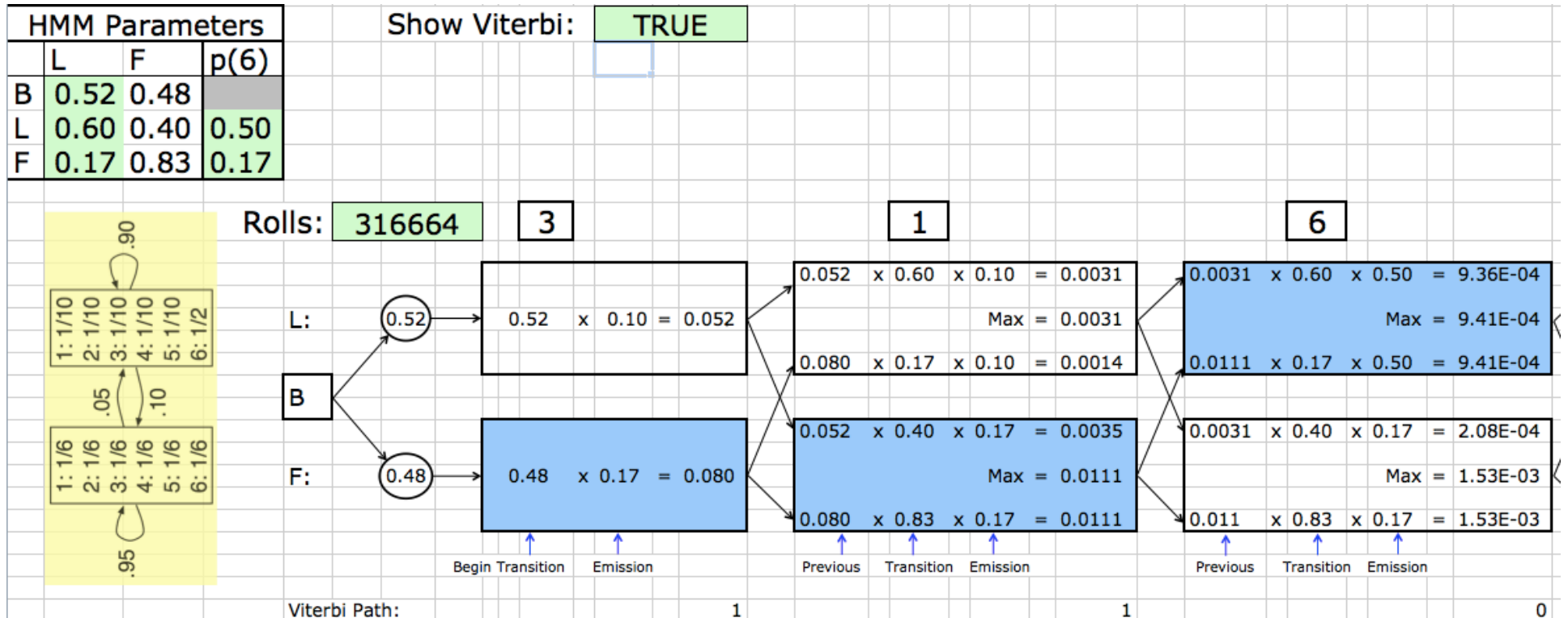
$$v_l(i+1) = e_l(x_{i+1}) \cdot \max_k (v_k(i) a_{k,l})$$

HMM Casino Example



(Excel spreadsheet on web; download & play...)

HMM Casino Example

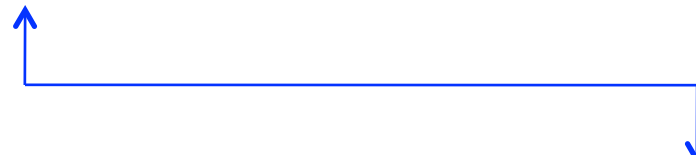


(Excel spreadsheet on web; download & play...)

Viterbi Traceback

Above finds *probability* of best path

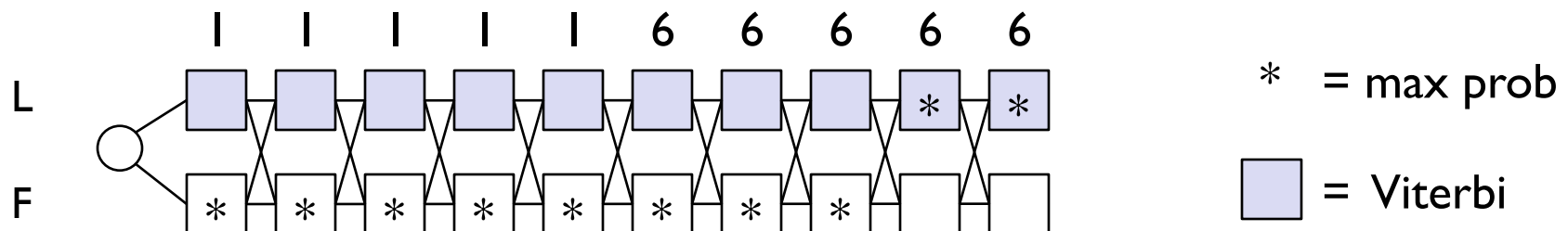
To find the path itself, trace *backward* to the state k attaining the max at each stage


$$v_l(i+1) = e_l(x_{i+1}) \cdot \max_k (v_k(i) a_{k,l})$$

Most probable path \neq Sequence of most probable states

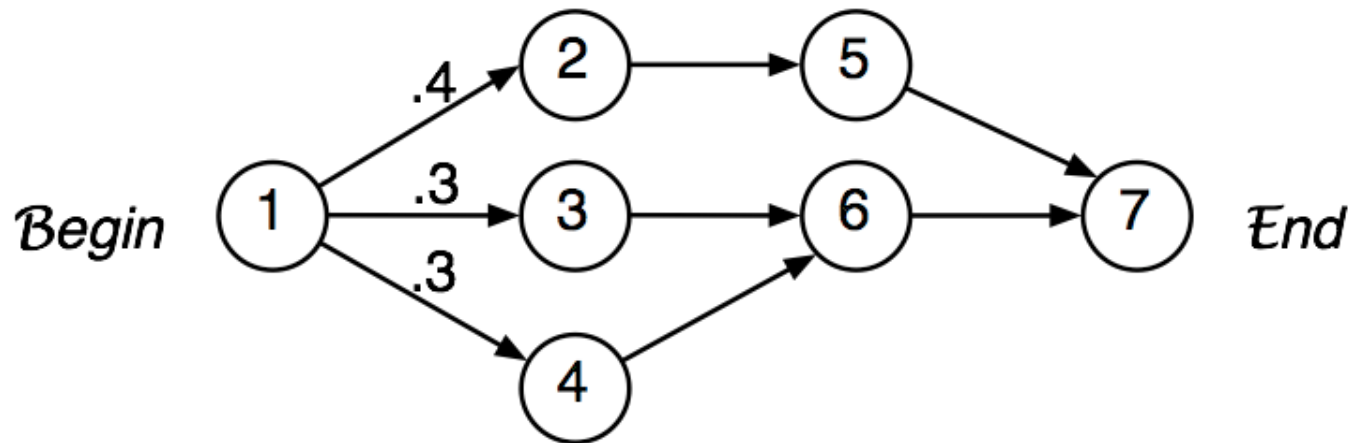
Another example, based on casino dice again

Suppose $p(\text{fair} \leftrightarrow \text{loaded})$ transitions are 10^{-99} and roll sequence is 1 1 1 1 ... 6 6 6 6 6; then fair state is more likely all through 1's & well into the run of 6's, but eventually loaded wins, and the improbable $F \rightarrow L$ transitions make Viterbi = *all* L.



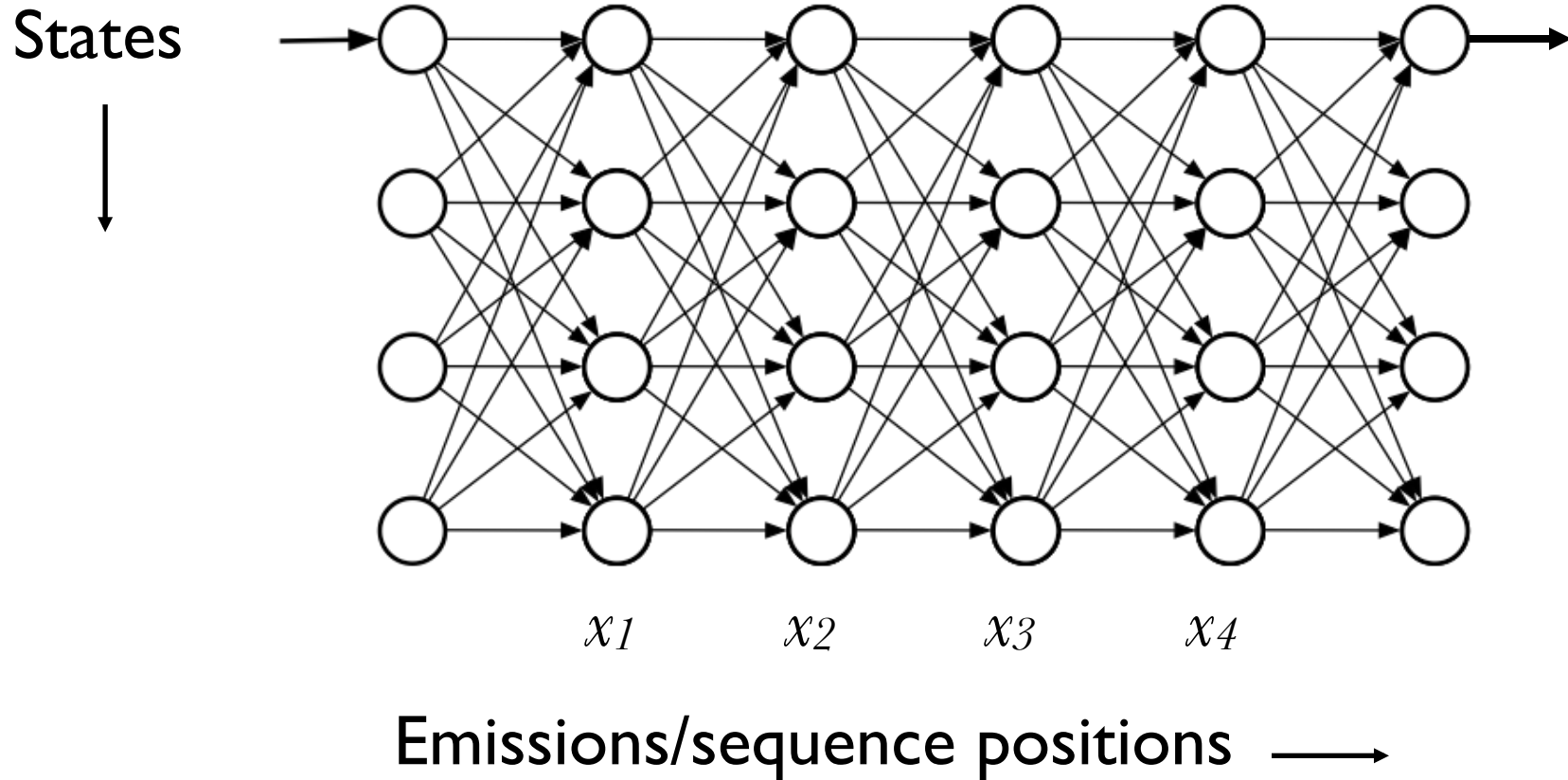
Is Viterbi “best”?

Viterbi finds $\pi^* = \arg \max_{\pi} P(x, \pi)$

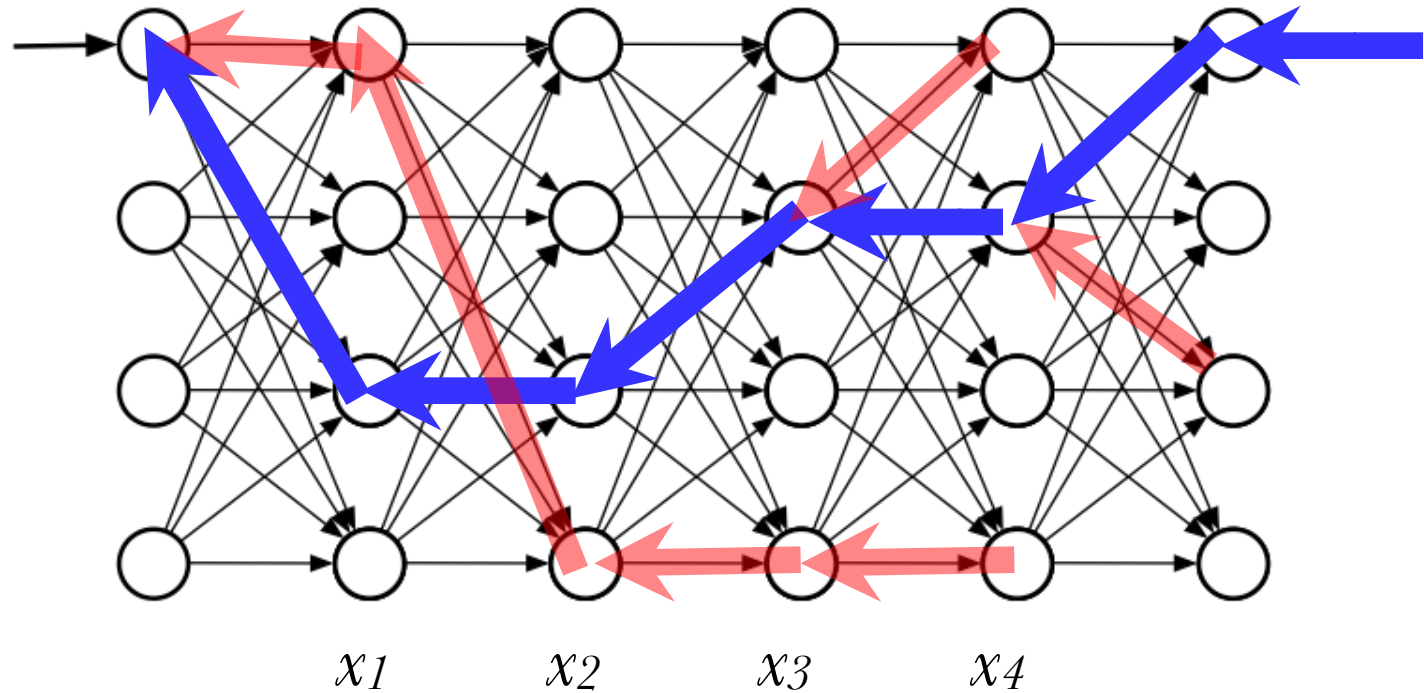


Most probable (Viterbi) *path* goes through 5, but most probable *state* at 2nd step is 6 (I.e., Viterbi is not the only interesting answer.)

An HMM (unrolled)



Viterbi: best path to each state

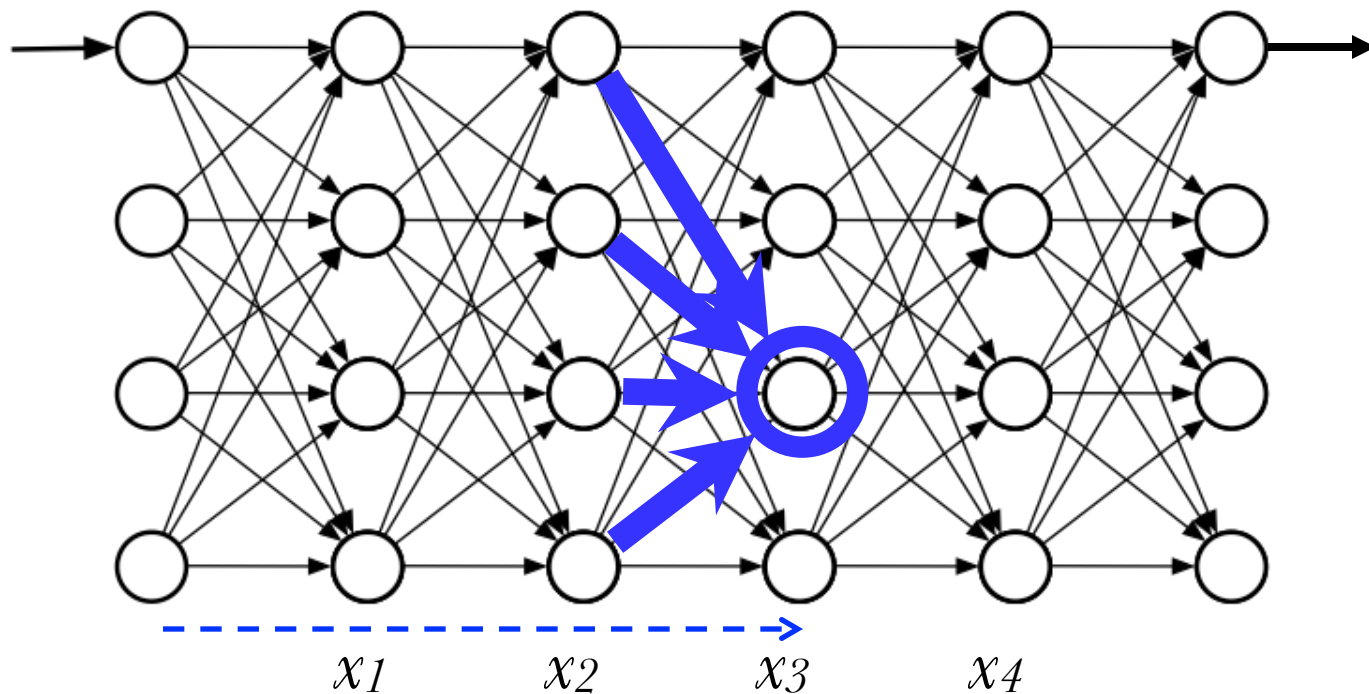


Viterbi score:
$$v_l(i + 1) = e_l(x_{i+1}) \cdot \max_k (v_k(i) a_{k,l})$$

Viterbi path^R:
$$back_l(i + 1) = \arg \max_k (v_k(i) a_{k,l})$$

The Forward Algorithm

For each state/time, want *total* probability of all paths leading to it, with given emissions



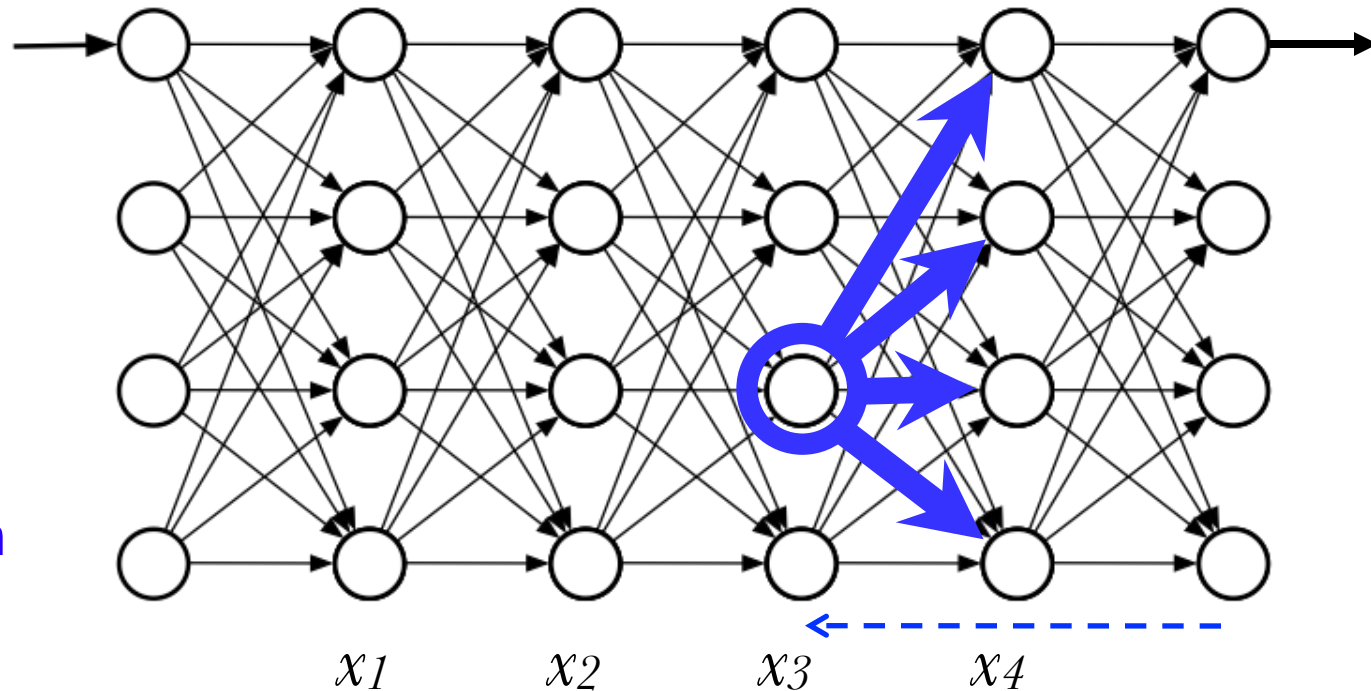
$$f_k(i) \triangleq P(x_1 \dots x_i, \pi_i = k)$$

$$f_l(i+1) = e_l(x_{i+1}) \sum_k f_k(i) a_{k,l}$$

$$P(x) = \sum_{\pi} P(x, \pi) = \sum_k f_k(n) a_{k,0}$$

The Backward Algorithm

Similar:
for each
state/time,
want total
probability
of all paths
from it, with
given
emissions,
conditional
on that
state.



$$b_k(i) \triangleq P(x_{i+1} \cdots x_n \mid \pi_i = k)$$

$$b_k(i) = \sum_l a_{k,l} e_l(x_{i+1}) b_l(i+1)$$

$$b_k(n) = a_{k,0}$$

In state k at step i ?

$$P(x, \pi_i = k)$$

$$= P(x_1, \dots, x_i, \pi_i = k) \cdot P(x_{i+1}, \dots, x_n \mid x_1, \dots, x_i, \pi_i = k)$$

$$= P(x_1, \dots, x_i, \pi_i = k) \cdot P(x_{i+1}, \dots, x_n \mid \pi_i = k)$$

$$= f_k(i) \cdot b_k(i)$$

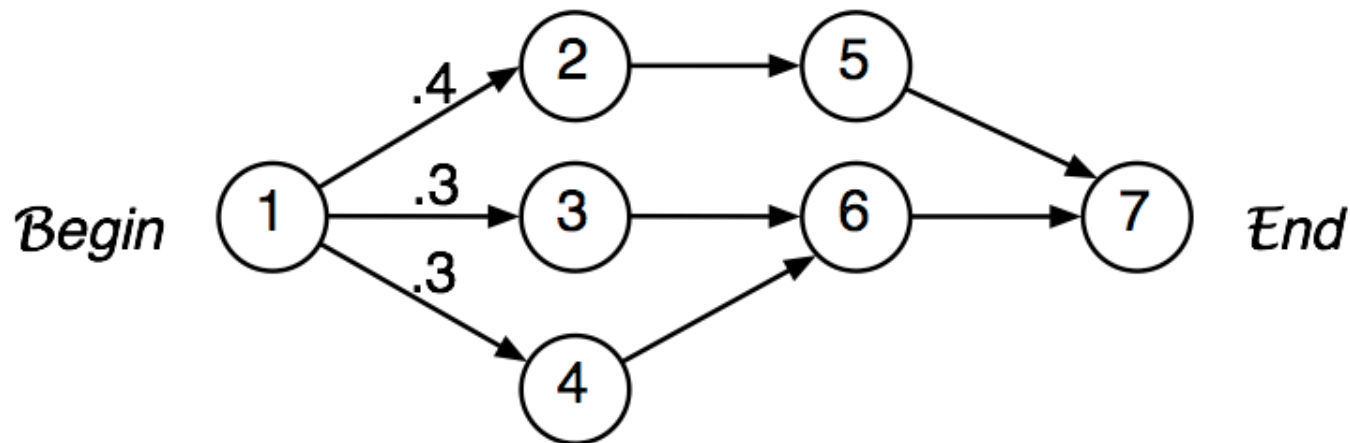
$$P(\pi_i = k \mid x) = \frac{P(x, \pi_i = k)}{P(x)} = \frac{f_k(i) \cdot b_k(i)}{P(x)}$$

Posterior Decoding, I

Alternative 1: what's the most likely state at step i ?

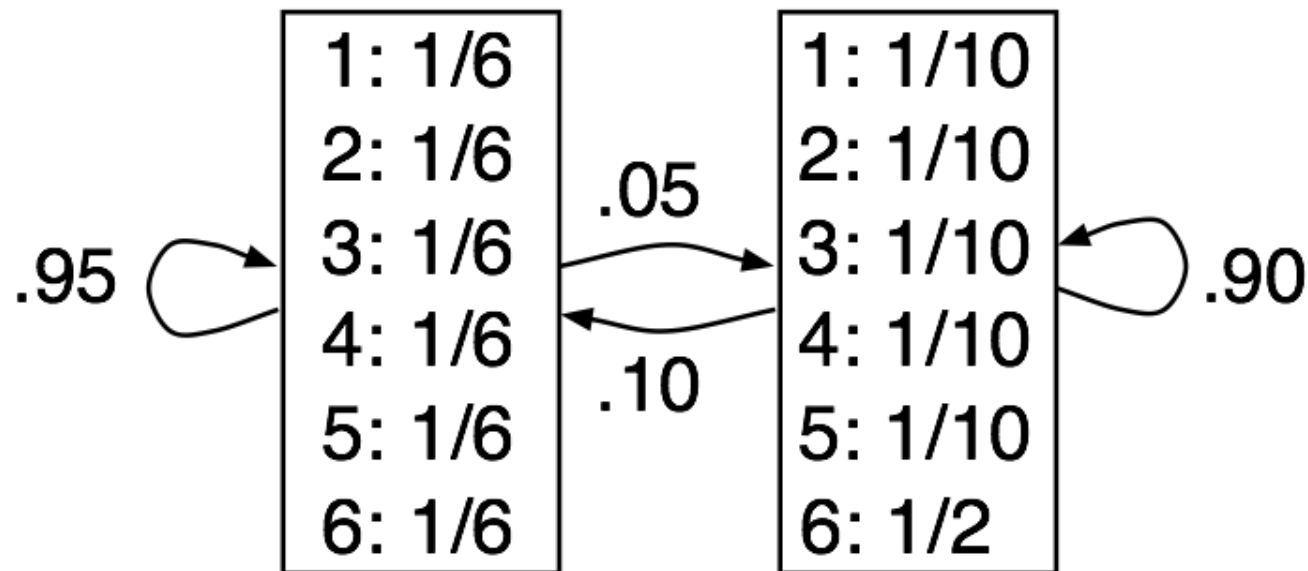
$$\hat{\pi}_i = \arg \max_k P(\pi_i = k \mid x)$$

Note: the sequence of most likely states \neq the most likely sequence of states. May not even be legal!



The Occasionally Dishonest Casino

1 fair die, 1 “loaded” die, occasionally swapped



Posterior Decoding

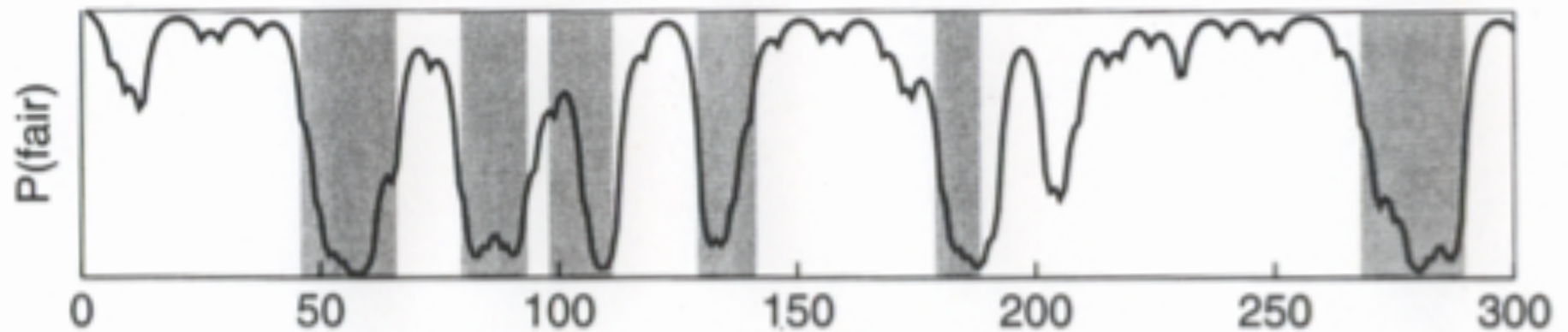


Figure 3.6 *The posterior probability of being in the state corresponding to the fair die in the casino example. The x axis shows the number of the roll. The shaded areas show when the roll was generated by the loaded die.*

Posterior Decoding, II

Alternative 1: what's most likely state at step i ?

$$\hat{\pi}_i = \arg \max_k P(\pi_i = k \mid x)$$

Alternative 2: given some function $g(k)$ on states, what's its expectation. E.g., what's probability of “+” model in CpG HMM ($g(k)=1$ iff k is “+” state)?

$$G(i \mid x) = \sum_k P(\pi_i = k \mid x) \cdot g(k)$$

CpG Islands again

Data: 41 human sequences, totaling 60kbp,
including 48 CpG islands of about 1kbp each

Viterbi:

Found 46 of 48
plus 121 “false positives”

Post-process:

46/48

67 false pos

Posterior Decoding:

same 2 false negatives
plus 236 false positives

46/48

83 false pos

Post-process: merge within
500; discard < 500

Training

Given model topology & training sequences,
learn transition and emission probabilities

If π known, then MLE is just frequency observed
in training data

$$\begin{aligned} a_{k,l} &= \frac{\text{count of } k \rightarrow l \text{ transitions}}{\text{count of } k \rightarrow \text{anywhere transitions}} \\ e_k(b) &= \dots \end{aligned}$$

← + pseudocounts?

If π hidden, then use EM:

given π , estimate θ ; given θ estimate π .

} 2 ways

Viterbi Training

given π , estimate θ ; given θ estimate π

Make initial estimates of parameters θ

Find Viterbi path π for each training sequence

Count transitions/emissions on those paths,
getting new θ

Repeat

Not rigorously optimizing desired likelihood, but
still useful & commonly used.

(Arguably good if you're doing Viterbi decoding.)

Baum-Welch Training

EM: given θ , estimate π ensemble; then re-estimate θ

$$\begin{aligned} P(\pi_i = k, \pi_{i+1} = l \mid x, \theta) \\ = \frac{f_k(i \mid \theta) a_{k,l} e_l(x_{i+1}) b_l(i+1 \mid \theta)}{P(x \mid \theta)} \end{aligned}$$

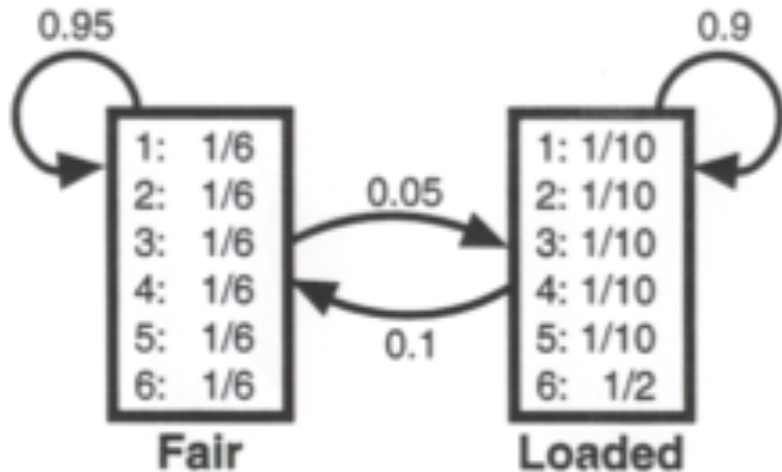
Estimated # of $k \rightarrow l$ transitions $\hat{A}_{k,l}$

$$= \sum_{\text{training seqs } x^j} \sum_i P(\pi_i = k, \pi_{i+1} = l \mid x^j, \theta)$$

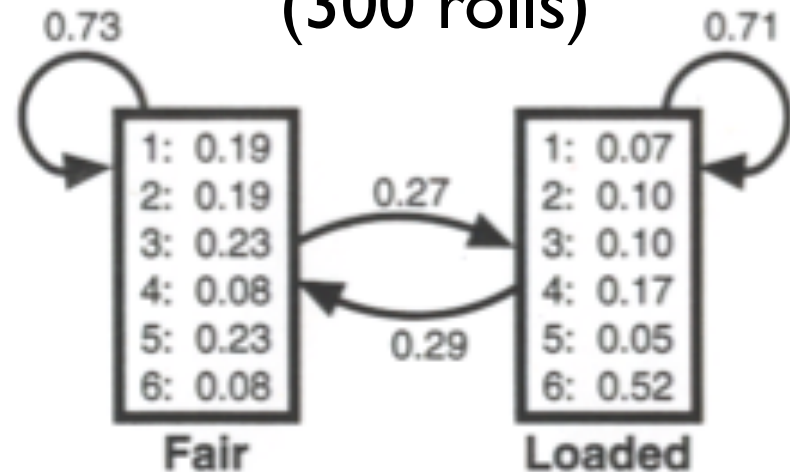
$$\text{New estimate } \hat{a}_{k,l} = \frac{\hat{A}_{k,l}}{\sum_l \hat{A}_{k,l}}$$

Emissions: similar

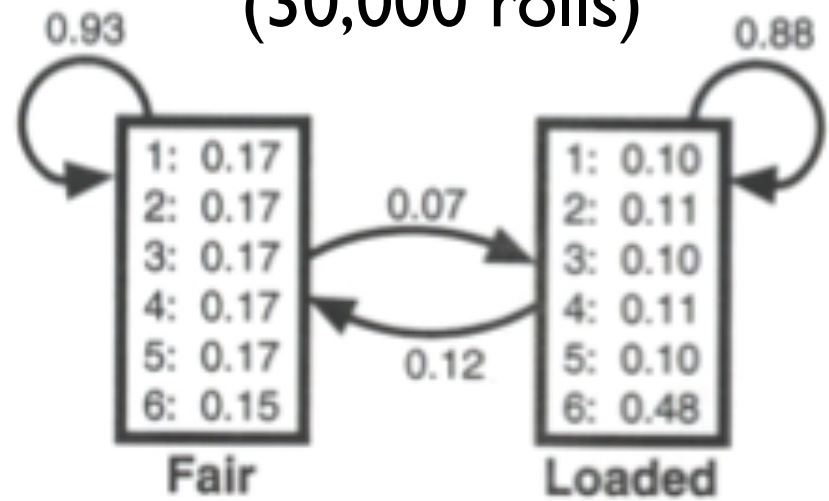
True Model



B-W Learned Model (300 rolls)



B-W Learned Model (30,000 rolls)



Log-odds (vs all F) per roll

True model 0.101 bits

300-roll est. 0.097 bits

30k-roll est. 0.100 bits

(NB: overestimated)

HMMs in Action: Pfam

<http://pfam.sanger.ac.uk/>

Proteins fall into families, both across & within species

Ex: Globins, GPCRs, Zinc fingers, Leucine zippers,...

Identifying family very useful: suggests function, etc.

So, search & alignment are both important

One very successful approach: profile HMMs

```

Helix          AAAAAAAAAAAAAAAAAA  BBBB BBBB BBBB BBBB BBBBBB CCCCCCCCCCCC
HBA_HUMAN      -----VLSPADKTNVKA AWGKVGA--HAGEYGAEALERMF LSFPTTKTYFPHF
HBB_HUMAN      -----VHLTPEEKSAVTALWGKV----NVDEVGGEALGRLLVVYPWTQRFFESF
MYG_PHYCA      -----VLSEGEWQLVLHVWAKVEA--DVAGHGQDILIRLFKSHPETLEKFDRLF
GLB3_CHITP     -----LSADQISTVQASFDKVKG-----DPVGILYAVFKADPSIMAKFTQF
GLB5_PETMA     PIVDTGSVAPLSAAEKT KIRSAWAPVYS--TYETSGVDILVKFFTSTPAAQEFFPKF
LGB2_LUPLU     -----GALTESQAALVKSSWEEFNA--NIPKHTRHFFILVLEIAPA AKDLFS-F
GLB1_GLYDI     -----GLSAAQRQVIAATWKDIAGADNGAGVGKDCLIKFLSAHPQMAAVFG-F
Consensus      Ls.... v a W kv . . g . L.. f . P . F F

```

```

Helix          DDDDDDDDEEEEEEEEEEEEEEEEEEEEEEEEEEE FFFFFFFF FFFF
HBA_HUMAN      -DLS-----HGSAQVKGHGKKVADALTNAVAHV---D--DMPNALSALSDLHA HKL-
HBB_HUMAN      GDLSTPD AVMGNPKVKAHGKKVLGAFSDGLAHL---D--NLKGT FATLSELHCDKL-
MYG_PHYCA      KHLKTEAEMKASEDLKKHGVTVLTALGAILKK---K-GHHEAELKPLAQSHATKH-
GLB3_CHITP     AG-KDLESIKGTAPFETHANRIVGFFSKIIGEL--P---NIEADVNTFVASHKPRG-
GLB5_PETMA     KGLTTADQLKKSADVRWHAERIINAVNDAVASM--DDTEKMSMKLRDL SGKHAKSF-
LGB2_LUPLU     LK-GTSEVPQNNPELQAHAGKVFKLVYEAAIQ LQVTGVVVT DATLKNLGSVHVSKG-
GLB1_GLYDI     SG----AS---DPGVAALGAKVLAQIGVAVSHL--GDEGKMVAQMKAVGV RHKGYGN
Consensus      . t . . . v..Hg kv. a a...l d . a l. l H .

```

```

Helix          FFGGGGGGGGGGGGGGGGGGGGGG HHHHHHHHHHHHHHHHHHHHHHHHHHHHH
HBA_HUMAN      -RVDPVNFKLLSHCLLVTLAAHLPAEFTPAVHASLDKFLASVSTVLT SKYR-----
HBB_HUMAN      -HVDPENFRLLGNVLVCVLAHHFGKEFTPPVQAAYQKVVAGVANALAHKYH-----
MYG_PHYCA      -KIPIKYLEFISEAIIHVLHSRHPGDFGADAQGAMNKALELFRKDIAAKYKELGYQG
GLB3_CHITP     --VTHDQLNNFRAGFVS YMKAHT--DFA-GAEAAWGATLDTFFGMIFSKM-----
GLB5_PETMA     -QVDPQYFKVLA AVIADTVAAG-----DAGFEKLMSMICILLRSAY-----
LGB2_LUPLU     --VADAHFPVVK EAILKTIKEVVGAKWSEELNSAWTIAYDELAIVIKKEMNDAA---
GLB1_GLYDI     KHIKAQYFEPLGASLLS AMEHRIGGKMNA AAKDAWAAAYADISGALISGLQS-----
Consensus      v. f l . . . . . f . aa. k. . l sky

```

Alignment of 7 globins. A-H mark 8 alpha helices.
Consensus line: upper case = 6/7, lower = 4/7, dot=3/7.
Could we have a profile (aka weight matrix) w/ indels?

Profile Hmm Structure

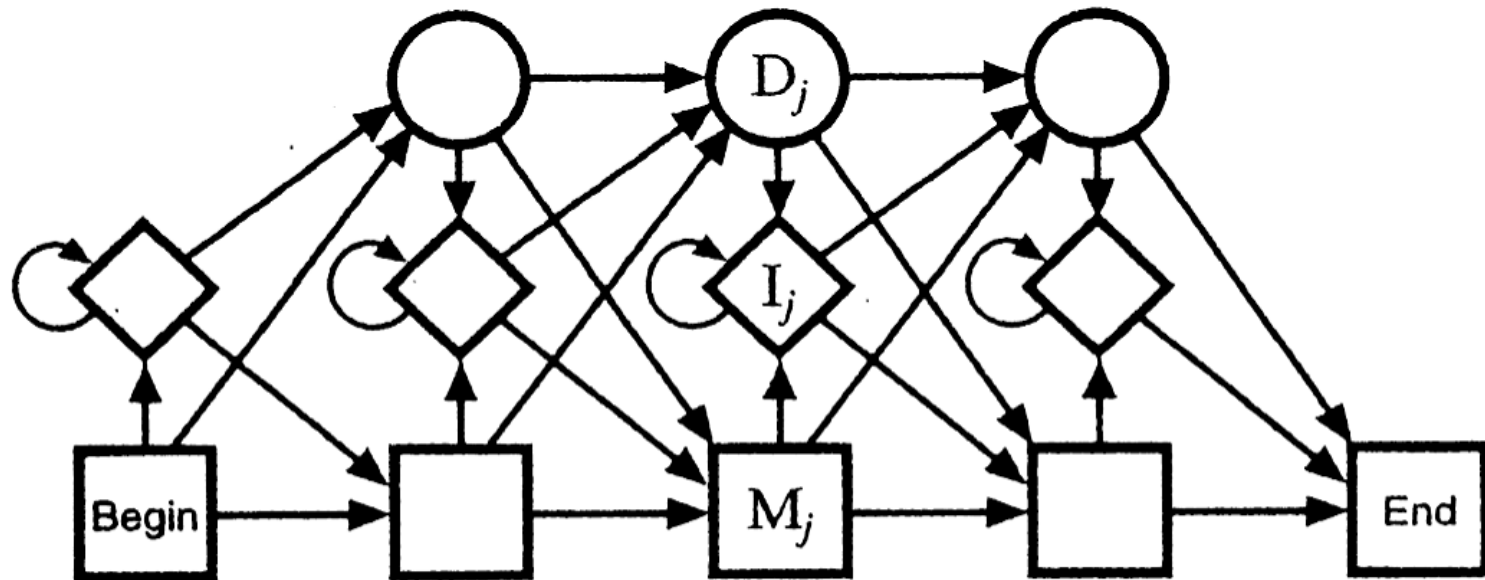


Figure 5.2 *The transition structure of a profile HMM.*

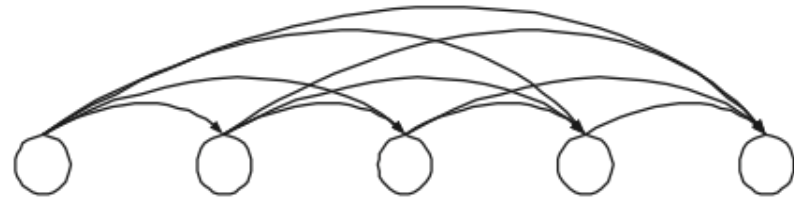
M_j : Match states (20 emission probabilities)

I_j : Insert states (Background emission probabilities)

D_j : Delete states (silent - no emission)

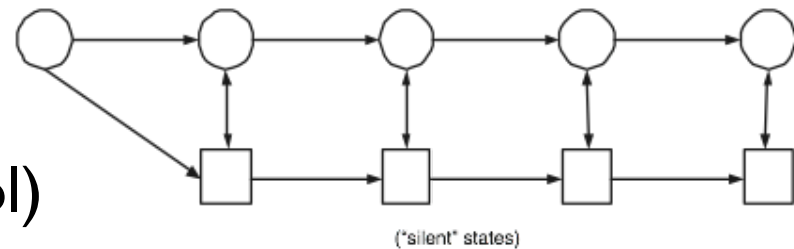
Silent States

Example: chain of states, can skip some



Problem: many parameters.

A solution: chain of “silent” states;
fewer parameters
(but less detailed control)



Algorithms: basically the same.

Using Profile HMM's

Search

Forward or Viterbi

Scoring

Log likelihood (length adjusted)

Log odds vs background

Z scores from either

} next slides

Alignment

Viterbi

Likelihood vs Odds Scores

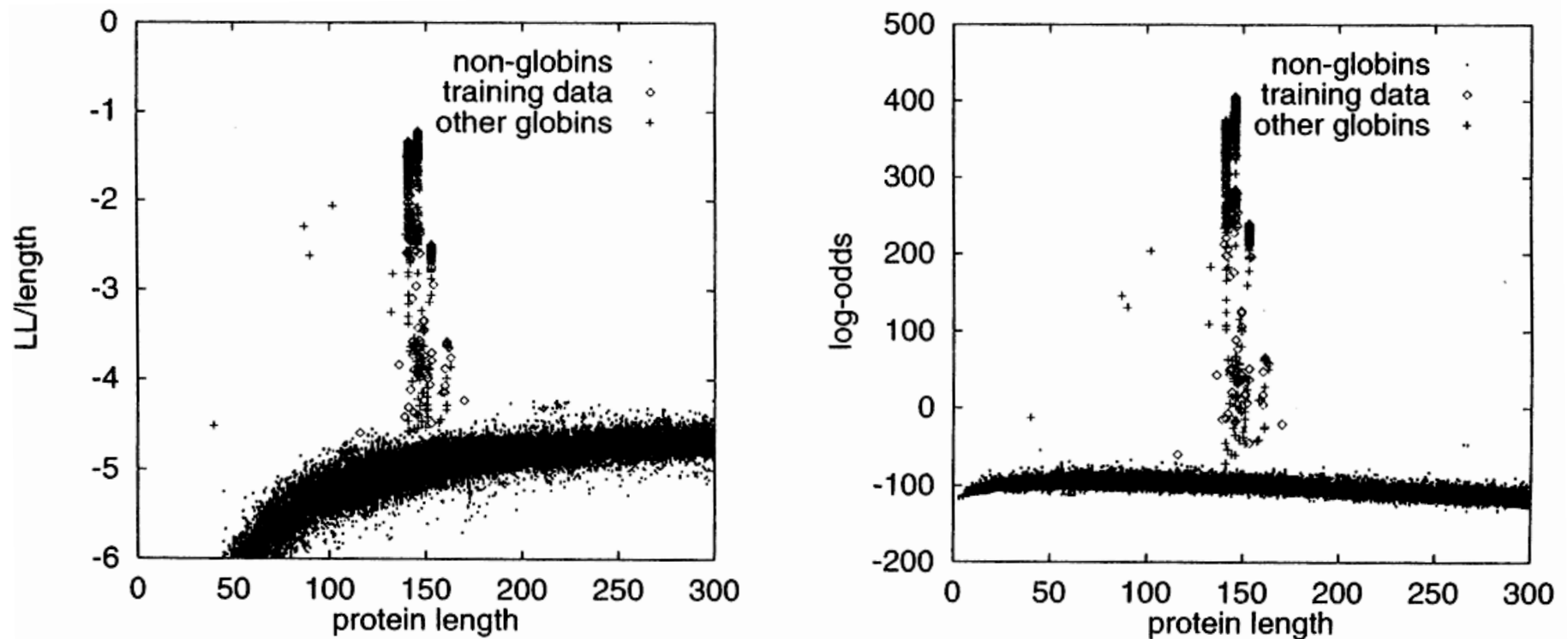


Figure 5.5 To the left the length-normalized LL score is shown as a function of sequence length. The right plot shows the same for the log-odds score.

Z-Scores

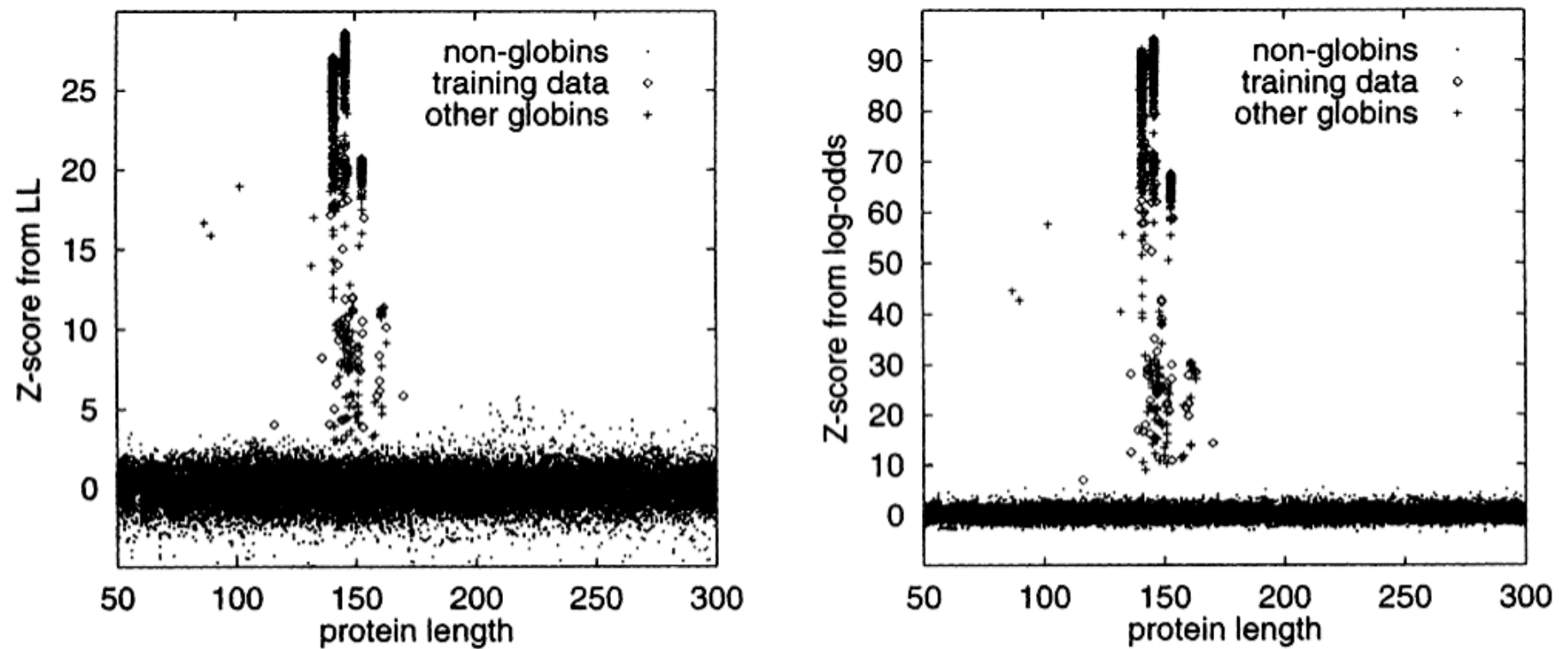


Figure 5.6 *The Z-score calculated from the LL scores (left) and the log-odds (right).*

Pfam Model Building

Hand-curated “seed” multiple alignments

Train profile HMM from seed alignment

Hand-chosen score threshold(s)

Automatic classification/alignment of all other
protein sequences

Pfam 25.0 (March 2011) 12273 families

(covers ~75% of proteins)

refinements

Model-building refinements

Pseudocounts (count = 0 common when training with 20 aa's)

$$e_i(a) = \frac{C_{i,a} + A \cdot q_a}{\sum_a C_{i,a} + A}, \quad A \sim 20, \quad q_a = \text{background}$$

(~50 training sequences)

Pseudocount “mixtures”, e.g. separate pseudocount vectors for various contexts (hydrophobic regions, buried regions,...)

(~10-20 training sequences)

More refinements

Weighting: may need to down weight highly similar sequences to reflect phylogenetic or sampling biases, etc.

Match/insert assignment: Simple threshold, e.g. “> 50% gap \Rightarrow insert”, may be suboptimal.

Can use forward-algorithm-like dynamic programming to compute max *a posteriori* assignment.

refinements

Numerical Issues

Products of many probabilities $\rightarrow 0$

For Viterbi: just add logs

For forward/backward: also work with logs, but
you need sums of products, so need
“log-of-sum-of-product-of-exp-of-logs”,
e.g., by table/interpolation

Keep high precision and perhaps scale factor

Working with log-odds also helps.

refinements

Model structure

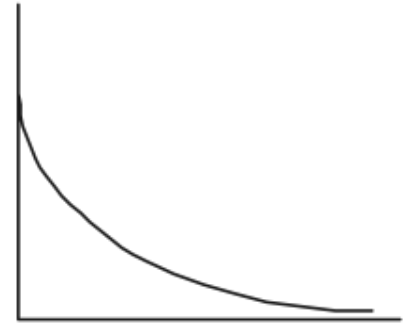
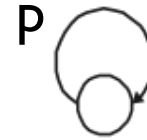
Define it as well as you can.

In principle, you can allow all transitions and hope to learn their probabilities from data, but it usually works poorly – too many local optima

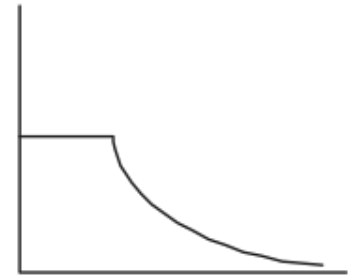
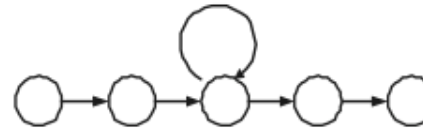
refinements

Duration Modeling

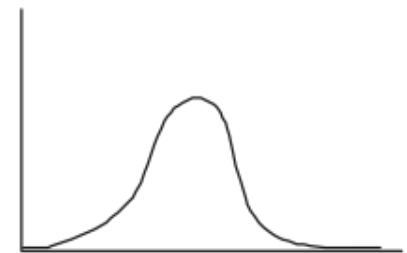
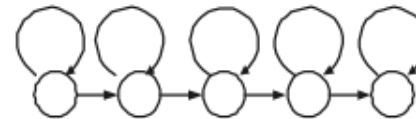
Self-loop duration:
geometric $p^n(1-p)$



min, then geometric



“negative binomial”



More general: possible (but slower)

HMM Summary

joint vs
conditional probs

Inference

- Viterbi – best single path (max of products)
- Forward – sum over all paths (sum of products)
- Backward – similar
- Posterior decoding

Model building

- Semi-supervised – typically fix architecture (e.g. profile HMM), then learn parameters
- Baum-Welch – training via EM and forward/backward (aka the forward/backward algorithm)
- Viterbi training – also “EM”, but Viterbi-based

HMM Summary (cont.)

Search:

Viterbi or forward

Scoring:

Odds ratio to background

Z-score

E-values, etc., too

Excellent tools available (SAM, HMMer, Pfam, ...)

A very widely used tool for biosequence analysis