

CSEP 590A
Summer 2006
Lecture 5
Motifs: Representation & Discovery

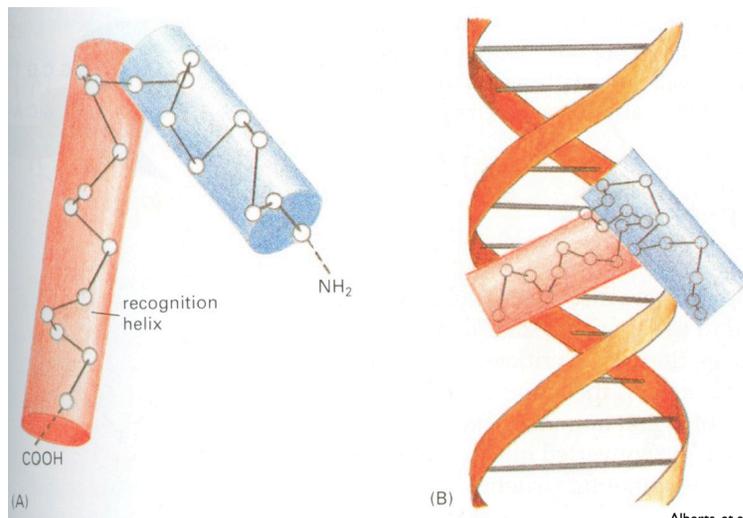
1

DNA Binding Proteins

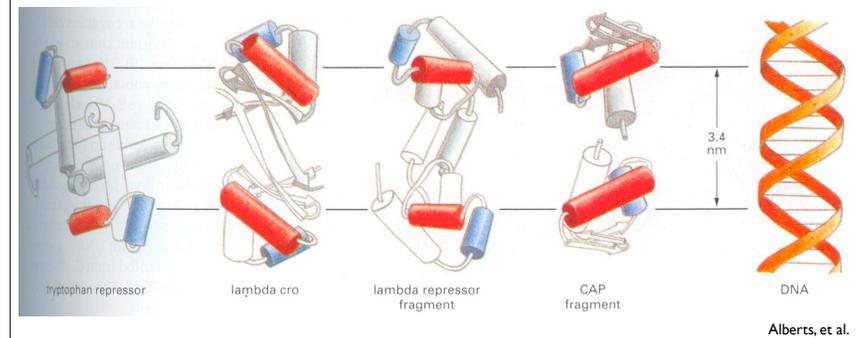
A variety of DNA binding proteins
("transcription factors"; a significant fraction,
perhaps 5-10%, of all human proteins)
modulate transcription of protein coding
genes

2

Helix-Turn-Helix DNA Binding Motif

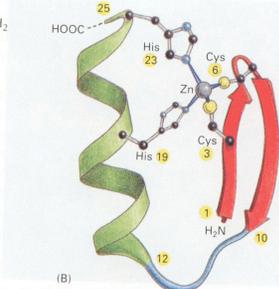
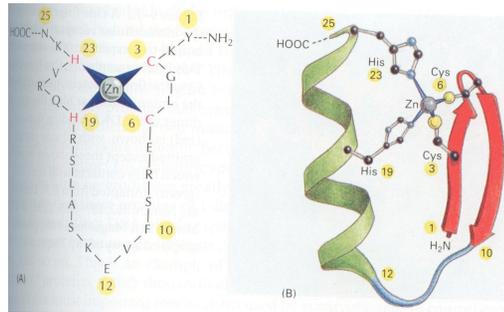


H-T-H Dimers

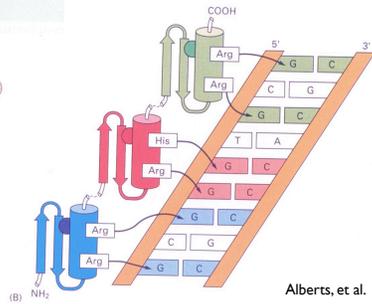
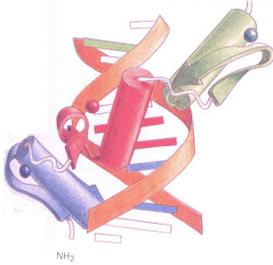


Bind 2 DNA patches, ~ 1 turn apart
Increases both specificity and affinity

4

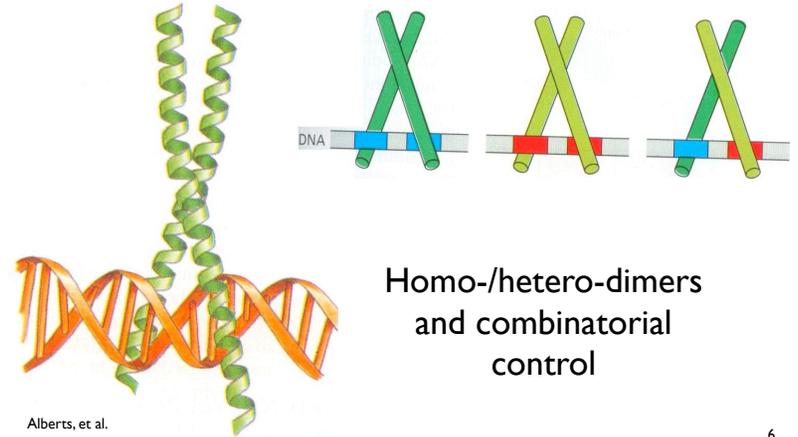


Zinc Finger Motif



Alberts, et al.

Leucine Zipper Motif

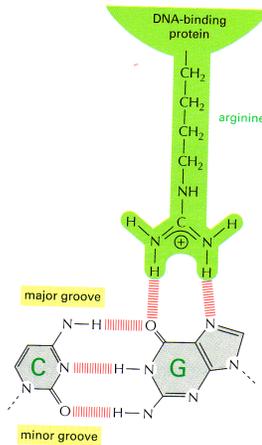
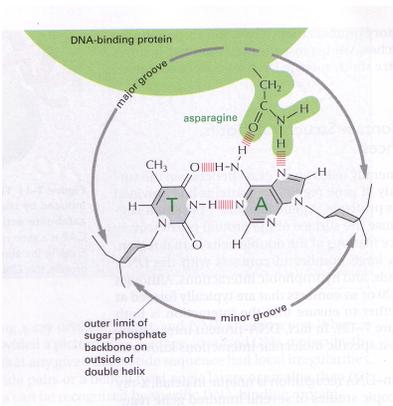


Homo-/hetero-dimers and combinatorial control

Alberts, et al.

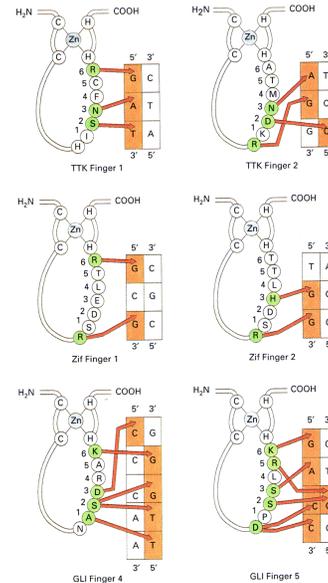
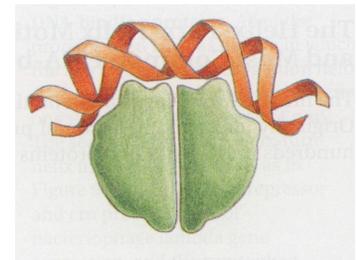
6

Some Protein/DNA interactions well-understood



7

But the overall DNA binding "code" still defies prediction



8

DNA binding site summary

- complex “code”
- short patches (6-8 bp)
- often near each other (1 turn = 10 bp)
- often reverse-complements
- not perfect matches

9

Sequence Motifs

10

E. coli Promoters

- “**TATA Box**” ~ 10bp upstream of transcription start
- How to define it?
 - *Consensus* is TATAAT
 - BUT all differ from it
 - allow k mismatches?
 - equally weighted?
 - wildcards like R,Y? ({A,G}, {C,T}, resp.)

TACGAT
TAAAAT
TATACT
GATAAT
TATGAT
TATGTT

11

E. coli Promoters

- “**TATA Box**” - consensus TATAAT ~ 10bp upstream of transcription start
- *Not exact*: of 168 studied (mid 80’s)
 - nearly all had 2/3 of TAx_zT
 - 80-90% had all 3
 - 50% agreed in each of x,y,z
 - **no** perfect match
- Other common features at -35, etc.

12

TATA Box Frequencies

pos base	1	2	3	4	5	6
A	2	95	26	59	51	1
C	9	2	14	13	20	3
G	10	1	16	15	13	0
T	79	3	44	13	17	96

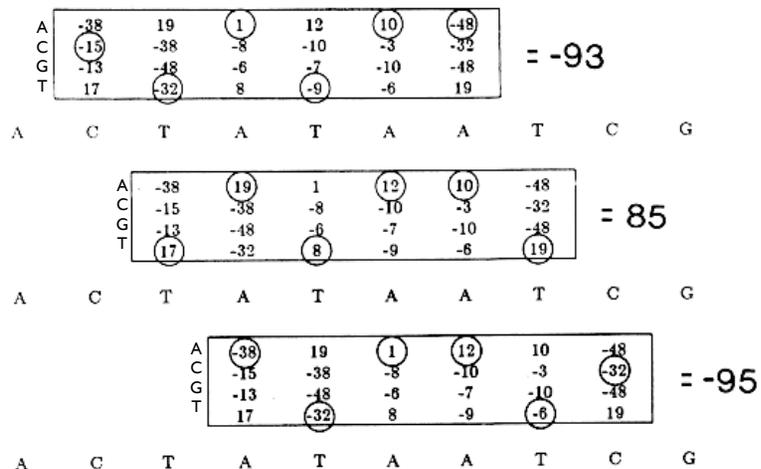
13

TATA Scores

pos base	1	2	3	4	5	6
A	-36	19	1	12	10	-46
C	-15	-36	-8	-9	-3	-31
G	-13	-46	-6	-7	-9	-46(?)
T	17	-31	8	-9	-6	19

14

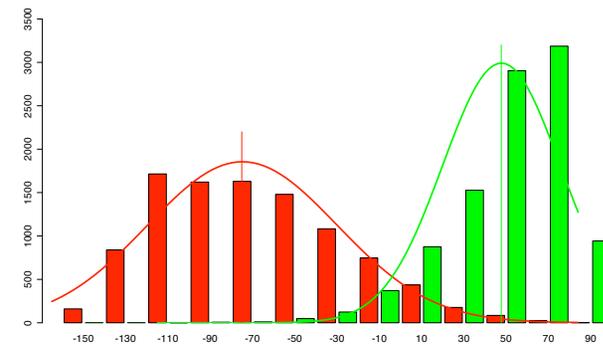
Scanning for TATA



Stormo, Ann. Rev. Biophys. Biophys. Chem, 17, 1988, 241-263

15

Score Distribution (Simulated)



16

Weight Matrices: Statistics

- Assume:

$f_{b,i}$ = frequency of base b in position i in TATA

f_b = frequency of base b in all sequences

- Log likelihood ratio, given $S = B_1 B_2 \dots B_6$:

$$\log \left(\frac{P(S|\text{"tata"})}{P(S|\text{"non-tata"})} \right) = \log \frac{\prod_{i=1}^6 f_{B_i,i}}{\prod_{i=1}^6 f_{B_i}} = \sum_{i=1}^6 \log \frac{f_{B_i,i}}{f_{B_i}}$$

Assumes independence

17

Neyman-Pearson

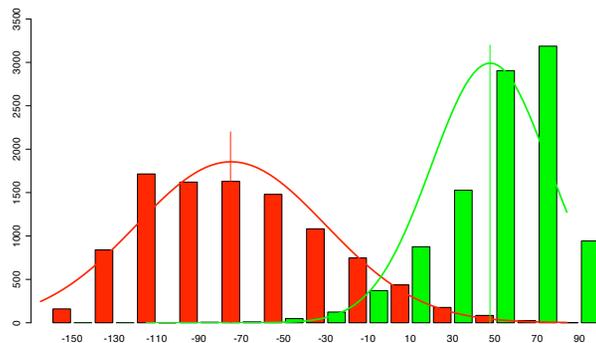
- Given a sample x_1, x_2, \dots, x_n , from a distribution $f(\dots|\Theta)$ with parameter Θ , want to test hypothesis $\Theta = \theta_1$ vs $\Theta = \theta_2$.
- Might as well look at *likelihood ratio*:

$$\frac{f(x_1, x_2, \dots, x_n|\theta_1)}{f(x_1, x_2, \dots, x_n|\theta_2)} > \tau$$

(or log likelihood difference)

18

Score Distribution (Simulated)



19

What's best WMM?

- Given 20 sequences s_1, s_2, \dots, s_k of length 8, assumed to be generated at random according to a WMM defined by $8 \times (4-1)$ parameters θ , what's the best θ ?
- E.g., what's MLE for θ given data s_1, s_2, \dots, s_k ?
- Answer: count frequencies per position.

20

Weight Matrices: Chemistry

- Experiments show ~80% correlation of log likelihood weight matrix scores to measured binding energy of RNA polymerase to variations on TATAAT consensus [Stormo & Fields]

21

Another WMM example

8 Sequences:

ATG
ATG
ATG
ATG
ATG
GTG
GTG
TTG

Freq.	Col 1	Col 2	Col3
A	.625	0	0
C	0	0	0
G	.250	0	1
T	.125	1	0

LLR	Col 1	Col 2	Col 3
A	1.32	$-\infty$	$-\infty$
C	$-\infty$	$-\infty$	$-\infty$
G	0	$-\infty$	2.00
T	-1.00	2.00	$-\infty$

Log-Likelihood Ratio:

$$\log_2 \frac{f_{x_i,i}}{f_{x_i}}, f_{x_i} = \frac{1}{4}$$

22

Non-uniform Background

- E. coli* - DNA approximately 25% A, C, G, T
- M. jannaschi* - 68% A-T, 32% G-C

LLR from previous example, assuming

LLR	Col 1	Col 2	Col 3
A	.74	$-\infty$	$-\infty$
C	$-\infty$	$-\infty$	$-\infty$
G	1.00	$-\infty$	3.00
T	-1.58	1.42	$-\infty$

$$f_A = f_T = 3/8$$

$$f_C = f_G = 1/8$$

e.g., G in col 3 is 8 x more likely via WMM than background, so (\log_2) score = 3 (bits).

23

Relative Entropy

- AKA Kullback-Liebler Distance/Divergence, AKA Information Content
- Given distributions P, Q

$$H(P||Q) = \sum_{x \in \Omega} P(x) \log \frac{P(x)}{Q(x)} \geq 0$$

Notes:

Let $P(x) \log \frac{P(x)}{Q(x)} = 0$ if $P(x) = 0$ [since $\lim_{y \rightarrow 0} y \log y = 0$]

Undefined if $0 = Q(x) < P(x)$

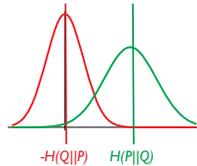
24

WMM: How “Informative”? Mean score of site vs bkg?

- For any fixed length sequence x , let
 $P(x)$ = Prob. of x according to WMM
 $Q(x)$ = Prob. of x according to background

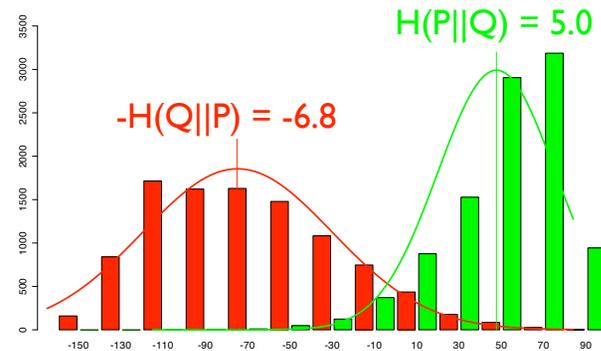
- Relative Entropy:

$$H(P||Q) = \sum_{x \in \Omega} P(x) \log_2 \frac{P(x)}{Q(x)}$$



- $H(P||Q)$ is *expected log likelihood score* of a sequence randomly chosen from **WMM**;
 $-H(Q||P)$ is expected score of **Background**

WMM Scores vs Relative Entropy



For WMM, you can show (based on the assumption of independence between columns), that :

$$H(P||Q) = \sum_i H(P_i||Q_i)$$

where P_i and Q_i are the WMM/background distributions for column i .

WMM Example, cont.

Freq.	Col 1	Col 2	Col 3
A	.625	0	0
C	0	0	0
G	.250	0	1
T	.125	1	0

Uniform				
LLR	Col 1	Col 2	Col 3	
A	1.32	$-\infty$	$-\infty$	
C	$-\infty$	$-\infty$	$-\infty$	
G	0	$-\infty$	2.00	
T	-1.00	2.00	$-\infty$	
RelEnt	.70	2.00	2.00	4.70

Non-uniform				
LLR	Col 1	Col 2	Col 3	
A	.74	$-\infty$	$-\infty$	
C	$-\infty$	$-\infty$	$-\infty$	
G	1.00	$-\infty$	3.00	
T	-1.58	1.42	$-\infty$	
RelEnt	.51	1.42	3.00	4.93

Pseudocounts

- Are the $-\infty$'s a problem?
 - Certain that a given residue *never* occurs in a given position? Then $-\infty$ just right
 - Else, it may be a small-sample artifact
- Typical fix: add a *pseudocount* to each observed count—small constant (e.g., .5, 1)
- Sounds *ad hoc*; there is a Bayesian justification

29

WMM Summary

- Weight Matrix Model (aka Position Specific Scoring Matrix, PSSM, "possum", 0th order Markov models)
- Simple statistical model assuming independence between adjacent positions
- To build: count (+ pseudocount) letter frequency per position, log likelihood ratio to background
- To scan: add LLRs per position, compare to threshold
- Generalizations to higher order models (i.e., letter frequency per position, conditional on neighbor) also possible, with enough training data

30

How-to Questions

- Given aligned motif instances, build model?
 - Frequency counts (above, maybe with pseudocounts)
- Given a model, find (probable) instances
 - Scanning, as above
- Given unaligned strings thought to contain a motif, find it? (e.g., upstream regions for co-expressed genes from a microarray experiment)
 - Hard... rest of lecture.

31

Motif Discovery

Unfortunately, finding a site of max relative entropy in a set of unaligned sequences is NP-hard [Akutsu]

32

Motif Discovery: 3 example approaches

- Greedy search
- Expectation Maximization
- Gibbs sampler

33

Greedy Best-First Approach [Hertz & Stormo]

Input:

- Sequence s_1, s_2, \dots, s_k ; motif length l ; “breadth” d

Algorithm:

- create singleton set with each length l subsequence of each s_1, s_2, \dots, s_k
- for each set, add each possible length l subsequence not already present
- compute relative entropy of each
- discard all but d best
- repeat until all have k sequences

usual “greedy” problems

34

Expectation Maximization [MEME, Bailey & Elkan, 1995]

Input (as above):

- Sequence s_1, s_2, \dots, s_k ; motif length l ; background model; again assume one instance per sequence (variants possible)

Algorithm: EM

- Visible data: the sequences
- Hidden data: where’s the motif

$$Y_{i,j} = \begin{cases} 1 & \text{if motif in sequence } i \text{ begins at position } j \\ 0 & \text{otherwise} \end{cases}$$

- Parameters θ : The WMM

35

MEME Outline

Typical EM algorithm:

- Parameters θ^t at t^{th} iteration, used to estimate where the motif instances are (the hidden variables)
- Use those estimates to re-estimate the parameters θ to maximize likelihood of observed data, giving θ^{t+1}
- Repeat

Key: given a few good matches to best motif, expect to pick out more

36

Expectation Step

(where are the motif instances?)

$$\begin{aligned}
 \hat{Y}_{i,j} &= E(Y_{i,j} | s_i, \theta^t) \xrightarrow{E = 0 \cdot P(0) + 1 \cdot P(1)} \\
 &= P(Y_{i,j} = 1 | s_i, \theta^t) \xrightarrow{\text{Bayes}} \\
 &= P(s_i | Y_{i,j} = 1, \theta^t) \frac{P(Y_{i,j}=1|\theta^t)}{P(s_i|\theta^t)} \\
 &= cP(s_i | Y_{i,j} = 1, \theta^t) \\
 &= c' \prod_{k=1}^l P(s_{i,j+k-1} | \theta^t)
 \end{aligned}$$

where c' is chosen so that $\sum_j \hat{Y}_{i,j} = 1$.

Maximization Step

(what is the motif?)

Find θ maximizing expected value:

$$\begin{aligned}
 Q(\theta | \theta^t) &= E_{Y \sim \theta^t} [\log P(s, Y | \theta)] \\
 &= E_{Y \sim \theta^t} [\log \prod_{i=1}^k P(s_i, Y_i | \theta)] \\
 &= E_{Y \sim \theta^t} [\sum_{i=1}^k \log P(s_i, Y_i | \theta)] \\
 &= E_{Y \sim \theta^t} [\sum_{i=1}^k \sum_{j=1}^{|s_i|-l+1} Y_{i,j} \log P(s_i, Y_{i,j} = 1 | \theta)] \\
 &= E_{Y \sim \theta^t} [\sum_{i=1}^k \sum_{j=1}^{|s_i|-l+1} Y_{i,j} \log (P(s_i | Y_{i,j} = 1, \theta) P(Y_{i,j} = 1 | \theta))] \\
 &= \sum_{i=1}^k \sum_{j=1}^{|s_i|-l+1} E_{Y \sim \theta^t} [Y_{i,j}] \log P(s_i | Y_{i,j} = 1, \theta) + C \\
 &= \sum_{i=1}^k \sum_{j=1}^{|s_i|-l+1} \hat{Y}_{i,j} \log P(s_i | Y_{i,j} = 1, \theta) + C
 \end{aligned}$$

M-Step (cont.)

$$Q(\theta | \theta^t) = \sum_{i=1}^k \sum_{j=1}^{|s_i|-l+1} \hat{Y}_{i,j} \log P(s_i | Y_{i,j} = 1, \theta) + C$$

Exercise: Show this is maximized by “counting” letter frequencies over all possible motif instances, with counts weighted by $\hat{Y}_{i,j}$, again the “obvious” thing.

s_1 : ACGGATT...
 ...
 s_k : GC...TCGGAC

 $\hat{Y}_{1,1}$ ACGG
 $\hat{Y}_{1,2}$ CGGA
 $\hat{Y}_{1,3}$ GGAT
 ⋮ ⋮
 $\hat{Y}_{k,l-1}$ CGGA
 $\hat{Y}_{k,l}$ GGAC

Initialization

1. Try every motif-length substring, and use as initial θ a WMM with, say 80% of weight on that sequence, rest uniform
2. Run a few iterations of each
3. Run best few to convergence
(Having a supercomputer helps)

Another Motif Discovery Approach The Gibbs Sampler

Lawrence, et al. "Detecting Subtle Sequence Signals: A Gibbs Sampling Strategy for Multiple Sequence Alignment," *Science* 1993

Sigma-37	223	IIDLTYIQNK	SQKETGDILGISQMHVSR	LQRKAVKKLR	240	A25944
SpoIIIC	94	RFGLDLKKEK	TQREIAKELGISRSYVSR	IEKRALMKMF	111	A28627
NahR	22	VVFNQLLVDR	RVSITAENLGLTQPAVSN	ALKRLRTSLQ	39	A32837
Antennapedia	326	FHFNRYLTRR	RRIEIAHALCLTERQIKI	WFQNRMRWK	343	A23450
NtrC (Brady.)	449	LTAALAAATRG	NQIRAADLLGLNRNLTLR	KIRDLDIQVY	466	B26499
DicA	22	IRYRRKNLKH	TQRLAKALKISHVSVSQ	WERGDSEPTG	39	B24328 (BVECD)
MerD	5	MNAY	TVSRLALDAGVSVHIVRD	YLLRGLLRPV	22	C29010
Fis	73	LDMVMQYTRG	NQTRAAALMMGINRGTLRK	KLKKYGMN	90	A32142 (DNECF)
MAT a1	99	FRKQSLNSK	EKEEVAKCKGITPLQVRV	WFINKRMRSK	116	A90983 (JEBY1)
Lambda cII	25	SALLNKIAML	GTEKTAEAVGVDSQISR	WKRDWIPKFS	42	A03579 (QCBP2L)
Crp (CAP)	169	THPDGMQIKI	TRQEIGQIVGCSRETVGR	ILKMLEDQNL	186	A03553 (QRECC)
Lambda Cro	15	ITLKYAMRF	GQTKTAKDLGVYQSAINK	AIHAGRKIFL	32	A03577 (RCPBL)
P22 Cro	12	YKDVIDHFG	TQRAVAKALGISDAAVSQ	WKEVIPEKDA	29	A25867 (RGBP22)
AraC	196	ISDHLADSNF	DIASVAQHVCLSPRSLSH	LFRQQLGISV	213	A03554 (RGCEA)
Fnr	196	FSPREFRLTM	TRGDIGNYLGLTVETISR	LLGRFQKSGM	213	A03552 (RGECF)
HtpR	252	ARWLDEDNKS	TLQELADRYGVAERVRQ	LEKNAMKKLR	269	A00700 (RGEC)
NtrC (K.a.)	444	LTALRHTQG	HKQEAARLGLWGRNLTTR	KLKELGME	461	A03564 (RGKBCP)
CytR	11	MKAKQETA	TMKDVALKAKVSTATVSR	ALMNPDKVSQ	28	A24963 (RPECCT)
DeoR	23	LQELKRSDKL	HLKDAALGLVSEMTIRR	DLNNSAPVV	40	A24076 (RPECDO)
GalR	3	MA	TIKDVARLAGVSVATVSR	VINNSPKASE	20	A03559 (RPEC)
LacI	5	MKPV	TLYDVAEYAGVSYQTVSR	VVNQASHVSA	22	A03558 (RPECL)
TetR	26	LLNEVEIGEL	TTRKLAQKLGVEQPTLYW	HVNKRALLD	43	A03576 (RPECTN)
TrpR	67	IVEELLRGEM	SQRELKNELGAGIATITR	GSNSLKAAPV	84	A03568 (RPECW)
NifA	495	LIAALEKAGW	VQAKAARLLGMTPRQVAY	RIQIMDIIMP	512	S02513
SpoIIG	205	RFGLVGEEK	TQKDVADMMGISQSYISR	LEKRIIKRLR	222	S07337
Pin	160	QAGRLIAAGT	PRQKVAIIYDVGVSITLYK	TFPAGDK	177	S07958
PurR	3	MA	TIKDVAKRANVSTTVSH	VINKTRFVAE	20	S08477
EbgR	3	MA	TLKDIAIEAGVSLATVSR	VLNDDPTLVN	20	S09205
LexA	27	DHISQTMGPP	TRAEIAQRLGFRSPNAAE	EHLKALARKG	44	S11945
P22 cI	25	SSILNRIR	GQRKVADALGINESQISR	WKGDFIPKMG	42	B25867 (Z1BPC2)
			*****	***		

B	Position in site																	
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18
Arg	94	222	265	137	9	9	137	137	9	9	9	52	222	94	94	9	265	606
Lys	9	133	442	380	9	71	380	194	9	133	9	9	71	9	9	9	71	256
Glu	53	9	96	401	9	9	140	140	9	9	9	53	140	140	9	9	9	53
Asp	67	9	9	473	9	9	224	125	9	67	9	67	67	9	9	9	9	67
Gln	9	600	224	9	9	9	224	9	9	9	9	9	278	63	278	9	9	170
His	240	9	9	9	9	9	125	125	9	9	9	9	125	125	125	9	9	240
Asn	168	9	9	9	9	9	168	89	9	89	9	248	9	168	89	9	89	89
Ser	117	9	117	117	9	9	9	9	9	9	9	819	63	387	63	9	819	9
Gly	151	9	56	9	9	151	9	9	9	1141	9	151	9	56	9	9	56	9
Ala	9	9	112	43	181	901	43	181	215	9	43	9	43	181	112	43	78	9
Thr	915	130	130	9	251	9	9	9	9	9	9	311	130	70	855	9	130	9
Pro	76	9	9	9	9	9	9	9	9	9	9	9	210	210	9	9	9	9
Cys	9	9	9	9	9	9	9	9	295	581	295	9	9	9	9	9	9	9
Val	58	107	9	9	500	9	9	9	156	9	598	9	205	58	9	746	9	58
Leu	9	121	9	9	149	9	93	149	458	9	149	9	37	37	9	177	9	9
Ile	9	166	114	61	323	9	114	166	9	9	427	9	61	9	61	427	9	61
Met	9	104	9	9	9	9	9	198	198	9	104	9	9	198	9	9	9	9
Tyr	9	9	136	9	9	9	9	262	262	9	9	136	136	9	262	9	262	136
Phe	9	9	9	9	9	9	9	9	9	9	108	9	9	9	9	9	9	9
Trp	9	9	9	9	9	9	9	9	9	9	366	9	9	9	9	9	9	366

Some History

- Geman & Geman, IEEE PAMI 1984
- Hastings, Biometrika, 1970
- Metropolis, Rosenbluth, Rosenbluth, Teller, & Teller, "Equations of State Calculations by Fast Computing Machines," *J. Chem. Phys.* 1953
- Josiah Williard Gibbs, 1839-1903, American physicist, a pioneer of thermodynamics

How to Average

An old problem:

- n random variables: x_1, x_2, \dots, x_k
- Joint distribution (p.d.f.): $P(x_1, x_2, \dots, x_k)$
- Some function: $f(x_1, x_2, \dots, x_k)$
- Want Expected Value: $E(f(x_1, x_2, \dots, x_k))$

How to Average

$$E(f(x_1, x_2, \dots, x_k)) = \int_{x_1} \int_{x_2} \dots \int_{x_k} f(x_1, x_2, \dots, x_k) \cdot P(x_1, x_2, \dots, x_k) dx_1 dx_2 \dots dx_k$$

- Approach 1: direct integration (rarely solvable analytically, esp. in high dim)
- Approach 2: numerical integration (often difficult, e.g., unstable, esp. in high dim)
- Approach 3: Monte Carlo integration
sample $\vec{x}^{(1)}, \vec{x}^{(2)}, \dots, \vec{x}^{(n)} \sim P(\vec{x})$ and average:

$$E(f(\vec{x})) \approx \frac{1}{n} \sum_{i=1}^n f(\vec{x}^{(i)})$$

Markov Chain Monte Carlo (MCMC)

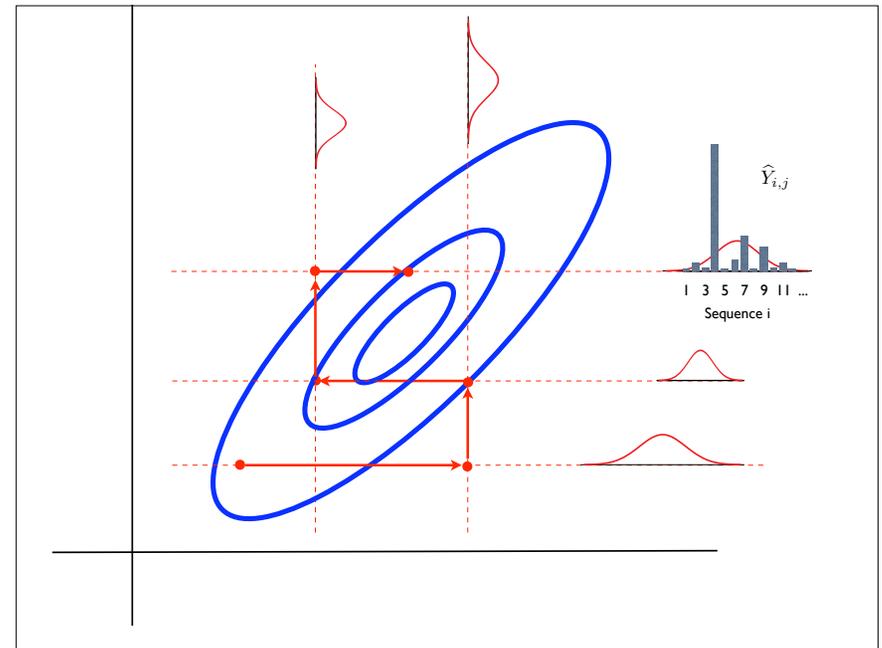
- Independent sampling also often hard, but *not* required for expectation
- MCMC $\vec{X}_{t+1} \sim P(\vec{X}_{t+1} | \vec{X}_t)$ w/ stationary dist = P
- Simplest & most common: Gibbs Sampling
 $P(x_i | x_1, x_2, \dots, x_{i-1}, x_{i+1}, \dots, x_k)$

Algorithm

for t = 1 to ∞

for i = 1 to k do :

$$x_{t+1,i} \sim P(x_{t+1,i} | \underbrace{x_{t+1,1}, x_{t+1,2}, \dots, x_{t+1,i-1}}_{t+1}, \underbrace{x_{t,i+1}, \dots, x_{t,k}}_t)$$



- Input: again assume sequences s_1, s_2, \dots, s_k with one length w motif per sequence
- Motif model: WMM
- Parameters: Where are the motifs?
for $1 \leq i \leq k$, have $1 \leq x_i \leq |s_i| - w + 1$
- “Full conditional”: to calc

$$P(x_i = j \mid x_1, x_2, \dots, x_{i-1}, x_{i+1}, \dots, x_k)$$
 build WMM from motifs in all sequences except i , then calc prob that motif in i^{th} seq occurs at j by usual “scanning” alg.

Overall Gibbs Alg

Randomly initialize x_i 's

for $t = 1$ to ∞

 for $i = 1$ to k

 discard motif instance from s_i ;

 recalc WMM from rest

 for $j = 1 \dots |s_i| - w + 1$

 calculate prob that i^{th} motif is at j :

Similar to MEME, but it would average over, rather than sample from $\longrightarrow P(x_i = j \mid x_1, x_2, \dots, x_{i-1}, x_{i+1}, \dots, x_k)$

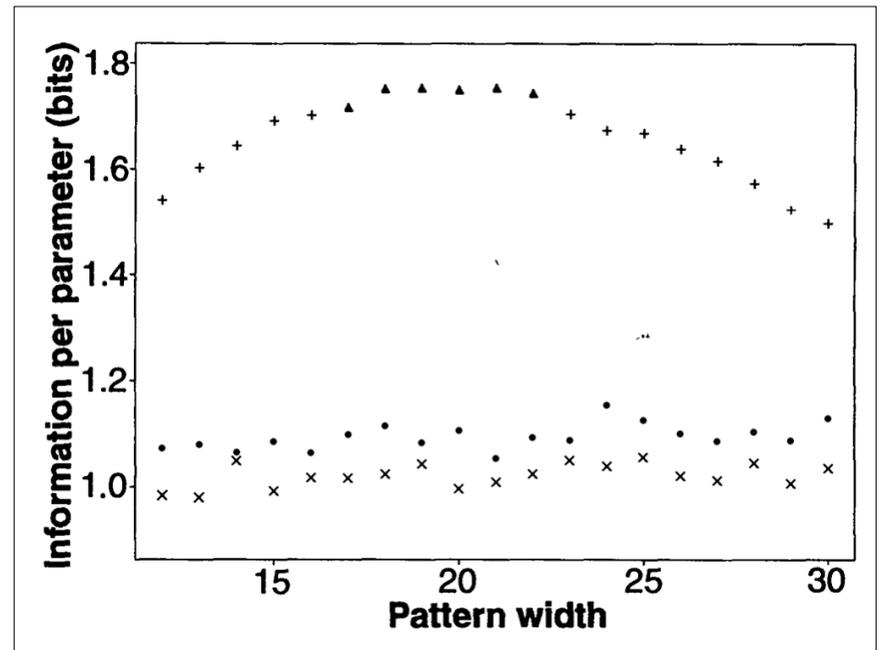
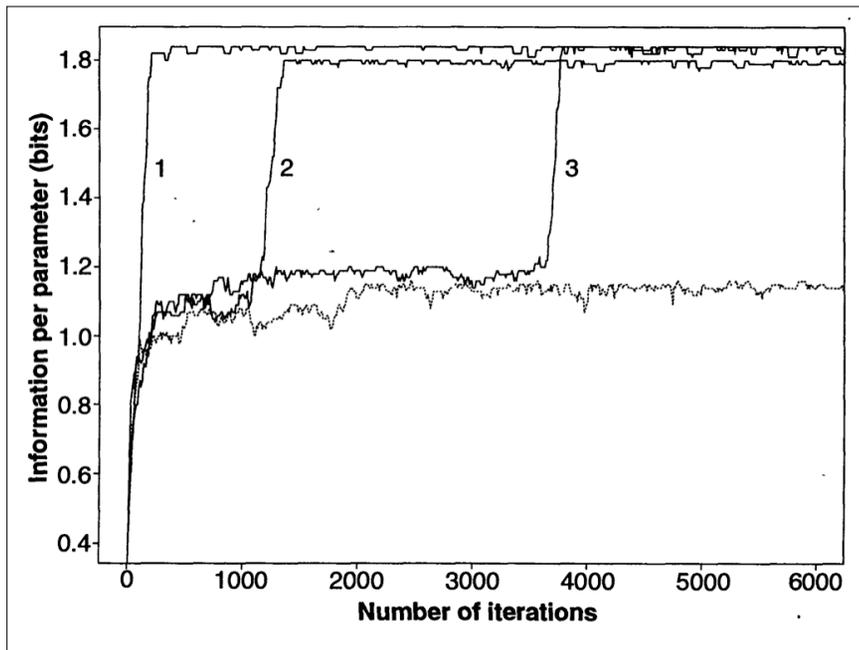
 pick new x_i according to that distribution

Issues

- Burnin - how long must we run the chain to reach stationarity?
- Mixing - how long a post-burnin sample must we take to get a good sample of the stationary distribution? (Recall that individual samples are not independent, and may not “move” freely through the sample space. Also, many isolated modes.)

Variants & Extensions

- “Phase Shift” - may settle on suboptimal solution that overlaps part of motif. Periodically try moving all motif instances a few spaces left or right.
- Algorithmic adjustment of pattern width: Periodically add/remove flanking positions to maximize (roughly) average relative entropy per position
- Multiple patterns per string

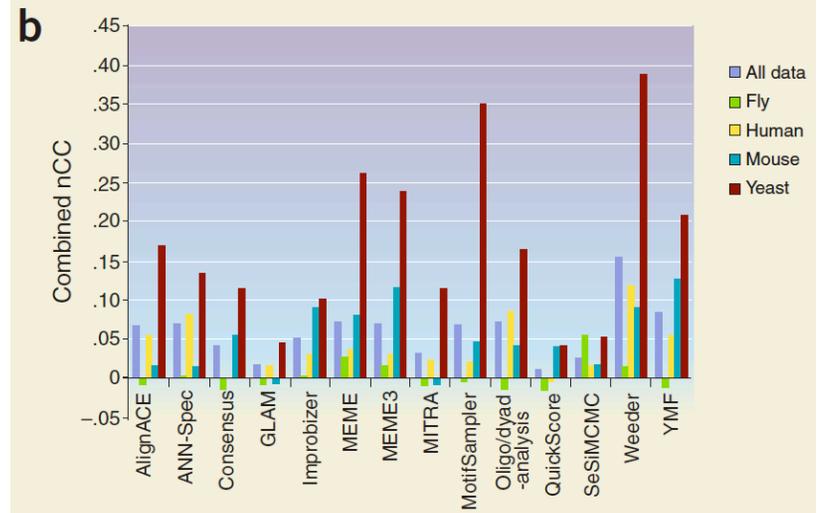
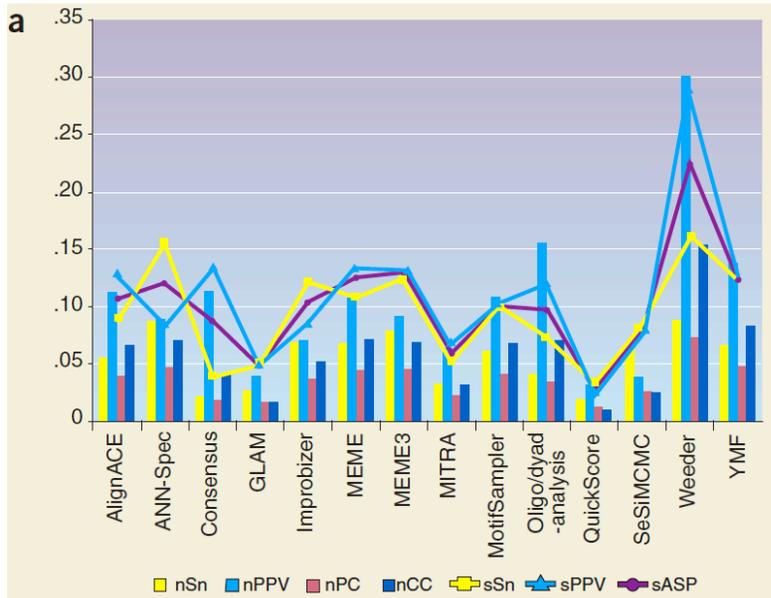


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Assessing computational tools for the discovery of transcription factor binding sites

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- dozen tools
- Real 'motifs' (Transfac)
- 56 data sets (human, mouse, fly, yeast)
- 'real', 'generic', 'Markov'
- expert users, top prediction only



Summary

- Important problem: a key to understanding gene regulation
- Hard problem: short, degenerate signals amidst much noise
- Many variants have been tried, for representation, search, and discovery. We looked at only a few:
 - Weight matrix models for representation & search
 - greedy, MEME and Gibbs for discovery
- Still much room for improvement. *Comparative genomics*, i.e. cross-species comparison is very promising