

CSE P 527

Markov Models and Hidden Markov Models



http://upload.wikimedia.org/wikipedia/commons/b/ba/Calico_cat

Dosage Compensation and X-Inactivation

2 copies (mom/dad) of each chromosome 1-22

Mostly, both copies of each gene are expressed

E.g., A B O blood group defined by 2 alleles of 1 gene

Women (XX) get double dose of X genes (vs XY)?

So, early in embryogenesis:

- One X randomly inactivated in each cell
 - Choice maintained in daughter cells
- } How?

Calico: a major coat color gene is on X

Reminder: Proteins “Read” DNA

E.g.:

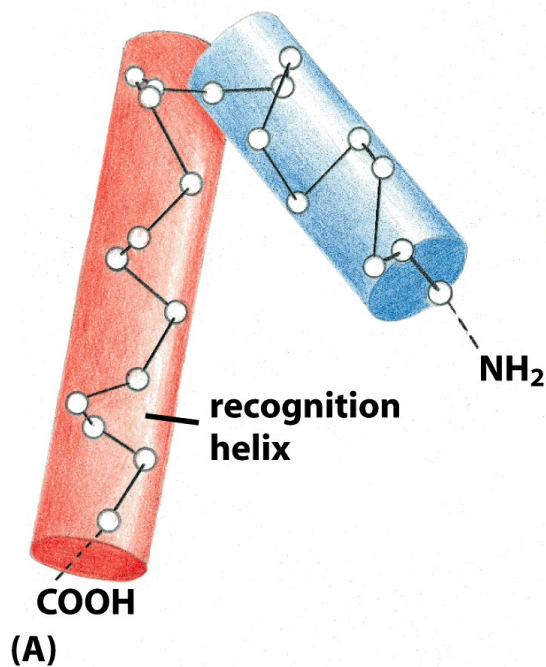


Figure 7-10 Molecular Biology of the Cell 5/e (© Garland Science 2008)

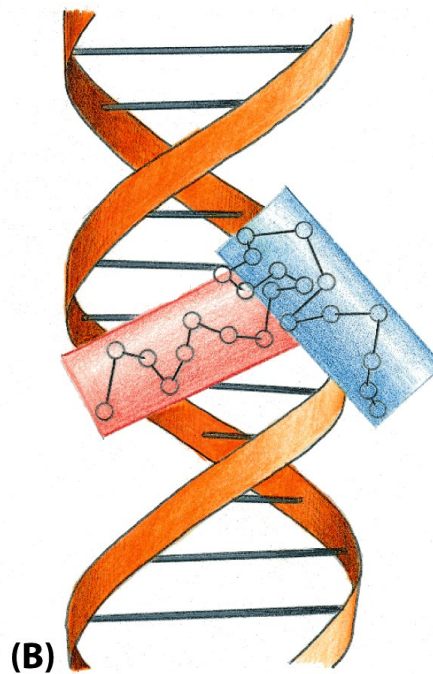


Figure 7-19 Molecular Biology of the Cell 5/e (© Garland Science 2008)



Down in the Groove

Different patterns of hydrophobic methyls, potential H bonds, etc. at edges of different base pairs. They're accessible, esp. in major groove

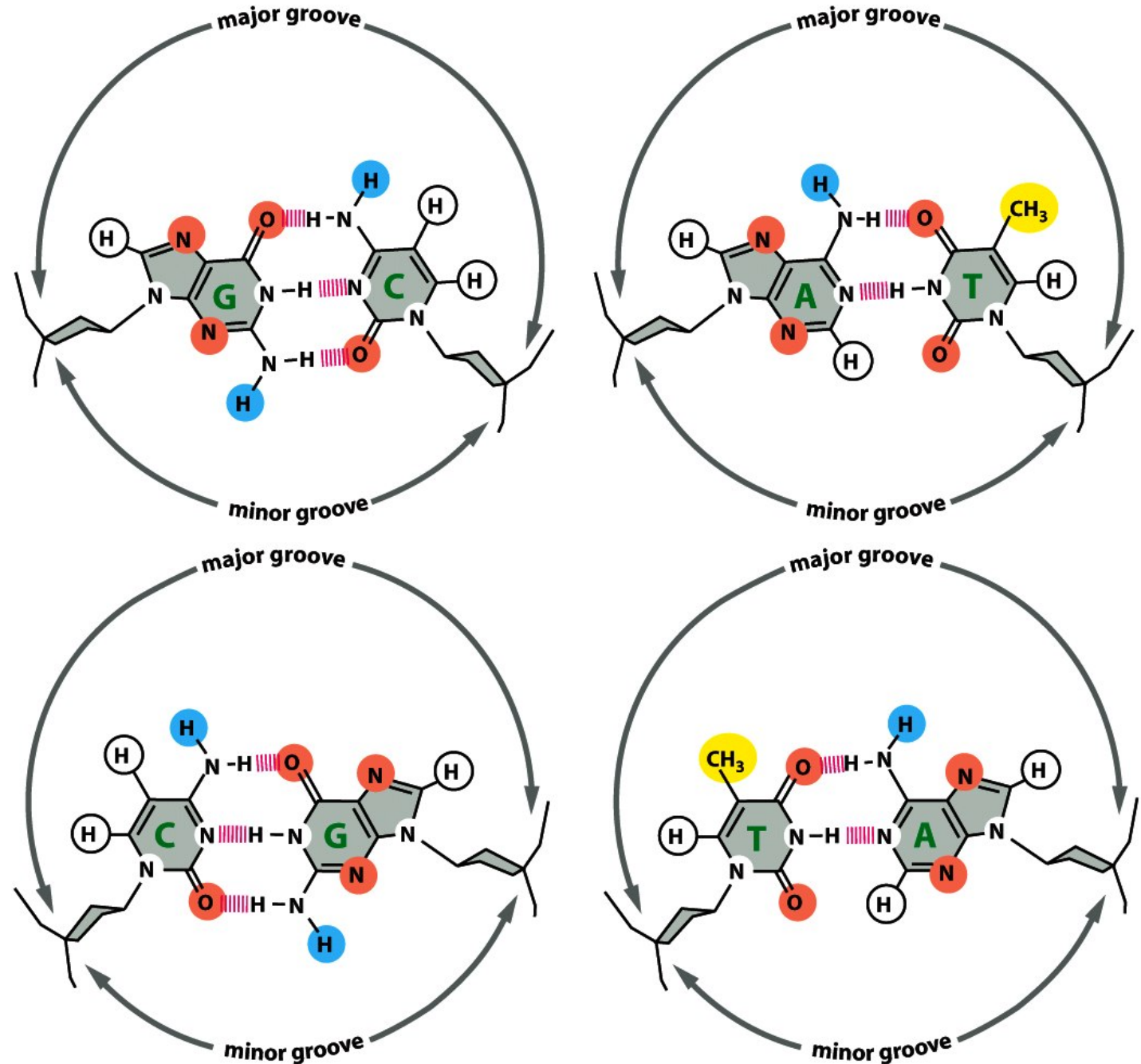
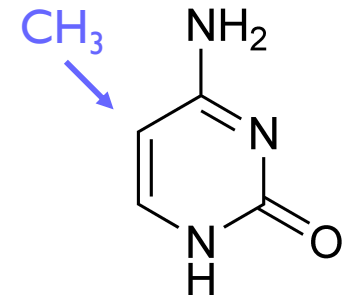


Figure 7-7 Molecular Biology of the Cell 5/e (© Garland Science 2008)

DNA Methylation

CpG - 2 adjacent nts, same strand
(not Watson-Crick pair; “p” mnemonic for the phosphodiester bond of the DNA backbone)

C of CpG is often (70-80%) methylated in mammals i.e., CH₃ group added (both strands)



cytosine

Same Pairing

Methyl-C
alters major
groove
profile (\therefore TF
binding), but
not base-
pairing,
transcription
or replication

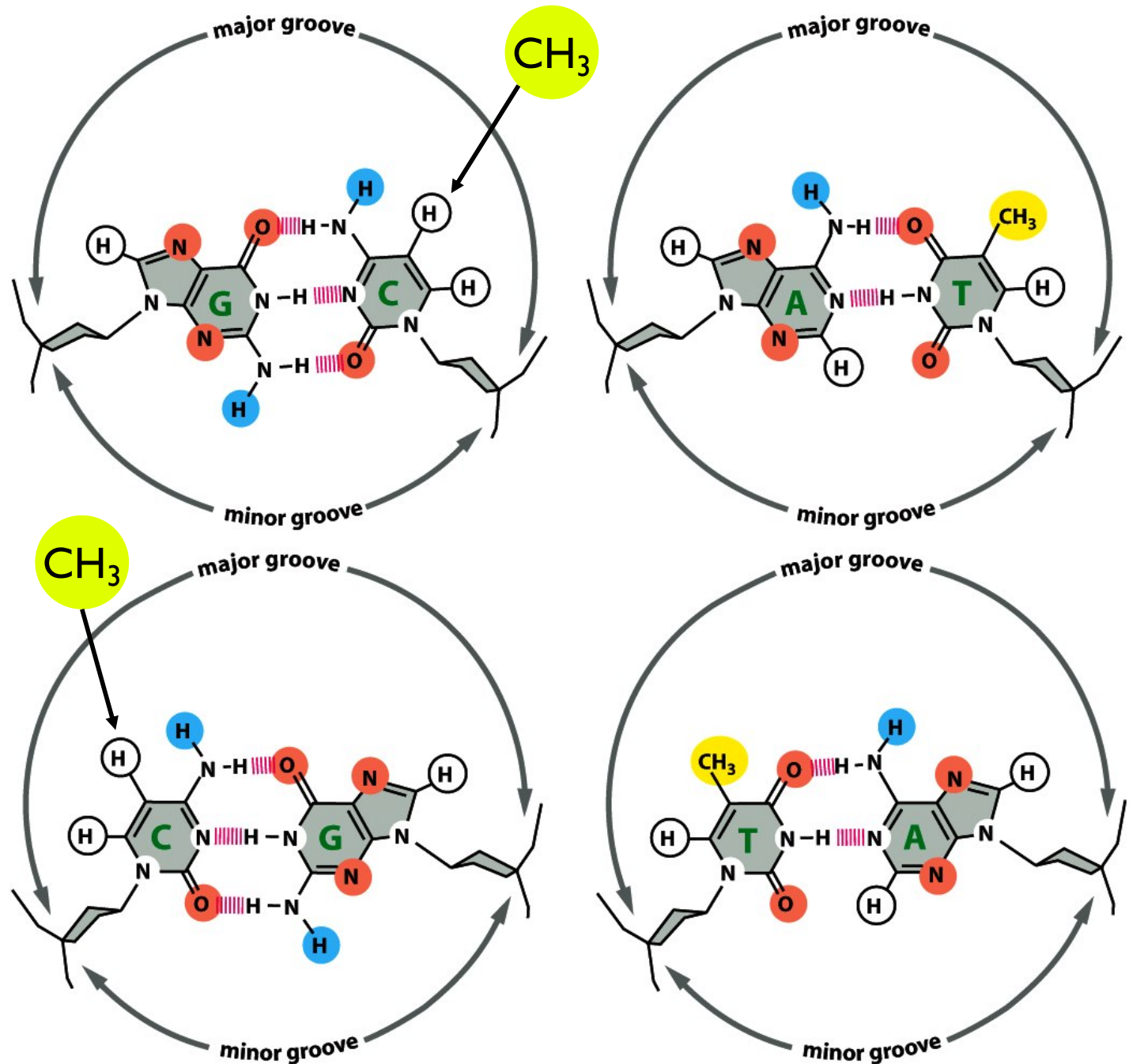


Figure 7-7 Molecular Biology of the Cell 5/e (© Garland Science 2008)

Calico Cats



http://upload.wikimedia.org/wikipedia/commons/b/ba/Calico_cat

Calico cat story: patchwork coat-color in some female cats partially explained by X-inactivation
And heavily methylating the inactive X is part of the mechanism of X-inactivation
And methylation is broadly important for other reasons, and sculpts the genome...

DNA Methylation—Why

In vertebrates, it generally silences transcription

(Epigenetics) X-inactivation, imprinting, repression of mobile elements, cancers, aging, and developmental differentiation

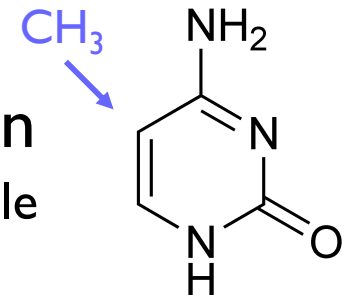
E.g., if a stem cell divides, one daughter fated to be liver, other kidney, need to

- (a) Turn off liver genes in kidney & vice versa, and
- (b) *Remember that through subsequent cell divisions*

How? One way:

- (a) Methylate genes, esp. promoters, to silence them
- (b) After ÷, DNA methyltransferases convert hemi- to fully-methylated
(not trivial: deleting methyltransferase is embryonic-lethal in mice)

Major exception: promoters of “housekeeping” genes



cytosine

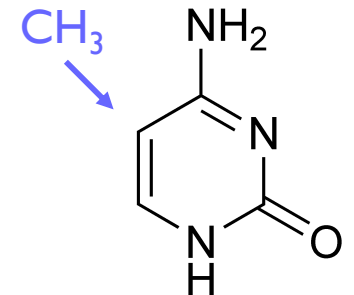
“CpG Islands”

Methyl-C mutates to T relatively easily

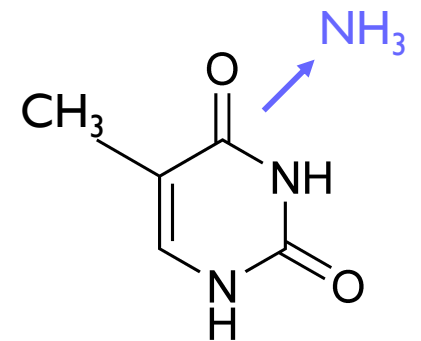
Net: CpG is less common than
expected genome-wide:

$$f(\text{CpG}) < f(\text{C}) * f(\text{G})$$

BUT in some regions (e.g. active
promoters), CpGs remain
unmethylated, so CpG \rightarrow TpG less
likely there: makes “CpG Islands”;
often mark gene-rich regions



cytosine



thymine

CpG Islands

CpG Islands

More CpG than elsewhere (say, $\text{CpG/GpC} > 50\%$)

More C & G than elsewhere, too (say, $\text{C+G} > 50\%$)

Typical length: few 100 to few 1000 bp

Questions

Is a short sequence (say, 200 bp) a CpG island or not?

Given long sequence (say, 10-100kb), find CpG islands?

Markov & Hidden Markov Models

References (see also online reading page):

Eddy, "What is a hidden Markov model?" *Nature Biotechnology*, 22, #10 (2004) 1315-6.

Durbin, Eddy, Krogh and Mitchison, "Biological Sequence Analysis", Cambridge, 1998 (esp. chs 3, 5)

Rabiner, "A Tutorial on Hidden Markov Models and Selected Application in Speech Recognition," *Proceedings of the IEEE*, v 77 #2, Feb 1989, 257-286

Independence

A key issue: Previous models we've talked about assume *independence* of nucleotides in different positions - definitely unrealistic.

Markov models allow us to relax that assumption.

Markov Chains

A sequence x_1, x_2, \dots of random variables is a *k-th order Markov chain* if, for all i , i^{th} value is independent of all but the previous k values:

$$P(x_i \mid \underbrace{x_1, x_2, \dots, x_{i-1}}_{i-1}) = P(x_i \mid \underbrace{x_{i-k}, x_{i-k+1}, \dots, x_{i-1}}_{k \text{ typically } \ll i-1})$$

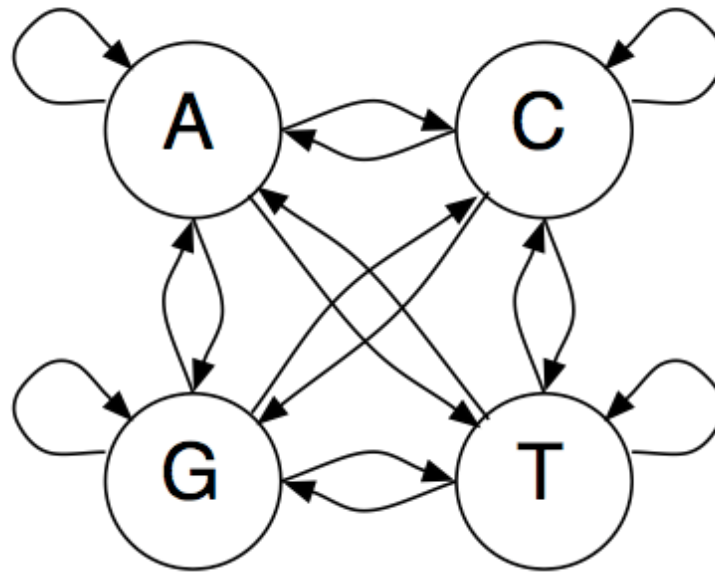
Example 1: Uniform random ACGT

Example 2: Weight matrix model

Example 3: ACGT, but $\downarrow \text{Pr}(\text{G following C})$

} 0th
order
}
1st
order

A Markov Model (1st order)

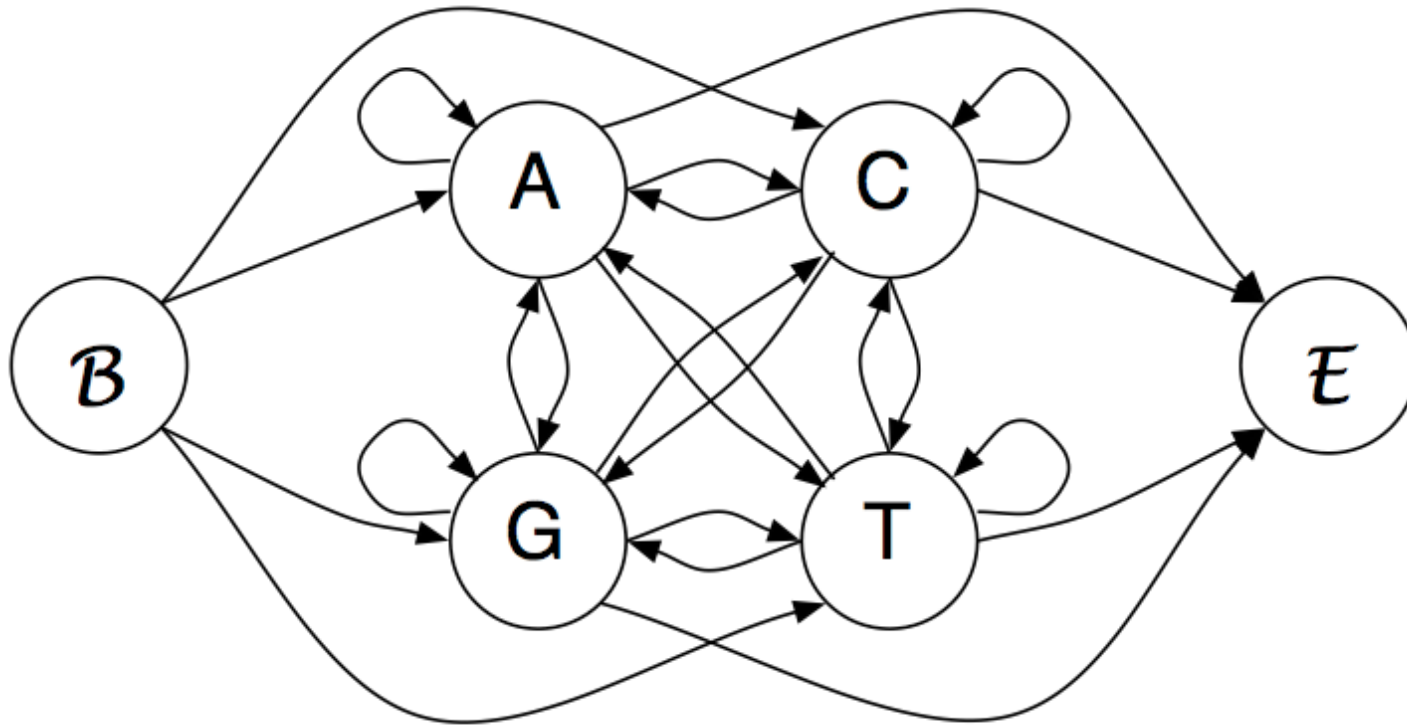


States: A,C,G,T

Emissions: corresponding letter

Transitions: $a_{st} = P(x_i = t \mid x_{i-1} = s)$ ← 1st order

A Markov Model (1st order)



States: A,C,G,T

Emissions: corresponding letter

Transitions: $a_{st} = P(x_i = t \mid x_{i-1} = s)$

Begin/End states

Pr of emitting sequence x

$$x = x_1 x_2 \dots x_n$$

$$P(x) = P(x_1, x_2, \dots, x_n)$$

law of probability
("chain rule")

$$= P(x_1) \cdot P(x_2 \mid x_1) \cdots P(x_n \mid x_{n-1}, \dots, x_1)$$

$$= P(x_1) \cdot P(x_2 \mid x_1) \cdots P(x_n \mid x_{n-1})$$

if 1st
order MC

$$= P(x_1) \prod_{i=1}^{n-1} a_{x_i, x_{i+1}}$$

$$= \prod_{i=0}^{n-1} a_{x_i, x_{i+1}} \quad (\text{with Begin state})$$

Training

Max likelihood estimates for transition probabilities are just the frequencies of transitions when emitting the training sequences

E.g., from 48 CpG islands in 60k bp:

+	A	C	G	T	-	A	C	G	T
A	0.180	0.274	0.426	0.120	A	0.300	0.205	0.285	0.210
C	0.171	0.368	<u>0.274</u>	0.188	C	0.322	0.298	<u>0.078</u>	0.302
G	0.161	0.339	0.375	0.125	G	0.248	0.246	0.298	0.208
T	0.079	0.355	0.384	0.182	T	0.177	0.239	0.292	0.292

Discrimination/Classification

Log likelihood ratio of CpG model vs background model

$$S(x) = \log \frac{P(x|+\text{model})}{P(x|-\text{model})} = \sum_{i=1}^L \log \frac{a_{x_{i-1},x_i}^+}{a_{x_{i-1},x_i}^-} = \sum_{i=1}^L \log \beta_{x_{i-1},x_i}$$

Input seq

Prev slide

β	A	C	G	T
A	-0.740	0.419	0.580	-0.803
C	-0.913	0.302	1.812	-0.685
G	-0.624	0.461	0.331	-0.730
T	-1.169	0.573	0.393	-0.679

CpG Island Scores

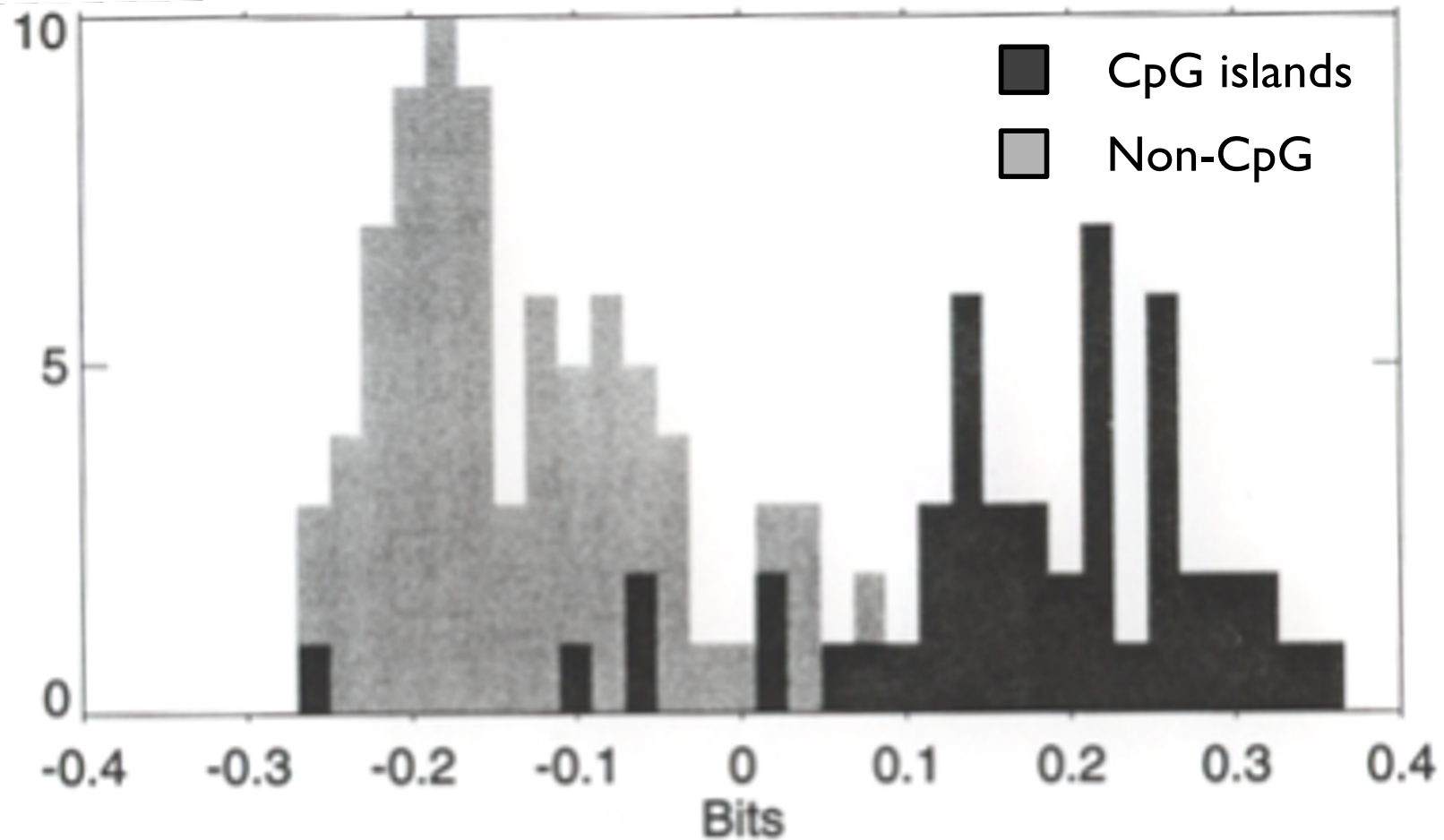


Figure 3.2 *Histogram of length-normalized scores.*

Questions

Q1: Given a *short* sequence, is it more likely from feature model or background model? [Above](#)

Q2: Given a *long* sequence, where are the features in it (if any)

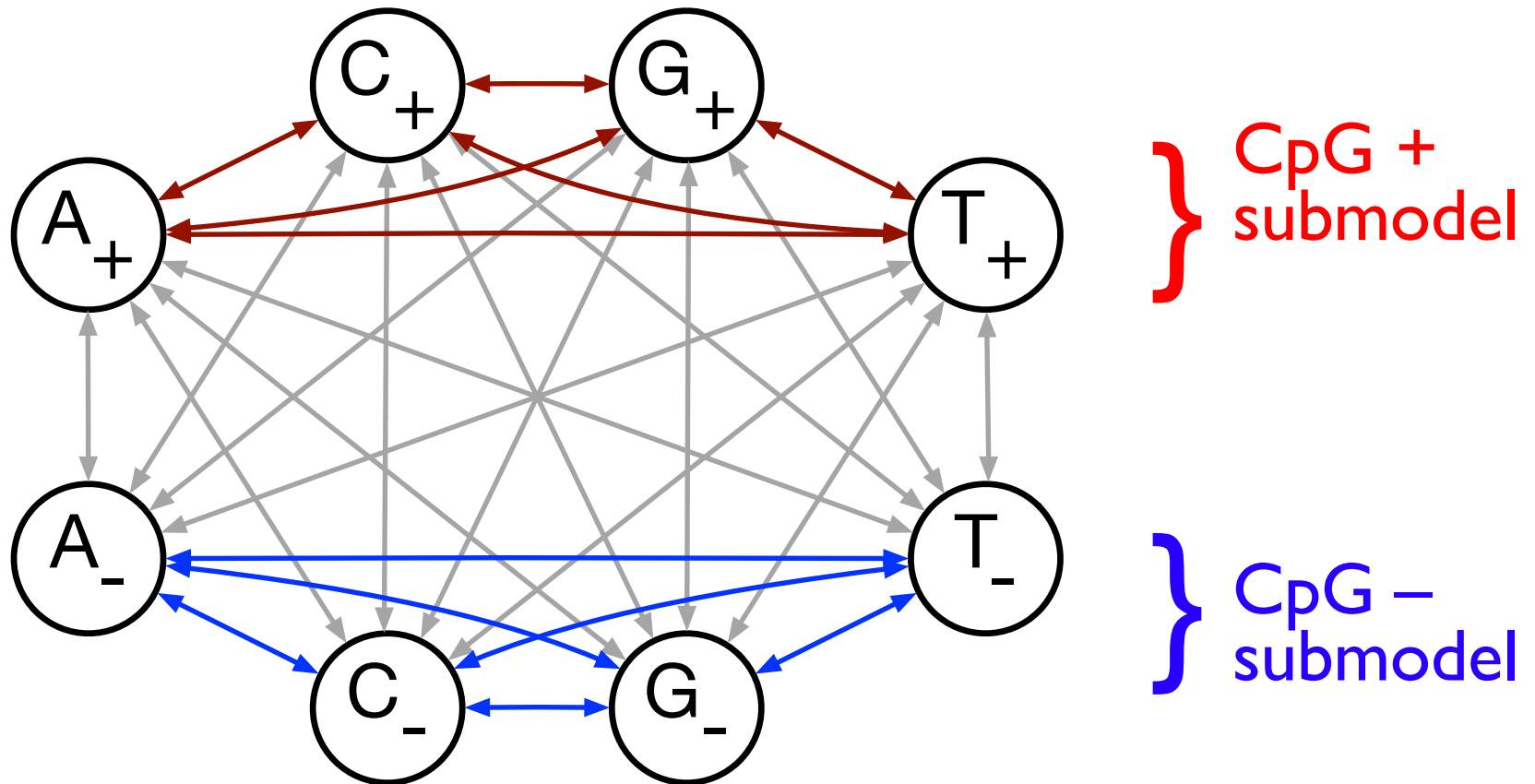
[Approach 1](#): score 100 bp (e.g.) windows

Pro: simple

Con: arbitrary, fixed length, inflexible

[Approach 2](#): *combine +/- models.*

Combined Model



Emphasis is “Which (hidden) state?” not “Which model?”

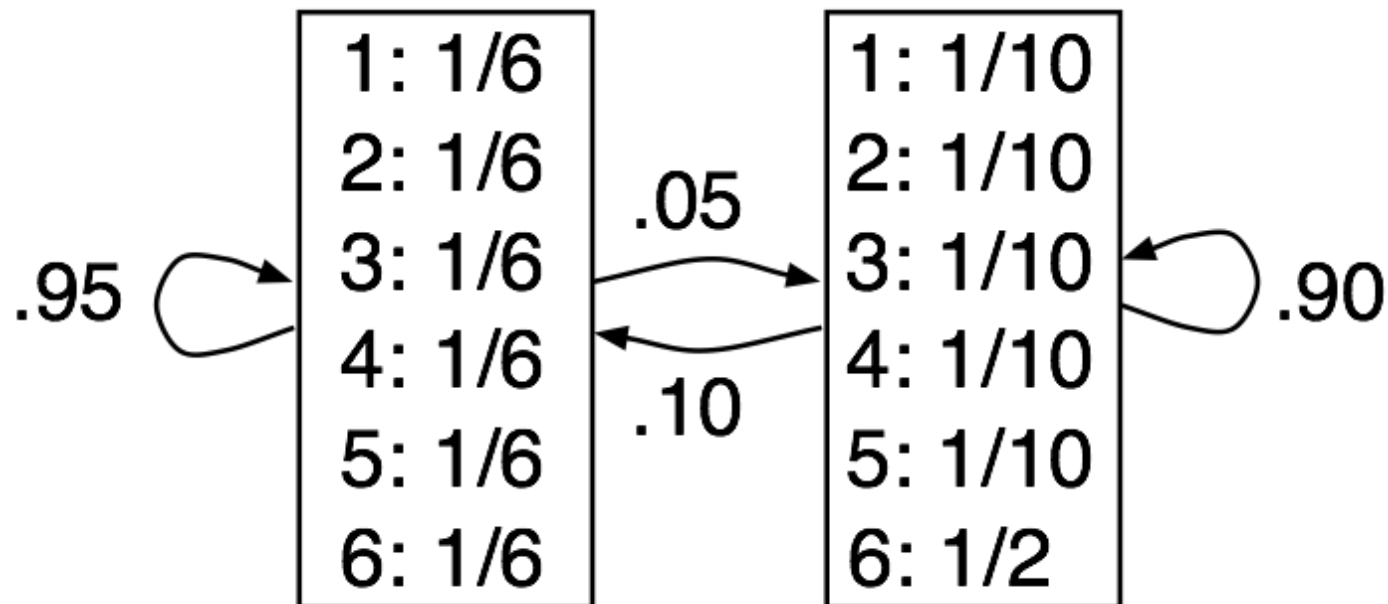
Hidden Markov Models

(HMMs; Claude Shannon, 1948)

States:	$1, 2, 3, \dots$
Paths:	sequences of states $\pi = (\pi_1, \pi_2, \dots)$
Transitions:	$a_{k,l} = P(\pi_i = l \mid \pi_{i-1} = k)$
Emissions:	$e_k(b) = P(x_i = b \mid \pi_i = k)$
Observed data:	emission sequence
Hidden data:	state/transition sequence

The Occasionally Dishonest Casino

1 fair die, 1 “loaded” die, occasionally swapped



[illegible]

Figure 3.5

Rolls: Visible data—300 rolls of a die as described above.

Die: Hidden data—which die was actually used for that roll (F = fair, L = loaded).

Viterbi: the prediction by the Viterbi algorithm is shown.

Inferring hidden stuff

Joint prob of a given path π & emission sequence x :

$$P(x, \pi) = a_{0, \pi_1} \prod_{i=1}^n e_{\pi_i}(x_i) \cdot a_{\pi_i, \pi_{i+1}}$$

But π is hidden; what to do? Some alternatives:

Most probable single path

$$\pi^* = \arg \max_{\pi} P(x, \pi)$$

Sequence of most probable states

$$\hat{\pi}_i = \arg \max_k P(\pi_i = k \mid x)$$

Etc.

Notation:

$\max_x F(x)$ = the maximum **y-value** attained by $F()$

$\arg \max_x F(x)$ = the **x-value** where that occurs

The Viterbi Algorithm: The most probable path

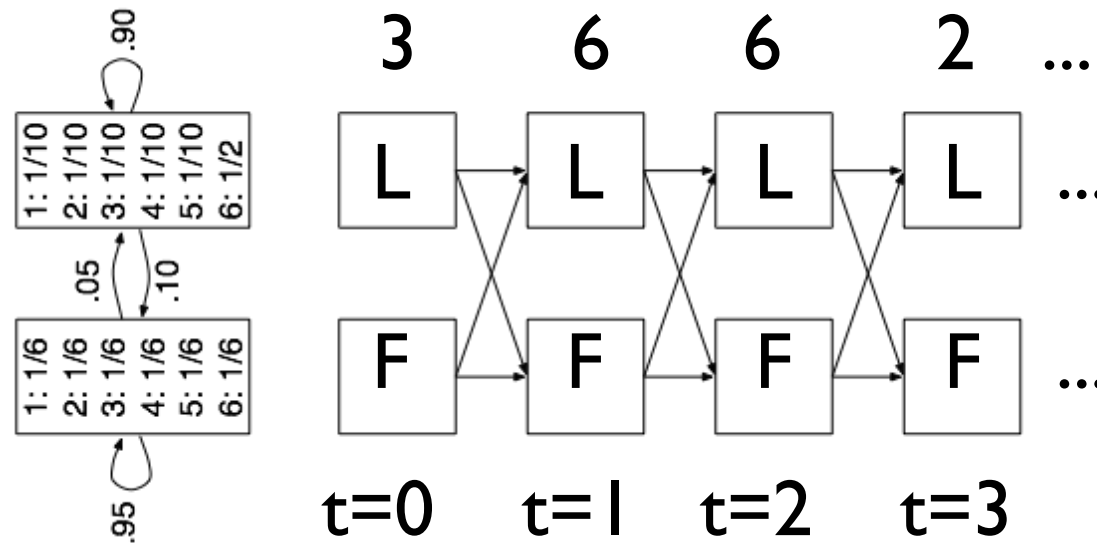
Viterbi finds: $\pi^* = \arg \max_{\pi} P(x, \pi)$

Possibly there are 10^{99} paths of prob 10^{-99}
(If so, non-Viterbi approaches may be preferable.)

More commonly, one path (+ slight variants)
dominate others; Viterbi finds that

Key problem: exponentially many paths π

Unrolling an HMM



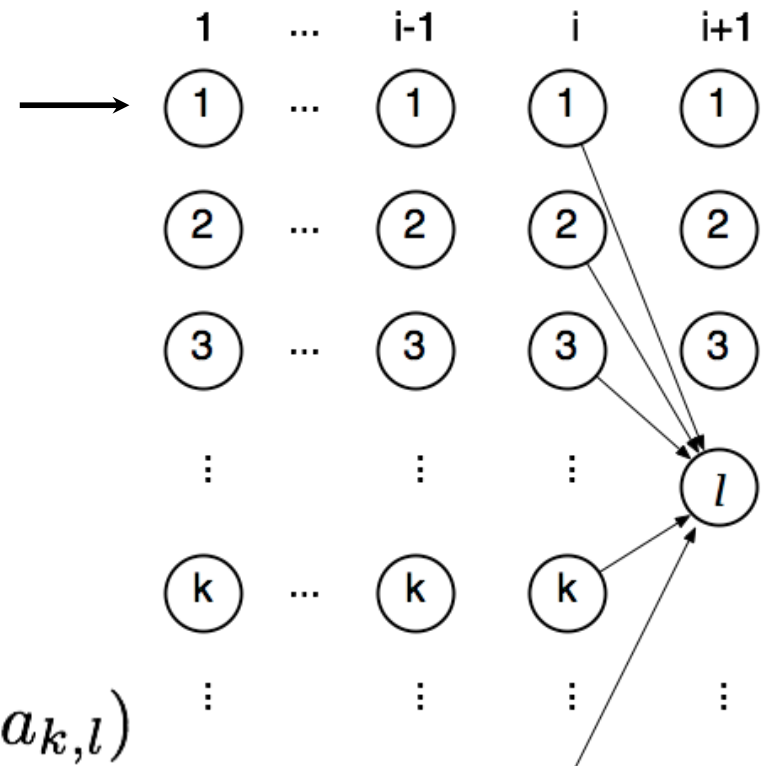
Conceptually, sometimes convenient
Note exponentially many paths

Viterbi

$v_l(i)$ = probability of the most probable path
emitting x_1, x_2, \dots, x_i and ending in state l

Initialize:

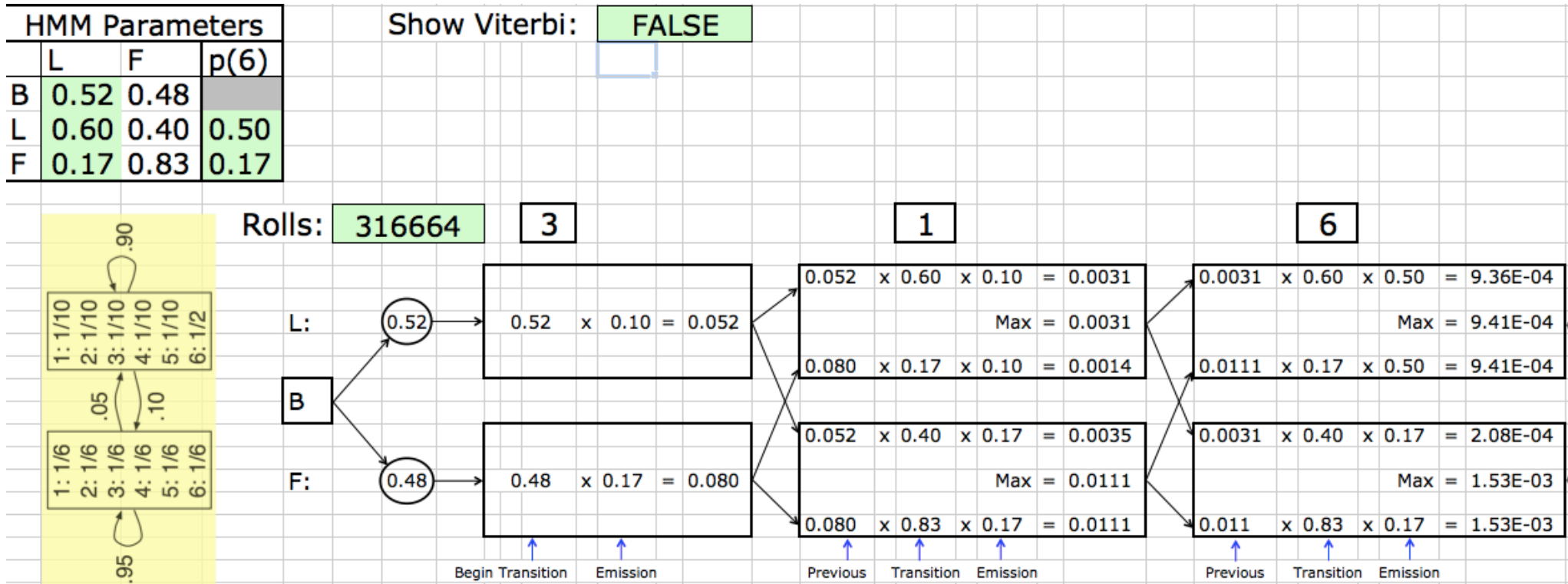
$$v_l(0) = \begin{cases} 1 & \text{if } l = \text{Begin state} \\ 0 & \text{otherwise} \end{cases}$$



General case:

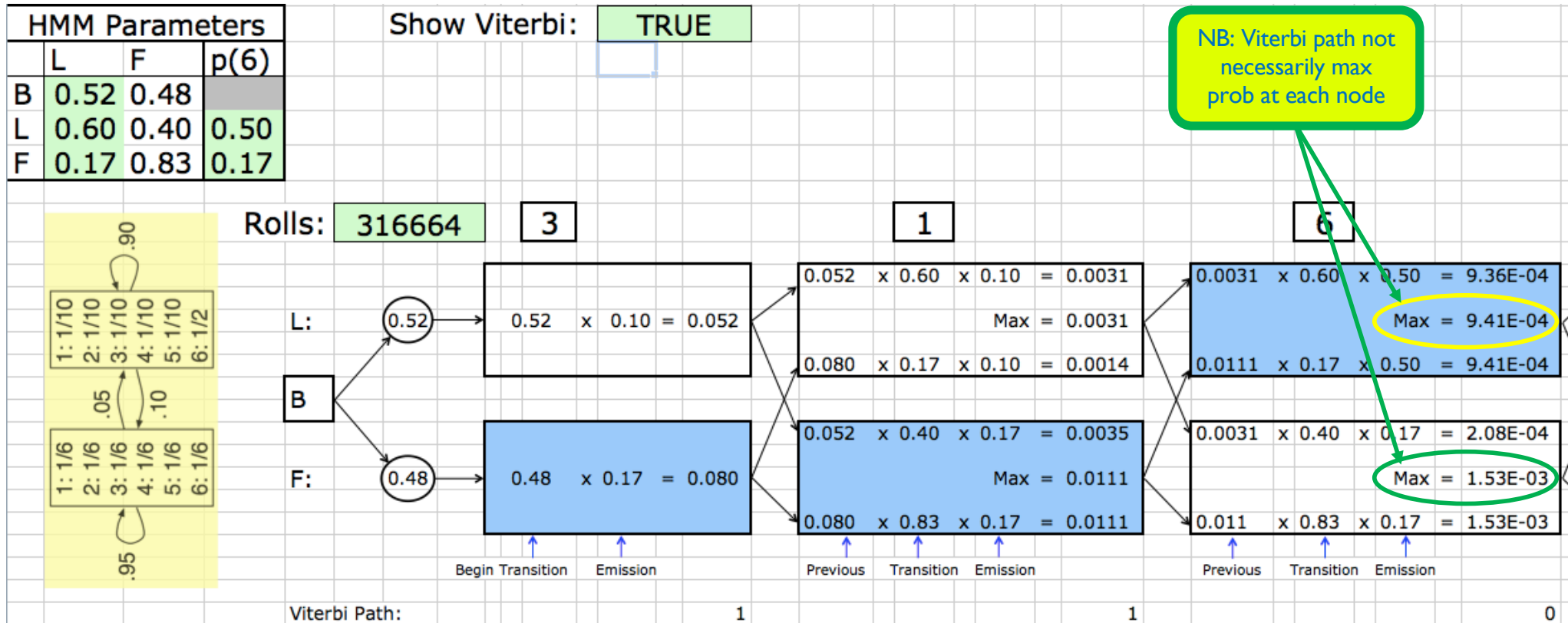
$$v_l(i+1) = e_l(x_{i+1}) \cdot \max_k (v_k(i) a_{k,l})$$

HMM Casino Example



(Excel spreadsheet on web; download & play...)

HMM Casino Example

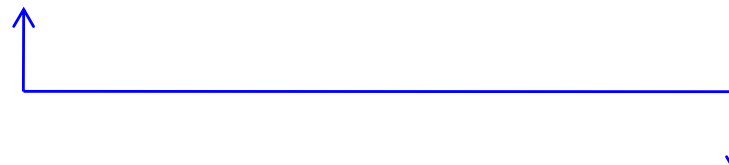


(Excel spreadsheet on web; download & play...)

Viterbi Traceback

Above finds *probability* of best path

To find the path itself, trace *backward* to the state k attaining the max at each stage


$$v_l(i+1) = e_l(x_{i+1}) \cdot \max_k (v_k(i) a_{k,l})$$

[illegible]

Figure 3.5

Rolls: Visible data—300 rolls of a die as described above.

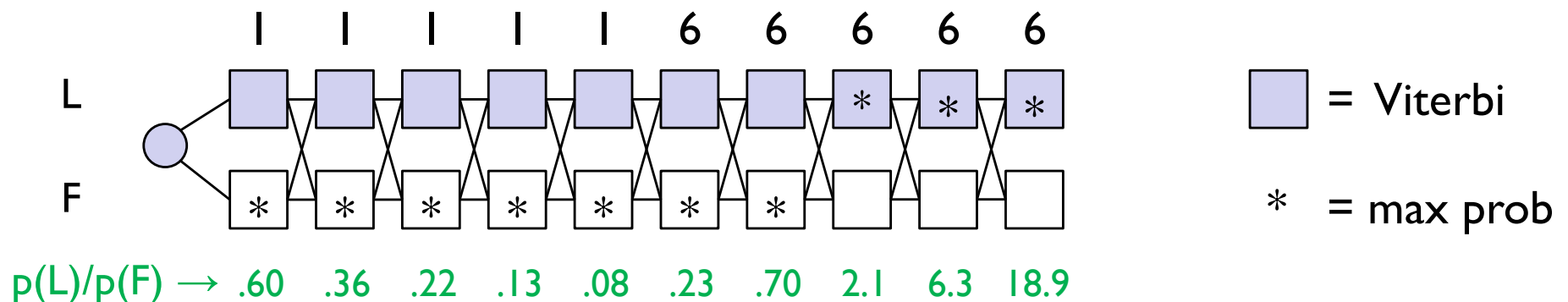
Die: Hidden data—which die was actually used for that roll (F = fair, L = loaded).

Viterbi: the prediction by the Viterbi algorithm is shown.

Most probable path \neq Sequence of most probable states

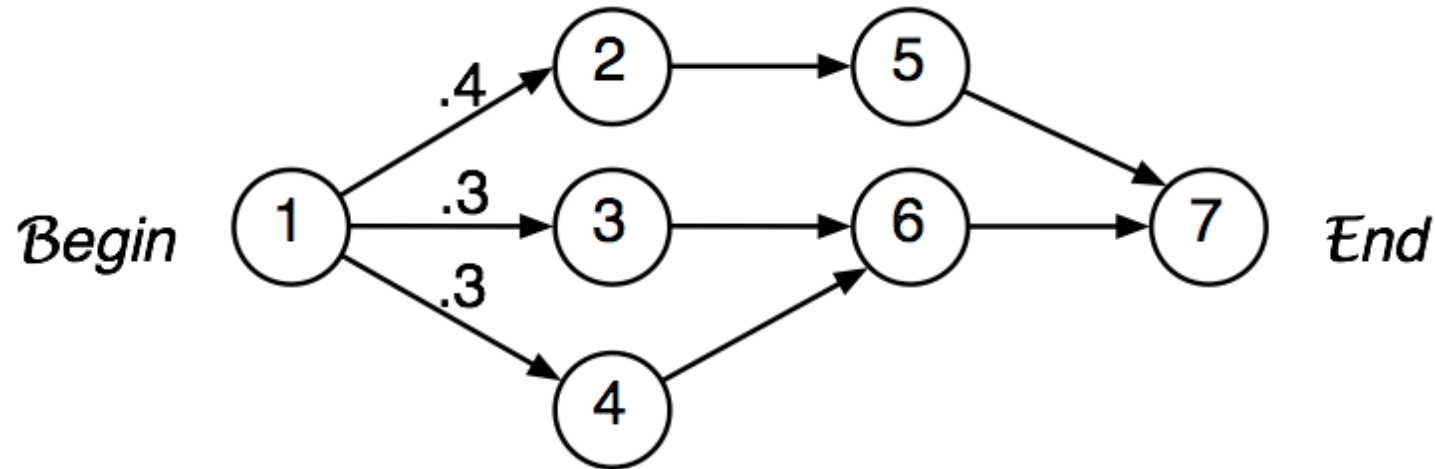
Another example, based on casino dice again:

Suppose $p(\text{fair} \leftrightarrow \text{loaded})$ transitions are 10^{-99} and roll sequence is 1 1 1 1 6 6...6 6 6; then fair state is more likely all through 1's & into the run of 6's, but eventually loaded wins, and the improbable $F \rightarrow L$ transitions make Viterbi = *all* L.



Is Viterbi “best”?

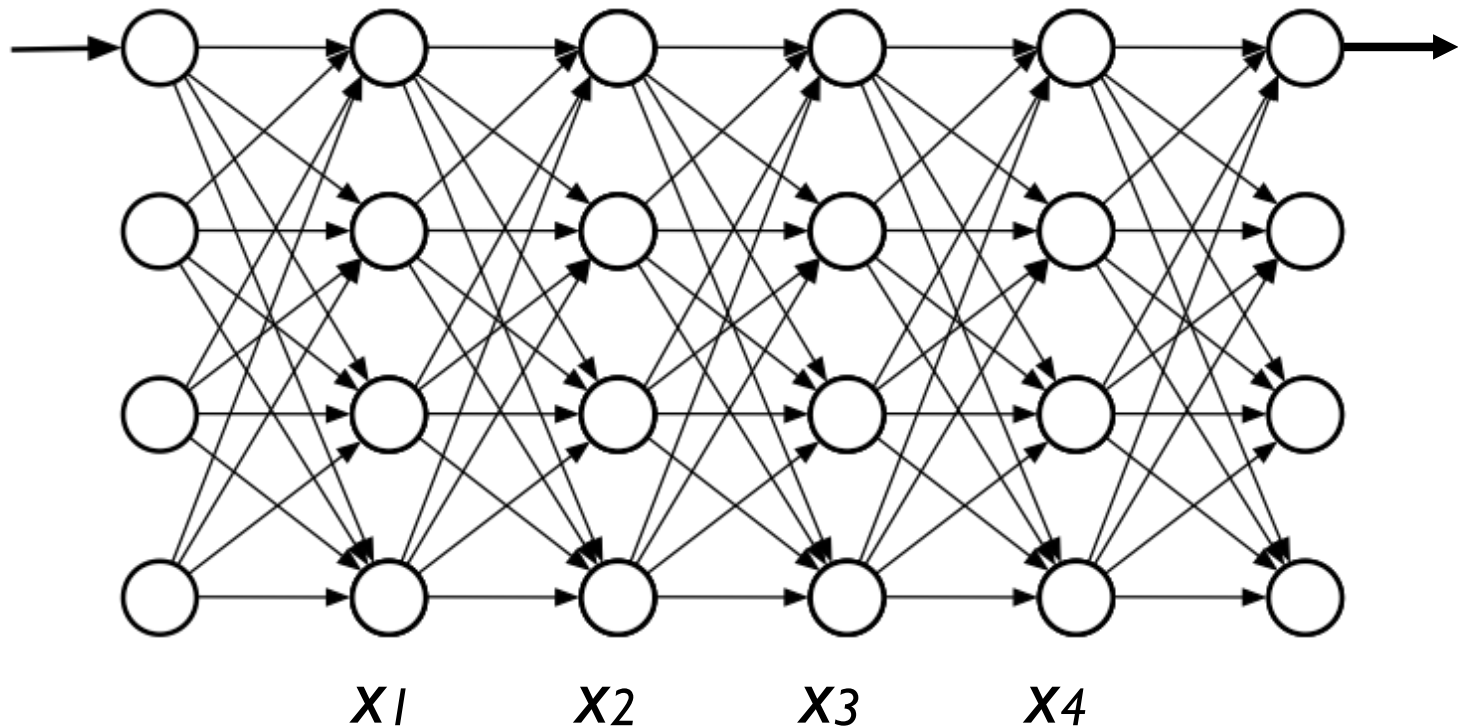
Viterbi finds $\pi^* = \arg \max_{\pi} P(x, \pi)$



Most probable (Viterbi) *path* goes through 5, but most probable *state* at 2nd step is 6 (i.e., Viterbi is not the only interesting answer.)

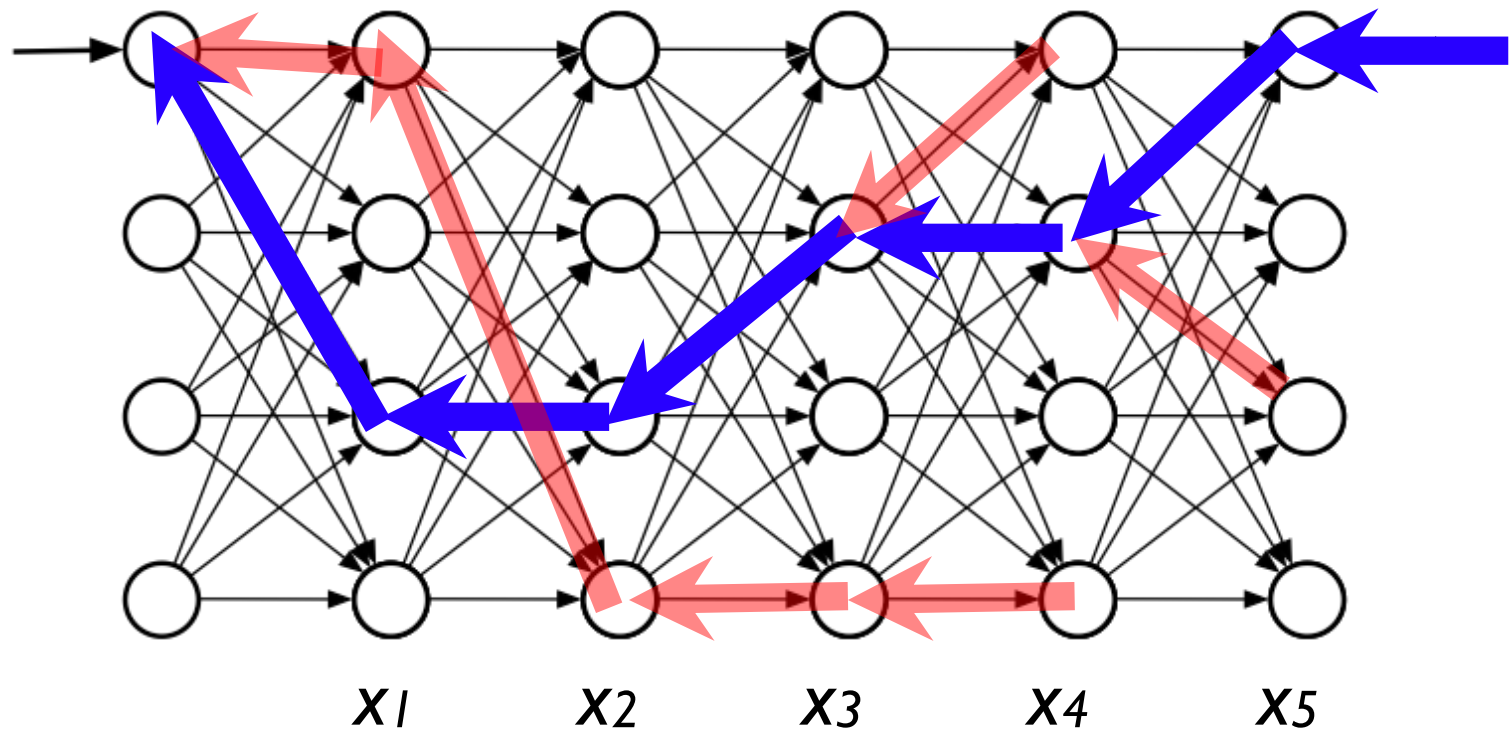
An HMM (unrolled)

States



Emissions/sequence positions →

Viterbi: best path to each state



Viterbi score:

$$v_l(i + 1) = e_l(x_{i+1}) \cdot \max_k (v_k(i) a_{k,l})$$

Viterbi
path^R:

$$back_l(i + 1) = \arg \max_k (v_k(i) a_{k,l})$$

Another Q: What's $P(x)$?

Given an HMM and a sequence x , Viterbi finds the single path π having the greatest probability of emitting x (and implicitly finds that probability $P(x, \pi)$)

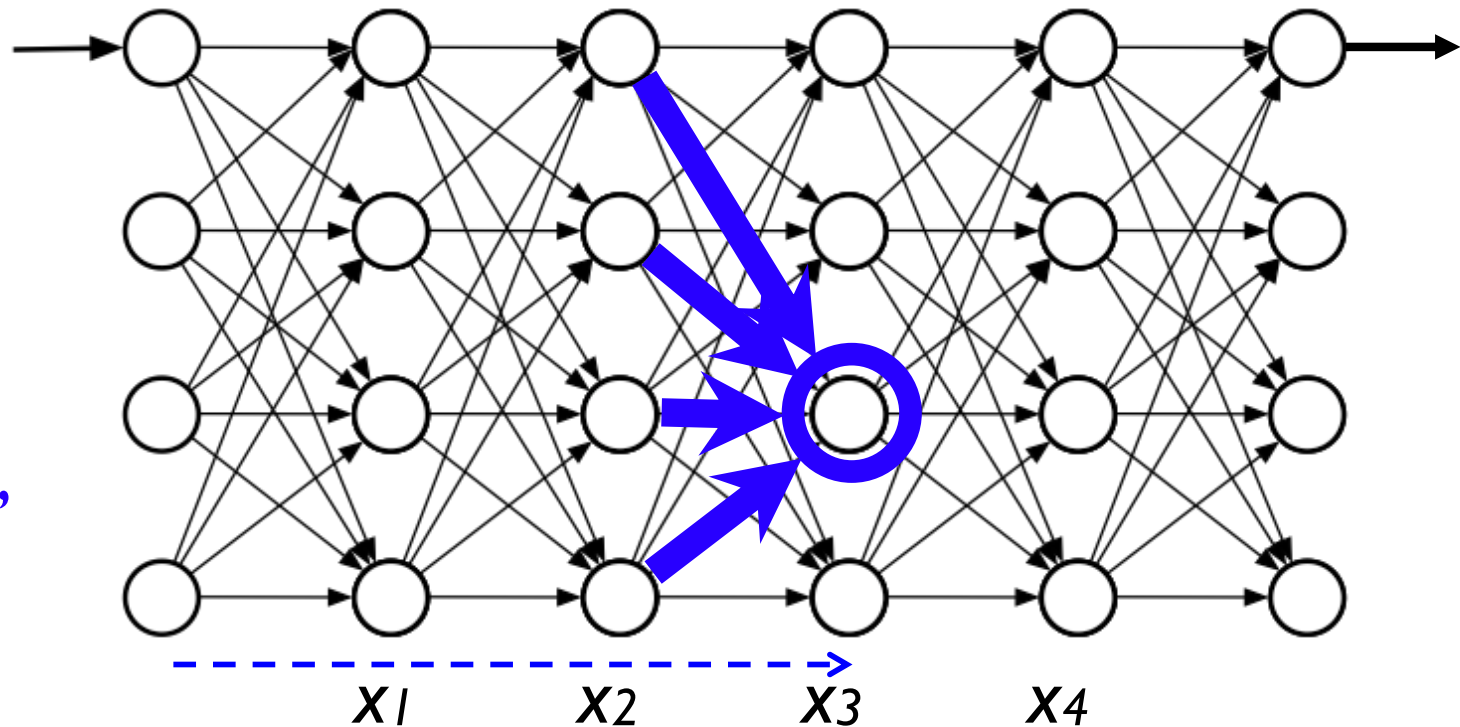
What if I don't care about π ? E.g., what is the probability $P(x)$ of emitting x , on *some* path?

Of course, $P(x) = \sum_{\pi} P(x, \pi)$, i.e. sum over all paths, but exponentially many, so nontrivial ...

Answer to this and related Qs is easiest to think about by focusing on intermediate states

The Forward Algorithm

For each state/time, want *total* probability of all paths leading to it, with previous emissions



$$f_k(i) \triangleq P(x_1 \dots x_i, \pi_i = k)$$

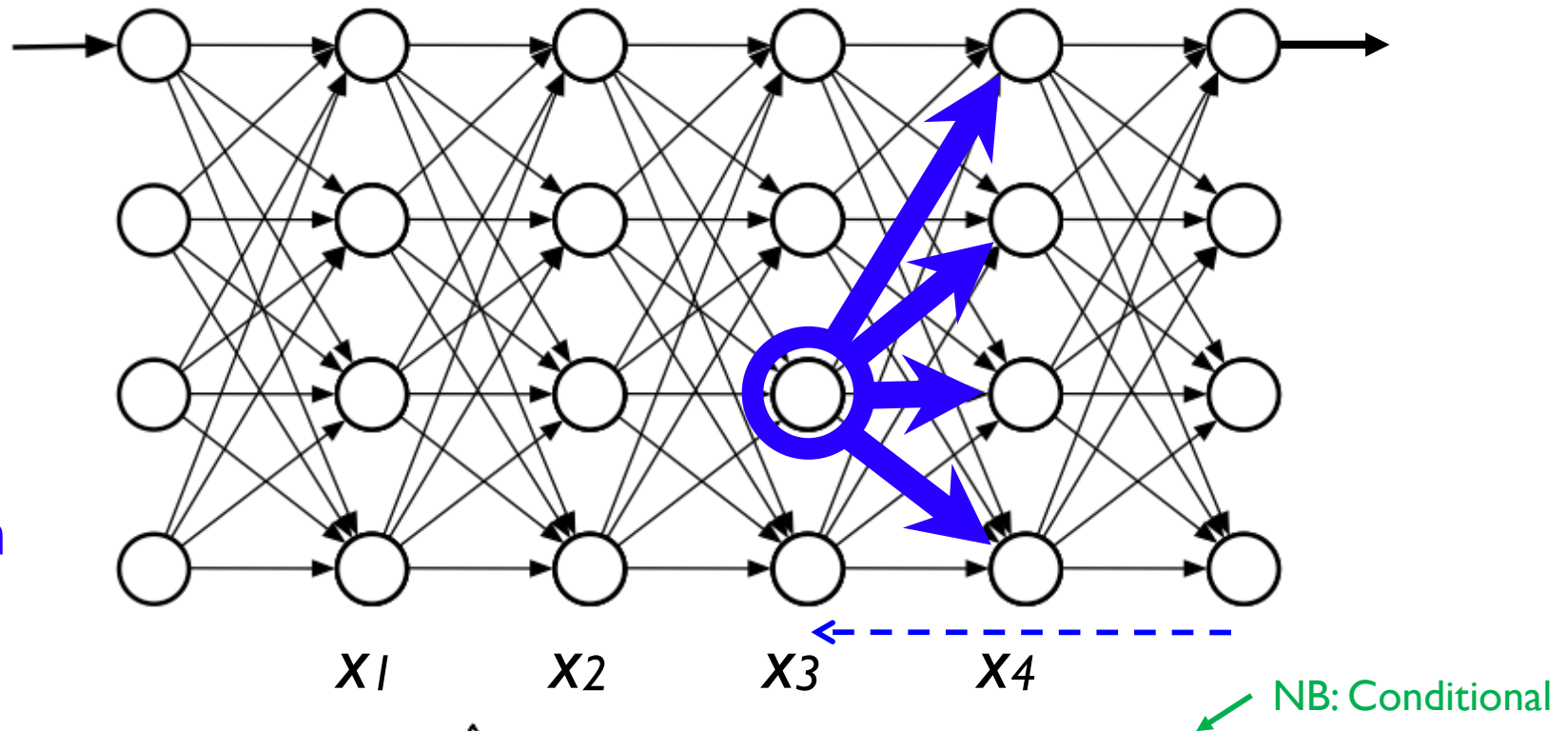
NB: Joint

$$f_l(i+1) = e_l(x_{i+1}) \sum_k f_k(i) a_{k,l}$$

$$P(x) = \sum_{\pi} P(x, \pi) = \sum_k f_k(n) a_{k,end}$$

The Backward Algorithm

Similar:
for each
state/time,
want total
probability
of all paths
from it, with
subsequent
emissions,
conditional
on that
state.



$$b_k(i) \triangleq P(x_{i+1} \cdots x_n \mid \pi_i = k)$$

$$b_k(i) = \sum_l a_{k,l} e_l(x_{i+1}) b_l(i+1)$$

$$b_k(n) = a_{k,end}$$

In state k at step i ?

$$P(x, \pi_i = k)$$

$$= P(x_1, \dots, x_i, \pi_i = k) \cdot P(x_{i+1}, \dots, x_n \mid x_1, \dots, x_i, \pi_i = k)$$

$$= P(x_1, \dots, x_i, \pi_i = k) \cdot P(x_{i+1}, \dots, x_n \mid \pi_i = k)$$

$$= f_k(i) \cdot b_k(i)$$

$$P(\pi_i = k \mid x) = \frac{P(x, \pi_i = k)}{P(x)} = \frac{f_k(i) \cdot b_k(i)}{P(x)}$$

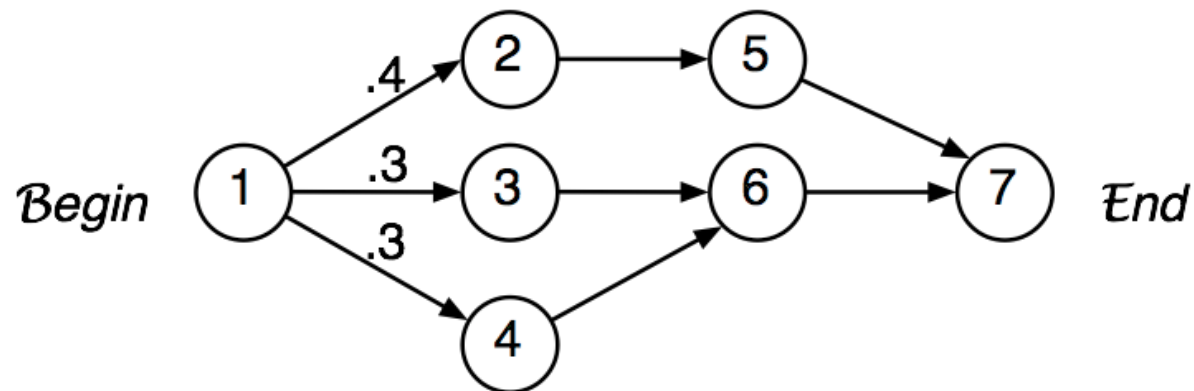
Posterior Decoding, I

Alternative 1: what's the most likely state at step i ?

$$\hat{\pi}_i = \arg \max_k P(\pi_i = k \mid x)$$

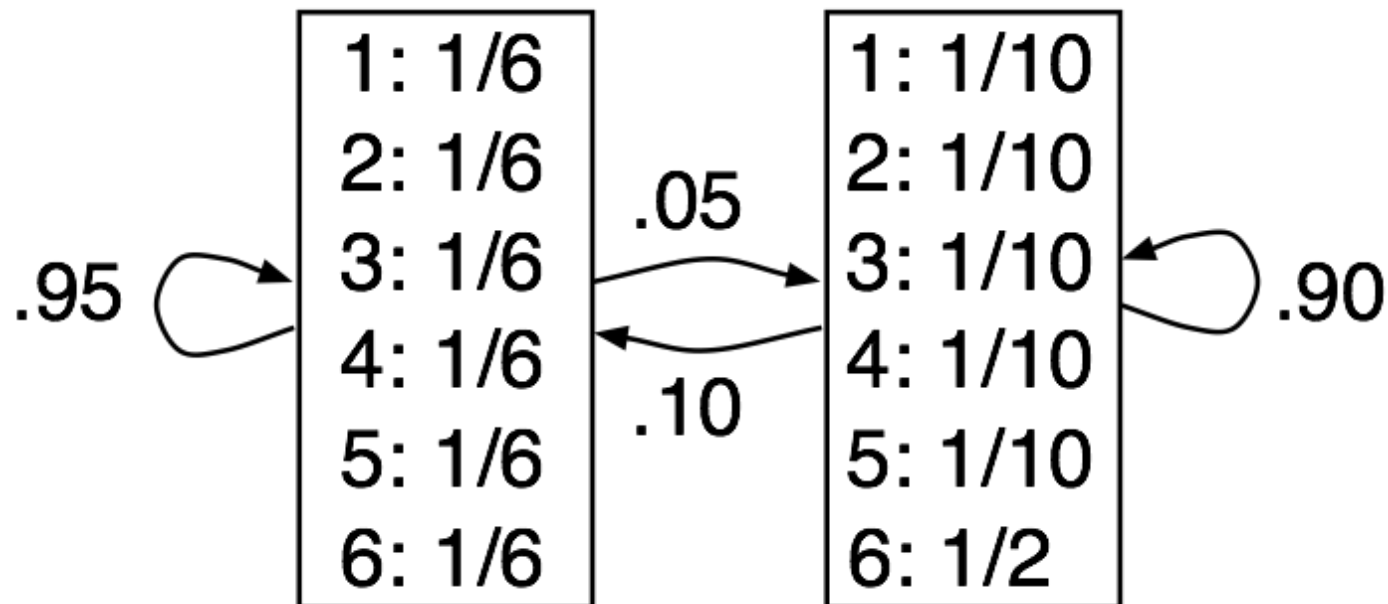
Note: the most likely sequence of states (a path) \neq the sequence of most likely states.

That may even be an illegal path! (E.g. 1,2,6,7 below)



The Occasionally Dishonest Casino

1 fair die, 1 “loaded” die, occasionally swapped



[illegible]

Figure 3.5

Rolls: Visible data—300 rolls of a die as described above.

Die: Hidden data—which die was actually used for that roll (F = fair, L = loaded).

Viterbi: the prediction by the Viterbi algorithm is shown.

Posterior Decoding

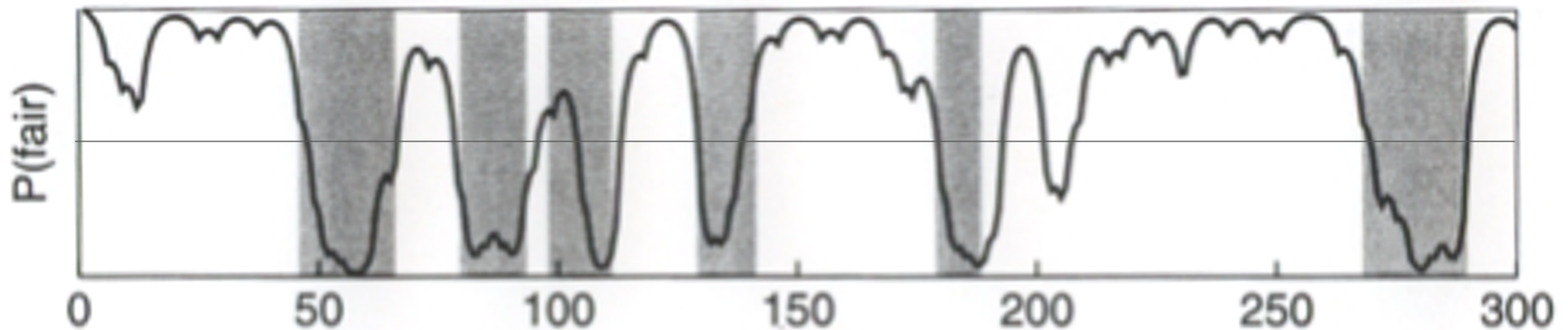


Figure 3.6 *The posterior probability of being in the state corresponding to the fair die in the casino example. The x axis shows the number of the roll. The shaded areas show when the roll was generated by the loaded die.*

Posterior Decoding, II

Alternative 1: what's most likely state at step i ?

$$\hat{\pi}_i = \arg \max_k P(\pi_i = k \mid x)$$

Alternative 2: given some function $g(k)$ on states, what's its expectation. E.g., what's probability of “+” model in CpG HMM ($g(k)=1$ iff k is “+” state)?

$$G(i \mid x) = \sum_k P(\pi_i = k \mid x) \cdot g(k)$$

CpG Islands again

Data: 41 human sequences, totaling 60kbp,
including 48 CpG islands of about 1kbp each

Viterbi:

Found 46 of 48
plus 121 “false positives”

Post-process:

46/48

67 false pos

Posterior Decoding:

same 2 false negatives
plus 236 false positives

46/48

83 false pos

Post-process: merge within
500; discard < 500

Training

Given model topology & training sequences,
learn transition and emission probabilities

If π known, then MLE is just frequency observed
in training data

$$\begin{aligned} a_{k,l} &= \frac{\text{count of } k \rightarrow l \text{ transitions}}{\text{count of } k \rightarrow \text{anywhere transitions}} \\ e_k(b) &= \dots \end{aligned}$$

← + pseudocounts?

If π hidden, then use EM:

given θ , estimate π ; given π estimate θ ; repeat } 2 ways

Viterbi Training

given θ , estimate π ; given π estimate θ ; repeat

Make initial estimates of parameters θ

Find Viterbi path π for each training sequence

Count transitions/emissions *on those paths*,
getting new θ

Repeat

Not rigorously optimizing desired likelihood, but
still useful & commonly used.

(Arguably good if you're doing Viterbi decoding.)

(And see note about “classification EM,” ~#45 in MLE-EM slides.)

Viterbi Training, II

given θ , estimate π ; given π estimate θ ; repeat

Not rigorously optimizing desired likelihood

What it IS doing is finding θ to maximize
contribution to likelihood from the most
probable paths

As noted earlier, with 10^{99} paths each with prob
near 10^{-99} , this may not be useful, but if a few
paths dominate the landscape, then it may be –
learned θ hopefully captures this

AKA “the forward-
backward alg”

Baum-Welch Training

EM: given θ , estimate π ensemble; then re-estimate θ

$$\begin{aligned} P(\pi_i = k, \pi_{i+1} = l \mid x, \theta) \\ = \frac{f_k(i \mid \theta) a_{k,l} e_l(x_{i+1}) b_l(i+1 \mid \theta)}{P(x \mid \theta)} \end{aligned}$$

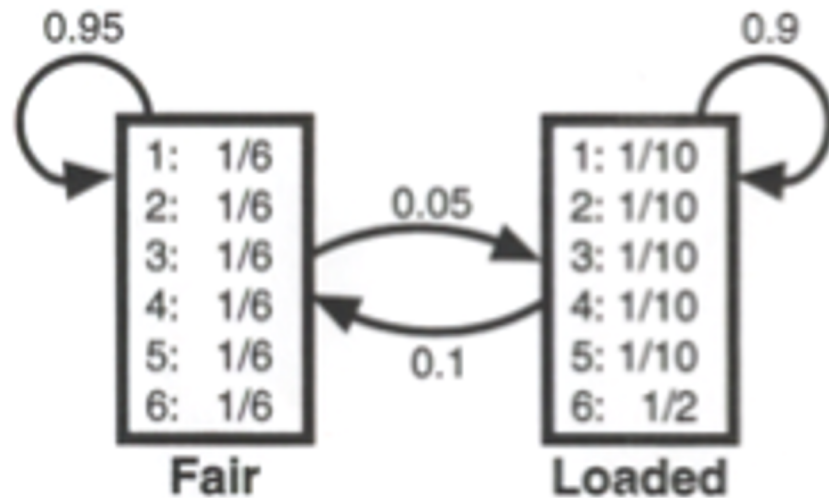
Estimated # of $k \rightarrow l$ transitions $\hat{A}_{k,l}$ on set of seqs x^j

$$= \sum_{\text{training seqs } x^j} \sum_i P(\pi_i = k, \pi_{i+1} = l \mid x^j, \theta)$$

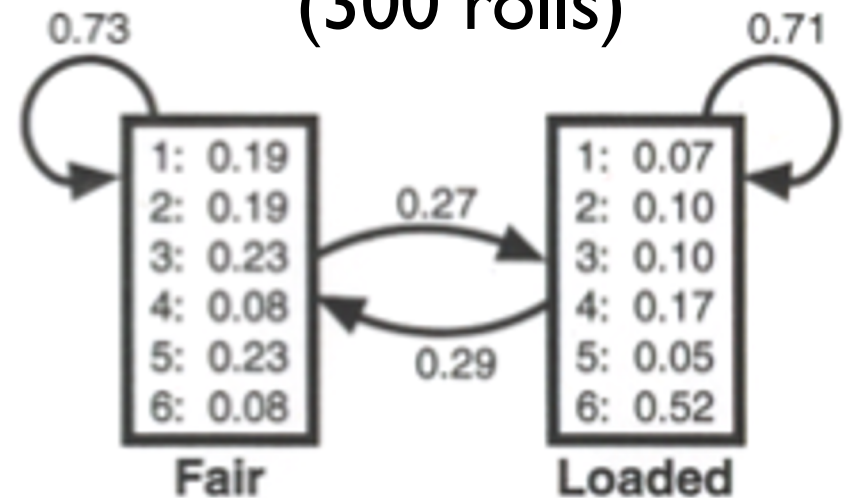
$$\text{New estimate } \hat{a}_{k,l} = \frac{\hat{A}_{k,l}}{\sum_l \hat{A}_{k,l}}$$

Emissions: similar

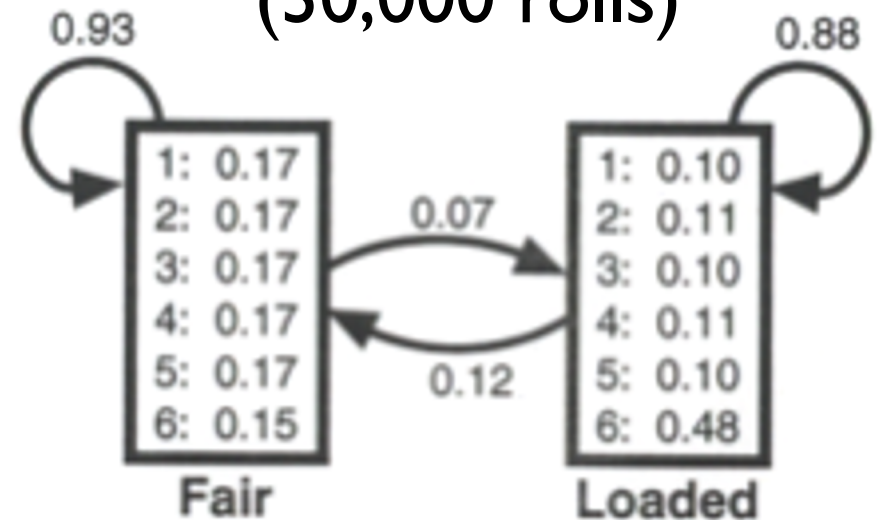
True Model



B-W Learned Model (300 rolls)



B-W Learned Model (30,000 rolls)



Log-odds (vs all F) per roll

True model 0.101 bits

300-roll est. 0.097 bits

30k-roll est. 0.100 bits

(NB: overestimated)

HMMs in Action: Pfam

<http://pfam.xfam.org>

Proteins fall into families, both across & within species

Ex: Globins, Zinc fingers, Leucine zippers, GPCRs, ...

Identifying family very useful: suggests function, etc.

So, search & alignment are both important

Q. Why not just use Blast/Smith-Waterman?

A. There is more info in *multiple* examples (e.g., psiBLAST)

One very successful approach: profile HMMs

```

Helix          AAAAAAAAAAAAAAAAAA  BBBB BBBB BBBB BBBB BBBBBB CCCCCCCCCCCC
HBA_HUMAN      -----VLSPADKTNVKA AWGKVGA--HAGEYGA EALERMFLSFPTTKTYFPHF
HBB_HUMAN      -----VHLTPEEKSAVTALWGKV---NVDEVGGEALGRLLVVYPWTQRFFESF
MYG_PHYCA      -----VLSEGEWQLVLHVWAKVEA--DVAGHGQDILIRLFKSHPETLEKFD RF
GLB3_CHITP     -----LSADQISTVQASFDKVKG-----DPVGILYAVFKADPSIMAKFTQF
GLB5_PETMA     PIVDTGSVAPLSAAEKT KIRSAWAPVYS--TYETSGVDILVKFFTSTPAAQE FFPKF
LGB2_LUPLU     -----GALTESQAALVKSSWEEFN A--NIPKHTHRFFILVLEIAPA AKDLFS-F
GLB1_GLYDI     -----GLSAAQRQVIAATWKDIAGADNGAGVGKDC LIKFLSAHPQMAAVFG-F
Consensus      Ls....  v a W kv . .    g . L.. f . P .    F F

```

```

Helix          DDDDDDDDEEEEEEEEEEEEEEEEEEEEEEEEEEE FFFFFFFF FFFF
HBA_HUMAN      -DLS-----HGSAQVKGHGKKVADALTNAVAHV---D--DMPNALSALS D LHAHKL-
HBB_HUMAN      GDLSTPD AVMGNPKVKAHGKKVLGAFSDGLAHL---D--NLKGT FATLSELHCDKL-
MYG_PHYCA      KHLKTEAEMKASEDLKKHGVTVLTALGAILKK---K-GHHEAELKPLAQSHATKH-
GLB3_CHITP     AG-KDLESIKGTAPFETHANRIVGFFSKIIGEL--P---NIEADVNTFVASHKPRG-
GLB5_PETMA     KGLTTADQLKKSADV RWHAE RIINAVNDAVASM--DDTEKMSMKLRDLSGKHAKSF-
LGB2_LUPLU     LK-GTSEVPQNNPELQA HAGKVF KLVYEAAIQ LQVTGVVVTDATLKNLGSVH VSKG-
GLB1_GLYDI     SG----AS---DPGVAALGAKVLAQIGVAVSHL--GDEGKMVAQMKAVGV RHKGYGN
Consensus      .  t    . . . v..Hg kv. a    a...l  d    . a l. l  H .

```

```

Helix          FFGGGGGGGGGGGGGGGGGGGGGG  HHHHHHHHHHHHHHHHHHHHHHHHHHHHHHH
HBA_HUMAN      -RVDPVNFKLLSHCLLVTLAAHLPAEFTPAVHASLDKFLASVSTVLTSKYR-----
HBB_HUMAN      -HVDPENFRLLGNVLVCVLAHHFGKEFTPPVQAAYQKVVAGVANALAHKYH-----
MYG_PHYCA      -KIPIKYLEFISEAIIHVLHSRHPGDFGADAQGAMNKALELFRKDIAAKYKELGYQG
GLB3_CHITP     --VTHDQLNNFRAGFVS YMKAHT--DFA-GAEAAWGATLD TFFGMIFSKM-----
GLB5_PETMA     -QVDPQYFKVLA AVIADTVAAG-----DAGFEKLMSMICILLRSAY-----
LGB2_LUPLU     --VADAHFPVVK EAILKTIKEVVGAKWSEELNSAWTIAYDELAIVIKKEMNDAA---
GLB1_GLYDI     KHIKAQYFEPLGASLLS AMEHRIGGKMNA AAKDAWAAAYADISGALISGLQS----
Consensus      v.    f  l . . . . .    f . aa. k. .    l sky

```

Alignment of 7 globins. A-H mark 8 alpha helices.
 Consensus line: upper case = 6/7, lower = 4/7, dot=3/7.
 Could we have a profile (aka weight matrix) w/ indels?

Profile Hmm Structure

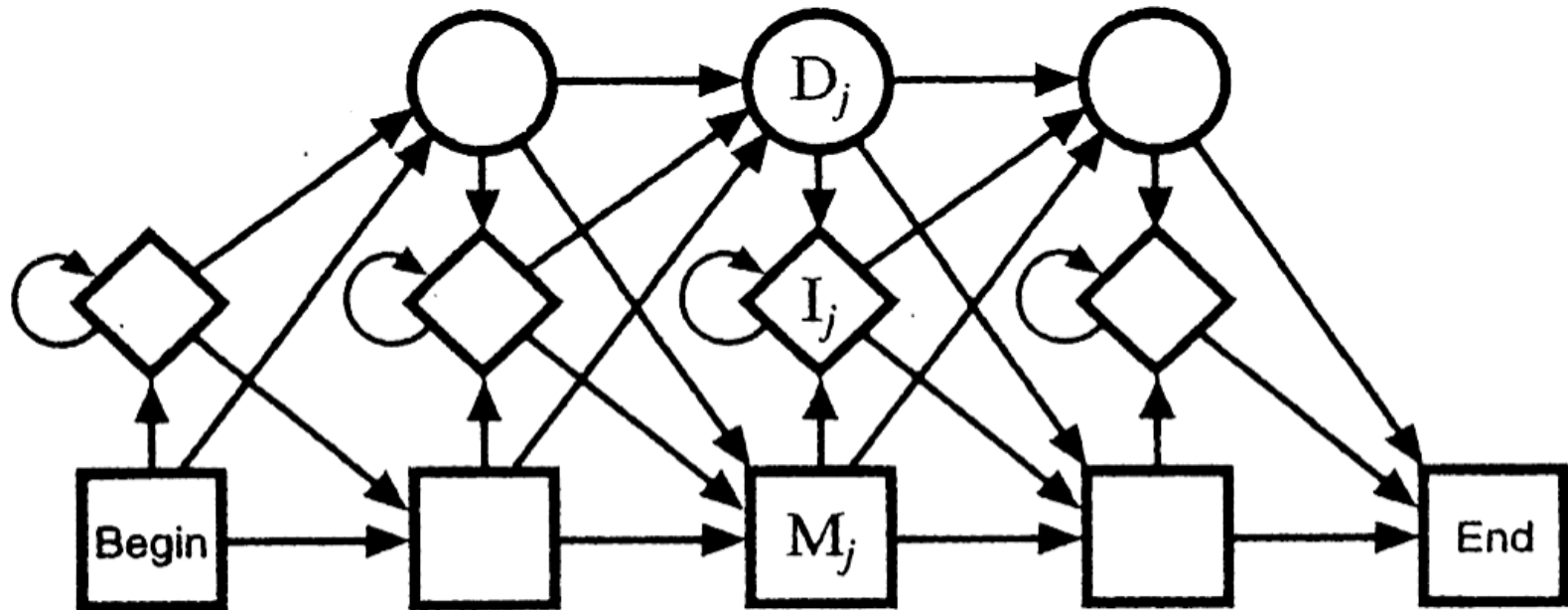


Figure 5.2 *The transition structure of a profile HMM.*

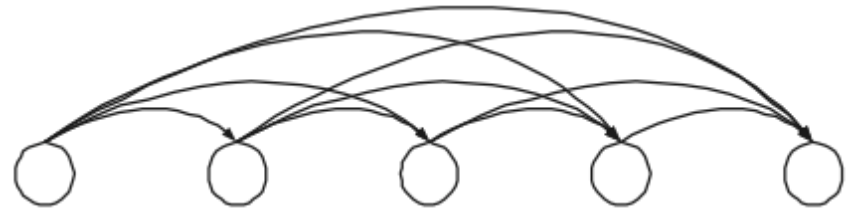
M_j : Match states (20 emission probabilities)

I_j : Insert states (Background emission probabilities)

D_j : Delete states (silent - no emission)

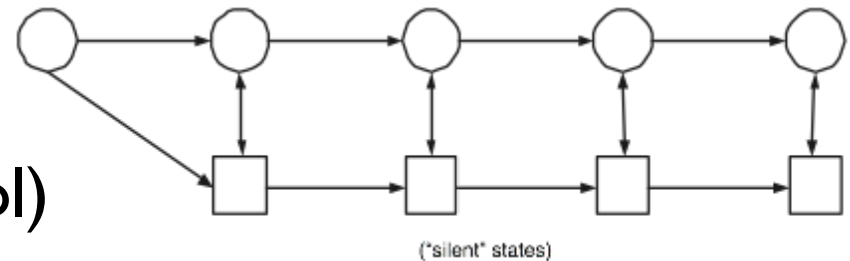
Silent States

Example: chain of states, can skip some



Problem: many parameters.

A solution: chain of “silent” states; fewer parameters (but less detailed control)



Algorithms: basically the same.

Using Profile HMM's

Search

Forward or Viterbi

Scoring

Log likelihood (length adjusted)

Log odds vs background

Z scores from either



next slides

Alignment

Viterbi

Likelihood vs Odds Scores

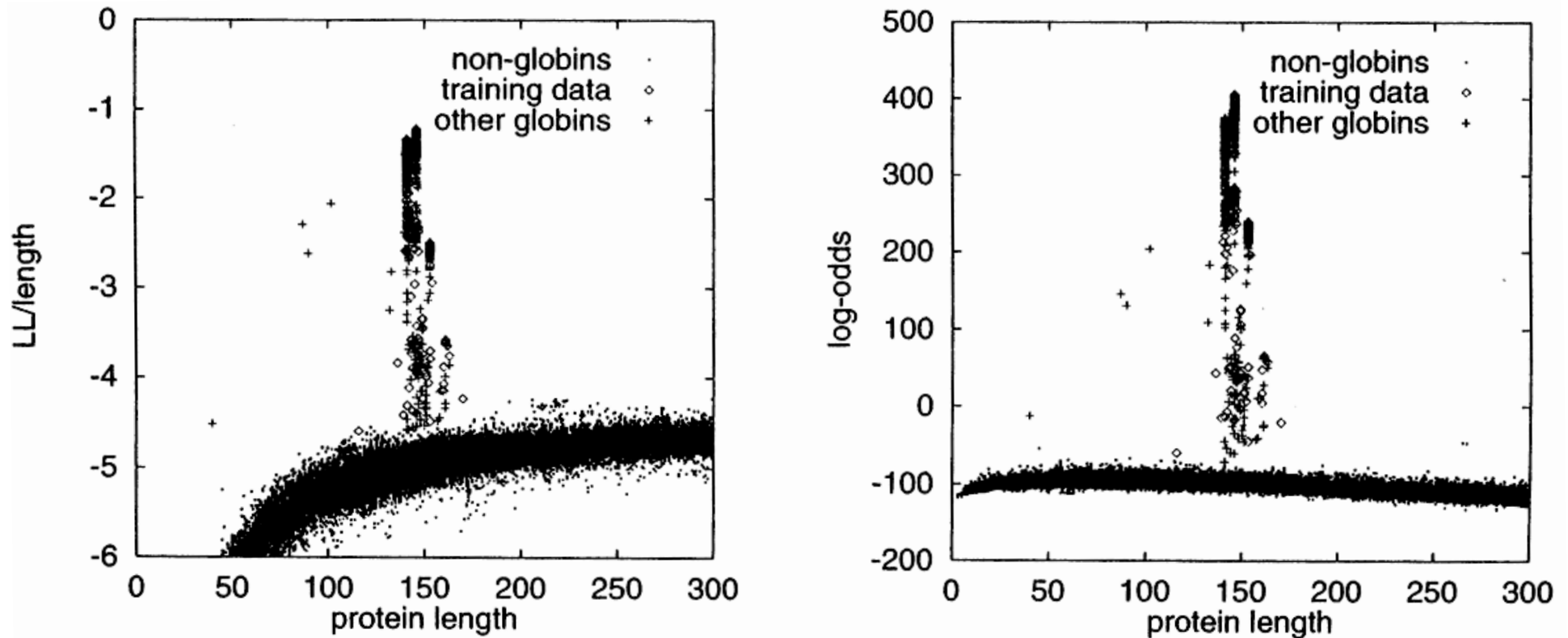


Figure 5.5 To the left the length-normalized LL score is shown as a function of sequence length. The right plot shows the same for the log-odds score.

Z-Scores

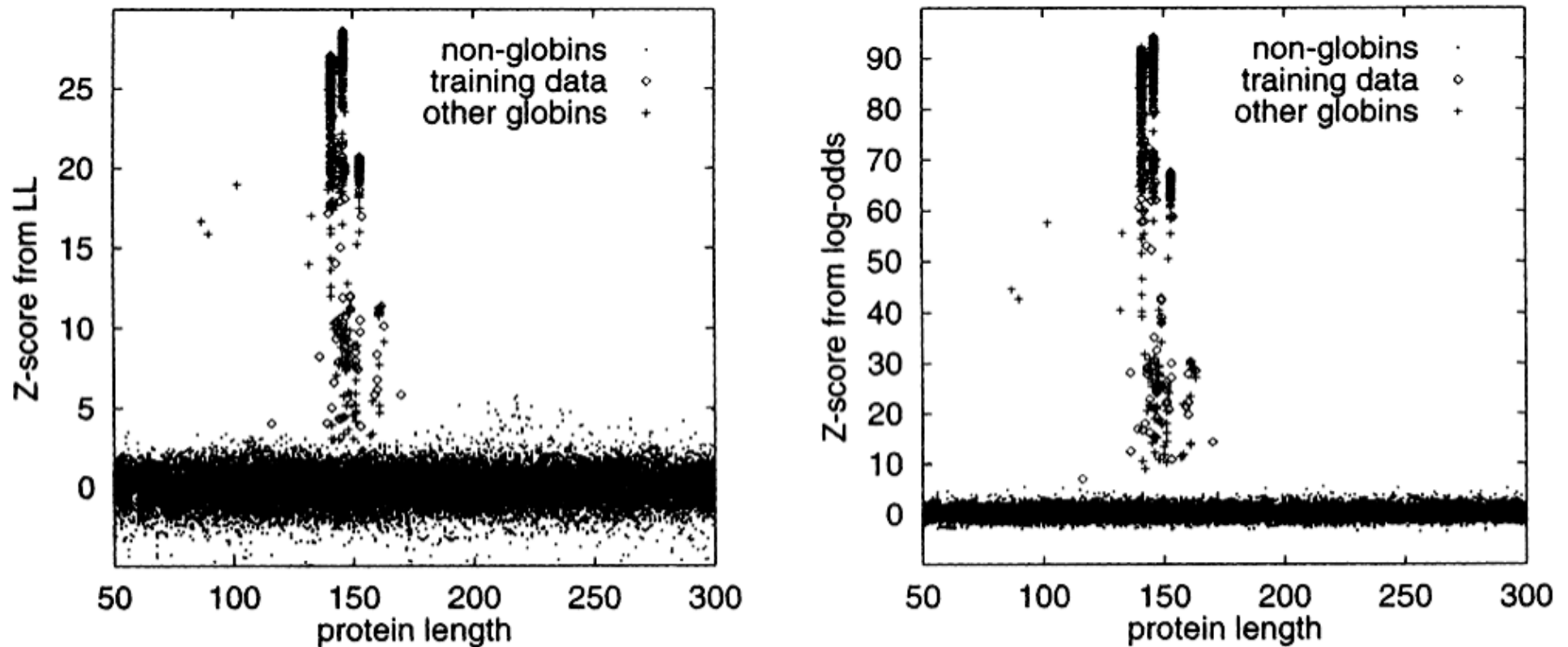


Figure 5.6 *The Z-score calculated from the LL scores (left) and the log-odds (right).*

Pfam Model Building


Hand-curated “seed” multiple alignments

Train profile HMM from seed alignment

Hand-chosen score threshold(s)

Automatic classification/alignment of all other protein sequences

% of human proteins containing ≥ 1 PFAM domain



Version	Date	#Families	Coverage
25.0	3/2011	12273	75
27.0	3/2013	14831	90
31.0	3/2017	16712	
32.0	9/2018	17929	
33.1	5/2020	18259	

Model-building refinements

Pseudocounts (with 20 aa's & few training seqs,
count = 0 common; adding $A > 0$ helps)

$$e_i(a) = \frac{C_{i,a} + A \cdot q_a}{\sum_a C_{i,a} + A}, \quad A \sim 20, \quad q_a = \text{background}$$

(~50 training sequences)

Pseudocount “mixtures”, e.g. separate
pseudocount vectors for various contexts
(hydrophobic regions, buried regions,...)

(~10-20 training sequences)

More refinements

Weighting: may need to down weight highly similar sequences to reflect phylogenetic or sampling biases, etc.

Match/insert assignment: Simple threshold, e.g. “> 50% gap \Rightarrow insert”, may be suboptimal. Can use forward-algorithm-like dynamic programming to compute max *a posteriori* assignment.

Numerical Issues

Products of many probabilities $\rightarrow 0$

For Viterbi: just add logs

For forward/backward: also work with logs, but you need sums of products, so need “log-of-sum-of-product-of-exp-of-logs”, e.g., by table/interpolation

Keep high precision and perhaps scale factor

Working with log-odds also helps.

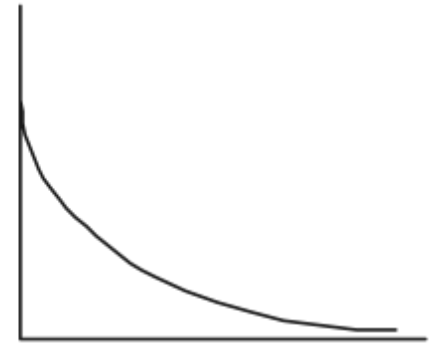
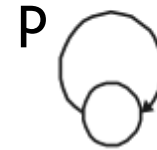
Model structure

Define it as well as you can.

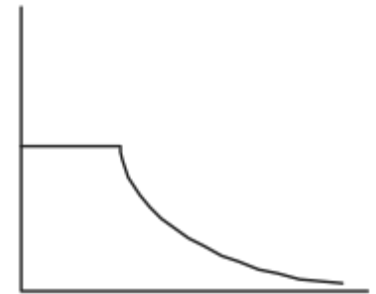
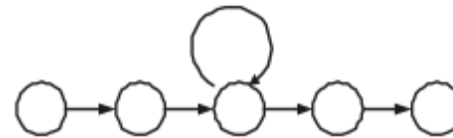
In principle, you can allow all transitions and hope to learn their probabilities from data, but it usually works poorly – too many local optima

Duration Modeling

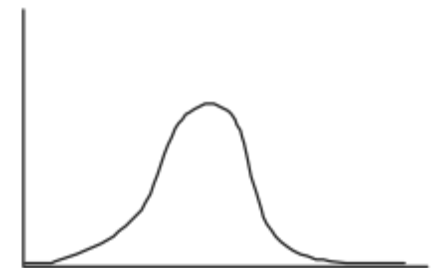
Self-loop duration:
geometric $p^n(1-p)$



min, then geometric



“negative binomial”



More general: possible (but slower)

HMM Summary

joint vs
conditional probs

Inference

- Viterbi – best single path (max of products)
- Forward – sum over all paths (sum of products)
- Backward – similar
- Posterior decoding

Model building

- Semi-supervised – typically fix architecture (e.g. profile HMM), then learn parameters
- Baum-Welch – training via EM and forward/backward (aka the forward/backward algorithm)
- Viterbi training – also “EM-like”, but Viterbi-based

HMM Summary (cont.)

Search:

Viterbi or forward

Scoring:

Odds ratio to background

Z-score

E-values, etc., too

Excellent tools available (HMMer, Pfam, ...)

Very widely used for bioseq analysis (& elsewhere)