## CSE P 527

## Markov Models and Hidden Markov Models


http://upload.wikimedia.org/wikipedia/commons/b/ba/Calico_cat

## Dosage Compensation and X -Inactivation

2 copies (mom/dad) of each chromosome I-23
Mostly, both copies of each gene are expressed
E.g., A B O blood group defined by 2 alleles of I gene

Women ( XX ) get double dose of X genes (vs XY )?
So, early in embryogenesis:

- One $X$ randomly inactivated in each cell
- Choice maintained in daughter cells ]

Calico: a major coat color gene is on $X$

## Reminder: Proteins "Read" DNA




## Down in the Groove

## Different

 patterns of hydrophobic methyls, potential H bonds, etc. at edges of different base pairs. They're accessible, esp. in major groove

Figure 7-7 Molecular Biology of the Cell 5/e (© Garland Science 2008)

## DNA Methylation

CpG - 2 adjacent nts, same strand (not Watson-Crick pair; "p" mnemonic for the phosphodiester bond of the DNA backbone)
C of CpG is often (70-80\%) methylated in mammals i.e., $\mathrm{CH}_{3}$ group added (both strands)

cytosine

## Same Pairing

 Methyl-C alters major groove profile (.: TF binding), but not basepairing,transcription or replication


Figure 7-7 Molecular Biology of the Cell 5/e ( © Garland Science 2008)

## DNA Methylation-Why

In vertebrates, it generally silences transcription
(Epigenetics) X-inactivation, imprinting, repression of mobile elements, cancers, aging, and developmental differentiation
E.g., if a stem cell divides, one daughter fated to be liver, other kidney, need to


(a) Turn off liver genes in kidney \& vice versa,
(b) Remember that through subsequent divisions

How? One way:
(a) Methylate genes, esp. promoters, to silence them
(b) After $\div$, DNA methyltransferases convert hemi- to fully-methylated (not trivial: deleting methyltransferase is embrionic-lethal in mice)
Major exception: promoters of "housekeeping" genes

## "CpG Islands"

Methyl-C mutates to $T$ relatively easily
Net: CpG is less common than expected genome-wide: $\mathrm{f}(\mathrm{CpG})<\mathrm{f}(\mathrm{C}) * \mathrm{f}(\mathrm{G})$

cytosine
BUT in some regions (e.g. active promoters), CpGs remain unmethylated, so $\mathrm{CpG} \rightarrow$ TpG less likely there: makes "CpG Islands"; often mark gene-rich regions

thymine

## CpG Islands

CpG Islands
More $C_{p G}$ than elsewhere (say, $\mathrm{CpG}^{\prime} / \mathrm{GpC}>50 \%$ )
More C \& G than elsewhere, too (say, C+G>50\%)
Typical length: few 100 to few 1000 bp
Questions
Is a short sequence (say, 200 bp ) a CpG island or not?
Given long sequence (say, $10-100 \mathrm{~kb}$ ), find CpG islands?

## Markov \& Hidden Markov Models

References (see also online reading page):
Eddy, "What is a hidden Markov model?" Nature Biotechnology, 22, \#IO (2004) I3I5-6.
Durbin, Eddy, Krogh and Mitchison, "Biological Sequence Analysis", Cambridge, 1998 (esp. chs 3, 5)
Rabiner, "A Tutorial on Hidden Markov Models and Selected Application in Speech Recognition," Proceedings of the IEEE, v 77 \#2,Feb 1989, 257-286

## Independence

A key issue: Previous models we've talked about assume independence of nucleotides in different positions - definitely unrealistic.

Markov models allow us to relax that assumption.

## Markov Chains

A sequence $x_{1}, x_{2}, \ldots$ of random variables is a $k$-th order Markov chain if, for all $i, i^{\text {th }}$ value is independent of all but the previous $k$ values:

$$
P\left(x_{i} \mid \underset{\mathrm{i}-1}{x_{1}, x_{2}, \ldots, x_{i-1}}\right)=P\left(x_{i} \mid \underset{\text { k typically < i-l }}{x_{i-k}, x_{i-k+1}, \ldots, x_{i-1}}\right)
$$

Example I: Uniform random ACGT
Example 2: Weight matrix model
Example 3: ACGT, but $\downarrow \operatorname{Pr}(G$ following C)

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## A Markov Model (Ist order)



States: A,C,G,T
Emissions: corresponding letter
Transitions: $a_{s t}=P\left(x_{i}=t \mid x_{i-1}=s\right) \longleftarrow$ Ist order

## A Markov Model (Ist order)



States: A,C,G,T
Emissions: corresponding letter
Transitions: $a_{s t}=P\left(x_{i}=t \mid x_{i-1}=s\right)$
Begin/End states

## Pr of emitting sequence $x$

$$
\begin{aligned}
& x \quad=x_{1} x_{2} \ldots x_{n}
\end{aligned}
$$

$$
\begin{aligned}
& =P\left(x_{1}\right) \cdot P\left(x_{2} \mid x_{1}\right) \cdots P\left(x_{n} \mid x_{n-1}, \ldots, x_{1}\right) \\
& \left.=P\left(x_{1}\right) \cdot P\left(x_{2} \mid x_{1}\right) \cdots P\left(x_{n} \mid x_{n-1}\right)\right\rangle_{\mathfrak{j} \backslash s t} \\
& =P\left(x_{1}\right) \prod_{i=1}^{n-1} a_{x_{i}, x_{i+1}} \\
& =\prod_{i=0}^{n-1} a_{x_{i}, x_{i+1}} \quad \text { (with Begin state) }
\end{aligned}
$$

## Training

Max likelihood estimates for transition probabilities are just the frequencies of transitions when emitting the training sequences
E.g., from 48 CpG islands in 60 kbp :

| $\mathbf{+}$ | $\mathbf{A}$ | $\mathbf{C}$ | $\mathbf{G}$ | $\mathbf{T}$ |  |  | $\mathbf{A}$ | $\mathbf{C}$ | $\mathbf{G}$ | $\mathbf{T}$ |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| $\mathbf{A}$ | 0.180 | 0.274 | 0.426 | 0.120 |  | $\mathbf{A}$ | 0.300 | 0.205 | 0.285 | 0.210 |
| $\mathbf{C}$ | 0.171 | 0.368 | 0.274 | 0.188 |  | $\mathbf{C}$ | 0.322 | 0.298 | 0.078 | 0.302 |
| $\mathbf{G}$ | 0.161 | 0.339 | 0.375 | 0.125 |  | $\mathbf{G}$ | 0.248 | 0.246 | 0.298 | 0.208 |
| $\mathbf{T}$ | 0.079 | 0.355 | 0.384 | 0.182 |  | $\mathbf{T}$ |  | 0.177 | 0.239 | 0.292 |
|  | 0.292 |  |  |  |  |  |  |  |  |  |

## Discrimination/Classification

Log likelihood ratio of CpG model vs background model


## CpG Island Scores



Figure 3.2 Histogram of length-normalized scores.

## Questions

QI: Given a short sequence, is it more likely from feature model or background model? Above
Q2: Given a long sequence, where are the features in it (if any)

Approach I: score 100 bp (e.g.) windows
Pro: simple
Con: arbitrary, fixed length, inflexible
Approach 2: combine +/- models.

## Combined Model



Emphasis is "Which (hidden) state?" not "Which model?"

## Hidden Markov Models

 (HMMs; Claude Shannon, I948)States:
Paths:
Transitions:
Emissions:
Observed data: emission sequence Hidden data:
$1,2,3, \ldots$
$e_{k}(b)=P\left(x_{i}=b \mid \pi_{i}=k\right)$ state/transition sequence
sequences of states $\pi=\left(\pi_{1}, \pi_{2}, \ldots\right)$
$a_{k, l}=P\left(\pi_{i}=l \mid \pi_{i-1}=k\right)$

## The Occasionally Dishonest Casino

1 fair die, 1 "loaded" die, occasionally swapped


| Rolls |  |
| :---: | :---: |
| Die | FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFLLLLLLLLLLLLLLL |
| Viterbi | FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFLLLLLLLLLLLL |
| Rolls | 651166453132651245636664631636663162326455236266666625151631 |
| Die | LLLLLLFFFFFFFFFFFFLLLLLLLLLLLLLLLLFFFLLLLLLLLLLLLLLFFFFFFFFF |
| Viterbi | LLLLLLFFFFFFFFFFFFLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLFFFFFFFF |
| Rolls | 222555441666566563564324364131513465146353411126414626253356 |
| Die | FFFFFFFFLLLLLLLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFLL |
| Viterbi | FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFL |
| Rolls | 366163666466232534413661661163252562462255265252266435353336 |
| Die | LLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF |
| Viterbi | LLLLLLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF |
| Rolls | 233121625364414432335163243633665562466662632666612355245242 |
| Die | FFFFFFFFFFFFFFFFFFFFFFFFFFFLLLLLLLLLLLLLLLLLLLLLLFFFFFFFFFFF |
| Viterbi | FLLLLLLLLLLLLLLLLLLLFFFFFFFF |

## Figure 3.5

Rolls: Visible data-300 rolls of a die as described above.
Die: Hidden data-which die was actually used for that roll ( $F=$ fair, $L=$ loaded).
Viterbi: the prediction by the Viterbi algorithm is shown.

## Inferring hidden stuff

Joint probability of a given path $\pi$ \& emission sequence $x$ :

$$
P(x, \pi)=a_{0, \pi_{1}} \prod_{i=1}^{n} e_{\pi_{i}}\left(x_{i}\right) \cdot a_{\pi_{i}, \pi_{i+1}}
$$

But $\pi$ is hidden; what to do? Some alternatives:
Most probable single path

$$
\pi^{*}=\arg \max _{\pi} P(x, \pi)
$$

Sequence of most probable states

$$
\hat{\pi}_{i}=\arg \max _{k} P\left(\pi_{i}=k \mid x\right)
$$

Etc.

# The Viterbi Algorithm: <br> <br> The most probable path 

 <br> <br> The most probable path}

Viterbi finds: $\pi^{*}=\arg \max _{\pi} P(x, \pi)$
Possibly there are $10^{99}$ paths of prob $10-99$ (If so, non-Viterbi approaches may be preferable.)
More commonly, one path (+ slight variants) dominate others; Viterbi finds that

Key problem: exponentially many paths $\pi$

## Unrolling an HMM



Conceptually, sometimes convenient Note exponentially many paths

## Viterbi

$v_{l}(i)=$ probability of the most probable path emitting $x_{1}, x_{2}, \ldots, x_{i}$ and ending in state $\ell$

Initialize:


## HMM Casino Example


(Excel spreadsheet on web; download \& play...)

## HMM Casino Example


(Excel spreadsheet on web; download \& play...)

## Viterbi Traceback

Above finds probability of best path
To find the path itself, trace backward to the state $k$ attaining the max at each stage


| Rolls |  |
| :---: | :---: |
| Die | FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFLLLLLLLLLLLLLLL |
| Viterbi | FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFLLLLLLLLLLLL |
| Rolls | 651166453132651245636664631636663162326455236266666625151631 |
| Die | LLLLLLFFFFFFFFFFFFLLLLLLLLLLLLLLLLFFFLLLLLLLLLLLLLLFFFFFFFFF |
| Viterbi | LLLLLLFFFFFFFFFFFFLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLFFFFFFFF |
| Rolls | 222555441666566563564324364131513465146353411126414626253356 |
| Die | FFFFFFFFLLLLLLLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFLL |
| Viterbi | FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFL |
| Rolls | 366163666466232534413661661163252562462255265252266435353336 |
| Die | LLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF |
| Viterbi | LLLLLLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF |
| Rolls | 233121625364414432335163243633665562466662632666612355245242 |
| Die | FFFFFFFFFFFFFFFFFFFFFFFFFFFLLLLLLLLLLLLLLLLLLLLLLFFFFFFFFFFF |
| Viterbi | FLLLLLLLLLLLLLLLLLLLFFFFFFFF |

## Figure 3.5

Rolls: Visible data-300 rolls of a die as described above. Die: Hidden data-which die was actually used for that roll ( $F=$ fair, $L=$ loaded). Viterbi: the prediction by the Viterbi algorithm is shown.

## Most probable path!= sequence of most probable states

Another example, based on casino dice again:
Suppose p(fair $\leftrightarrow$ loaded) transitions are 10-99 and roll sequence is $1 I\|I\| 66 \ldots 666$; then fair state is more likely all through I's \& well into the run of 6's, but eventually loaded wins, and the improbable $F \rightarrow L$ transitions make Viterbi $=$ all L .

F


* $=$ max prob
$\square=$ Viterbi


## Is Viterbi "best"?

Viterbi finds $\pi^{*}=\arg \max _{\pi} P(x, \pi)$


Most probable (Viterbi) path goes through 5, but most probable state at 2 nd step is 6
(I.e., Viterbi is not the only interesting answer.)

## An HMM (unrolled)

States

## $\downarrow$



Emissions/sequence positions $\longrightarrow$

## Viterbi: best path to each

 state

Viterbi score:

$$
v_{l}(i+1)=e_{l}\left(x_{i+1}\right) \cdot \max _{k}\left(v_{k}(i) a_{k, l}\right)
$$

> Viterbi path

## The Forward Algorithm

For each state/time, want total probability of all paths leading to it, with given emissions


## The Backward Algorithm

Similar: for each state/time, want total probability of all paths from it, with given
emissions, conditional on that state.


## In state $k$ at step $i$ ?

$$
\begin{aligned}
& P\left(x, \pi_{i}=k\right) \\
& \quad=P\left(x_{1}, \ldots, x_{i}, \pi_{i}=k\right) \cdot P\left(x_{i+1}, \ldots, x_{n} \mid x_{1}, \ldots, x_{i}, \pi_{i}=k\right) \\
& \quad=P\left(x_{1}, \ldots, x_{i}, \pi_{i}=k\right) \cdot P\left(x_{i+1}, \ldots, x_{n} \mid \pi_{i}=k\right) \\
& \quad=f_{k}(i) \cdot b_{k}(i) \\
& P\left(\pi_{i}=k \mid x\right)=\frac{P\left(x, \pi_{i}=k\right)}{P(x)}=\frac{f_{k}(i) \cdot b_{k}(i)}{P(x)}
\end{aligned}
$$

## Posterior Decoding,

Alternative 1: what's the most likely state at step i?

$$
\hat{\pi}_{i}=\arg \max _{k} P\left(\pi_{i}=k \mid x\right)
$$

Note: the sequence of most likely states $\boldsymbol{\neq}$ the most likely sequence of states. May not even be legal!


## The Occasionally Dishonest Casino

1 fair die, 1 "loaded" die, occasionally swapped


| Rolls |  |
| :---: | :---: |
| Die | FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFLLLLLLLLLLLLLLL |
| Viterbi | FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFLLLLLLLLLLLL |
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| Die | FFFFFFFFLLLLLLLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFLL |
| Viterbi | FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFL |
| Rolls | 366163666466232534413661661163252562462255265252266435353336 |
| Die | LLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF |
| Viterbi | LLLLLLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF |
| Rolls | 233121625364414432335163243633665562466662632666612355245242 |
| Die | FFFFFFFFFFFFFFFFFFFFFFFFFFFLLLLLLLLLLLLLLLLLLLLLLFFFFFFFFFFF |
| iterbi | FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFLLLLLLLLLLLLLLLLLLLFFFFFFFFF |

## Figure 3.5

Rolls: Visible data-300 rolls of a die as described above. Die: Hidden data-which die was actually used for that roll ( $F=$ fair, $L=$ loaded). Viterbi: the prediction by the Viterbi algorithm is shown.

## Posterior Decoding



Figure 3.6 The posterior probability of being in the state corresponding to the fair die in the casino example. The $x$ axis shows the number of the roll. The shaded areas show when the roll was generated by the loaded die.

## Posterior Decoding, II

Alternative 1: what's most likely state at step $i$ ?

$$
\hat{\pi}_{i}=\arg \max _{k} P\left(\pi_{i}=k \mid x\right)
$$

Alternative 2: given some function $g(k)$ on states, what's its expectation. E.g., what's probability of " + " model in CpG HMM ( $g(k)=1$ iff $k$ is " + " state)?

$$
G(i \mid x)=\sum_{k} P\left(\pi_{i}=k \mid x\right) \cdot g(k)
$$

## CpG Islands again

Data: 4 I human sequences, totaling 60 kbp , including 48 CpG islands of about Ikbp each

Viterbi:
Found 46 of 48
plus I2I "false positives"
Posterior Decoding:
same 2 false negatives
plus 236 false positives

Post-process:
46/48
67 false pos

46/48
83 false pos

## Training

Given model topology \& training sequences, learn transition and emission probabilities

If $\pi$ known, then MLE is just frequency observed in training data

$$
\begin{array}{ll}
a_{k, l} & =\frac{\text { count of } k \rightarrow l \text { transitions }}{\text { count of } k \rightarrow \text { anywhere transitions }} \\
e_{k}(b) & =\cdots
\end{array}
$$

If $\pi$ hidden, then use EM:
given $\theta$, estimate $\pi$; given $\pi$ estimate $\theta$; repeat $\}^{2}$ ways

## Viterbi Training

given $\theta$, estimate $\pi$; given $\pi$ estimate $\theta$; repeat
Make initial estimates of parameters $\theta$
Find Viterbi path $\pi$ for each training sequence
Count transitions/emissions on those paths, getting new $\theta$
Repeat
Not rigorously optimizing desired likelihood, but still useful \& commonly used.
(Arguably good if you're doing Viterbi decoding.)

## Baum-Welch Training

EM: given $\theta$, estimate $\pi$ ensemble; then re-estimate $\theta$

$$
\begin{aligned}
& P\left(\pi_{i}=k, \pi_{i+1}=l \mid x, \theta\right) \\
& \quad=\frac{f_{k}(i \mid \theta) a_{k, l} e_{l}\left(x_{i+1}\right) b_{l}(i+1 \mid \theta)}{P(x \mid \theta)}
\end{aligned}
$$

Estimated \# of $k \rightarrow l$ transitions $\hat{A}_{k, l}$ on set of seqs $\times \mathrm{j}$

$$
=\sum_{\text {training seqs } x^{j}} \sum_{i} P\left(\pi_{i}=k, \pi_{i+1}=l \mid x^{j}, \theta\right)
$$

New estimate $\hat{a}_{k, l}=\frac{\hat{A}_{k, l}}{\sum_{l} \hat{A}_{k, l}}$
Emissions: similar

## True Model



Log-odds (vs all F) per roll True model 0.IOI bits 300-roll est. 0.097 bits 30k-roll est. 0.100 bits (NB: overestimated)

B-W Learned Model (300 rolls)

B-W Learned Model


From DEKM 50

## HMMs in Action: Pfam http://pfam.xfam.org

Proteins fall into families, both across \& within species
Ex: Globins, Zinc fingers, Leucine zippers, GPCRs, ...
Identifying family very useful: suggests function, etc.
So, search \& alignment are both important
Q. Why not just use Blast/Smith-Waterman?
A. There is more info in multiple examples (e.g., psiBLAST)

One very successful approach: profile HMMs

HBA_HUMAN HBB_HUMAN MYG_PHYCA GLB3_CHITP GLB5_PETMA LGB2_LUPLU GLB1_GLYDI Consensus

Helix
HBA_HUMAN HBB_HUMAN MYG_PHYCA GLB3_CHITP GLB5_PETMA LGB2_LUPLU GLB1_GLYDI Consensus

## Helix

HBA_HUMAN
HBB_HUMAN MYG_PHYCA GLB3_CHITP GLB5_PETMA LGB2_LUPLU GLB1_GLYDI Consensus

## FFGGGGGGGGGGGGGGGGGGG

FFFFFFFFFFFF
-DLS-----HGSAQVKGHGKKVADALTNAVAHV---D--DMPNALSALSDLHAHKL-GDLSTPDAVMGNPKVKAHGKKVLGAFSDGLAHL---D--NLKGTFATLSELHCDKL-KHLKTEAEMKASEDLKKHGVTVLTALGAILKK----K-GHHEAELKPLAQSHATKH-AG-KDLESIKGTAPFETHANRIVGFFSKIIGEL--P---NIEADVNTFVASHKPRG-K̈GLTTADQLKKSADVRWHAERIINAVNDAVASM--DDTEKMSMKLRDLSGKHAKSF-LK-GTSEVPQNNPELQAHAGKVFKLVYEAAIQLQVTGVVVTDATLKNLGSVHVSKG-SG----AS---DPGVAALGAKVLAQIGVAVSHL--GDEGKMVAQMKAVGVRHKGYGN t .. . v..Hg kv. a a...l d . a 1. 1 H



## Alignment of 7 globins. A-H mark 8 alpha helices. Consensus line: upper case $=6 / 7$, lower $=4 / 7$, dot $=3 / 7$.

 Could we have a profile (aka weight matrix) w/ indels?
## Profile Hmm Structure



Figure 5.2 The transition structure of a profile HMM.
$\mathrm{Mj}: \quad$ Match states ( 20 emission probabilities)
l : $\quad$ Insert states (Background emission probabilities)
D : Delete states (silent - no emission)

## Silent States

Example: chain of states, can skip some


Problem: many parameters.
A solution: chain of "silent" states; fewer parameters (but less detailed control)


Algorithms: basically the same.

## Using Profile HMM's

## Search

Forward or Viterbi
Scoring
Log likelihood (length adjusted)
Log odds vs background
$Z$ scores from either


Alignment
Viterbi

## Likelihood vs Odds Scores




Figure 5.5 To the left the length-normalized LL score is shown as a function of sequence length. The right plot shows the same for the log-odds score.

## Z-Scores



Figure 5.6 The Z-score calculated from the LL scores (left) and the log-odds (right).

## Pfam Model Building

Hand-curated "seed" multiple alignments
Train profile HMM from seed alignment
Hand-chosen score threshold(s)
Automatic classification/alignment of all other protein sequences
Pfam 25.0 (March 201I, 12273 families; covers $\approx 75 \%$ of human proteins)
Pfam 27.0 (March 2013, I483I families; $\approx 90 \%$ )
Pfam 31.0 (March 2017, 16712 families)

## Model-building refinements

Pseudocounts (count $=0$ common when training with 20 aa's)

$$
e_{i}(a)=\frac{C_{i, a}+A \cdot q_{a}}{\sum_{a} C_{i, a}+A}, \quad A \sim 20, q_{a}=\text { background }
$$

(~50 training sequences)
Pseudocount "mixtures", e.g. separate pseudocount vectors for various contexts
(hydrophobic regions, buried regions,...)
(~10-20 training sequences)

## More refinements

Weighting: may need to down weight highly similar sequences to reflect phylogenetic or sampling biases, etc.
Match/insert assignment: Simple threshold, e.g. " $>50 \%$ gap $\Rightarrow$ insert", may be suboptimal. Can use forward-algorithm-like dynamic programming to compute max a posteriori assignment.

## Numerical Issues

Products of many probabilities $\rightarrow 0$
For Viterbi: just add logs
For forward/backward: also work with logs, but you need sums of products, so need "log-of-sum-of-product-of-exp-of-logs", e.g., by table/interpolation

Keep high precision and perhaps scale factor
Working with log-odds also helps.

## Model structure

Define it as well as you can.
In principle, you can allow all transitions and hope to learn their probabilities from data, but it usually works poorly - too many local optima

# Duration Modeling 

Self-loop duration: geometric $\mathrm{p}^{\mathrm{n}}(\mathrm{I}-\mathrm{p})$

min, then geometric

"negative binomial"


More general: possible (but slower)

## HMM Summary

Inference
Viterbi - best single path
Forward - sum over all paths
(max of products)
(sum of products)
Backward - similar
Posterior decoding
Model building
Semi-supervised - typically fix architecture (e.g. profile HMM), then learn parameters
Baum-Welch - training via EM and forward/backward (aka the forward/backward algorithm)
Viterbi training - also "EM-like", but Viterbi-based

## HMM Summary (cont.)

Search:
Viterbi or forward
Scoring:
Odds ratio to background
Z-score
E-values, etc., too
Excellent tools available (SAM, HMMer, Pfam, ...)
A very widely used tool for biosequence analysis

