# Overview of Single Cell Omics

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## Overview of Single Cell Omics

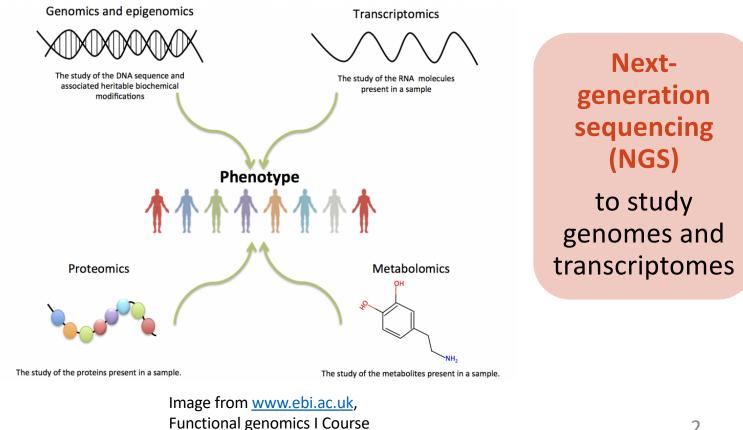
**Clinical Chemistry** 65:8 972-985 (2019)



### **Single-Cell Genomics**

Carmela Paolillo,<sup>1</sup> Eric Londin,<sup>2</sup> and Paolo Fortina<sup>3,4\*</sup>

### 1. Introduction: Next-generation sequencing



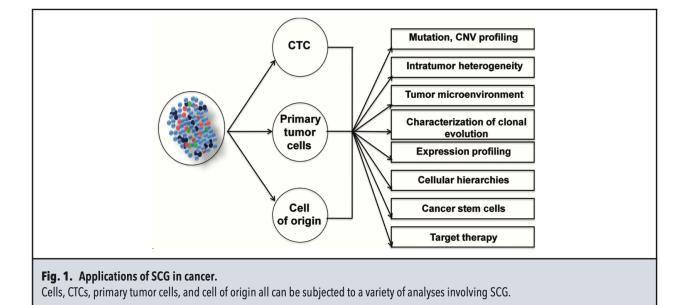
### 1. Introduction: Limitations of bulk approaches

### "Bulk" approaches

- Combining thousands of cells from a single tissue
- Treating them as a single unit
- Ignoring the role of each individual cell
- We now appreciate the **heterogeneous nature of various tissues** 
  - different cell populations may have vastly different transcriptomes and different contributions to cellular processes

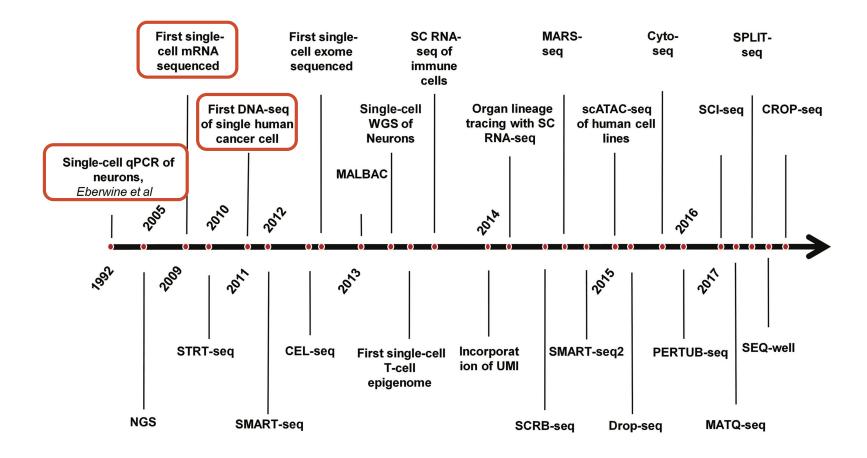
### 1. Introduction: Single-Cell Omics

Development of advanced methods to capture single cells and amplify genome-wide DNA and RNA -> Single-Cell Genomics (SCG)



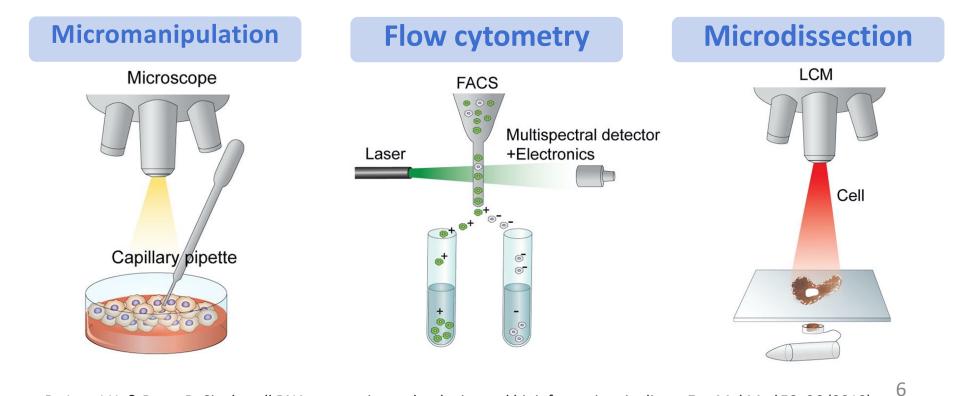
## SCG aims to provide a new understanding of genetics and transcriptomics at the single-cell level

### 1. Introduction: History of single-cell genomics



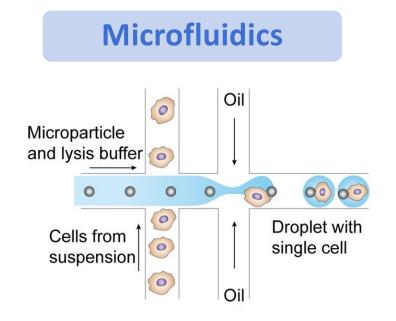
### 2.1 SCG Technology: Isolation of Single Cells

### 1<sup>st</sup> step = isolation of an intact single cell



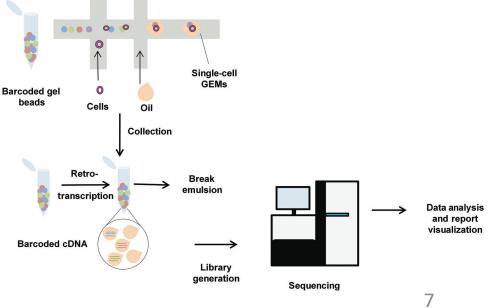
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### 2.1 SCG Technology: Isolation of Single Cells

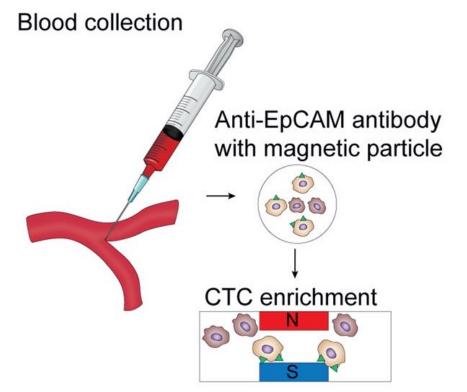


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# Chromium Single Cell 3' Solution (10x Genomics)



### 2.1 SCG Technology: Isolation of Single Cells



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Clinical applications of SCG involves the isolation of single cells from body fluids

CellSearch semiautomated cell-isolation system

Enables analysis of circulating tumor cells (CTCs)

## 2.2 SCG Technology: Amplification

### 1<sup>st</sup> step = isolation of an intact single cell

For sequencing we need enough genetic material to be analyzed **So, amplification is a crucial step** 

### 2<sup>nd</sup> step = amplification (WTA or WGA)

Amplification remains a major challenge for SCG Goal = minimize artifacts without loss in sensitivity and specificity

## 2.2 SCG Technology: Whole-Transcriptome Amplification (WTA)

### 1. Full-length methods

- Full representation of poly Acontaining transcripts
- Examples:
  - SMART-seqQuartz-Seq

- 2. Tag-based methods
  - Enable sequencing of either the 5' or 3' end
  - Allow strand specificity
  - CEL-seq is one of the most common

#### More recent protocols:

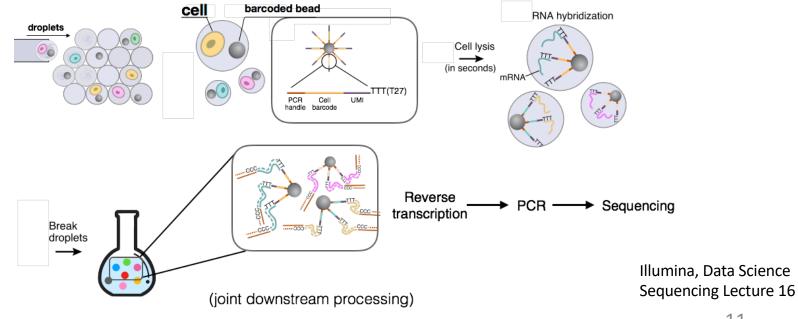
Label cells by using molecular barcodes

#### **Examples:**

- CytoSeq (Illumina) (oligonucleotide beads)
- inDrop and Drop-seq (Illumina) (droplets using a microfluidic device )
- SPLiT-seq (does not require encapsulating a single cell in droplets or microwells)

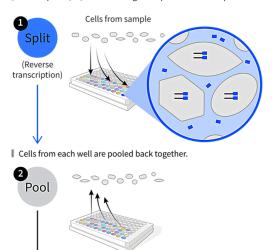
## 2.2 SCG Technology: Whole-Transcriptome Amplification (WTA)

#### **Drop-Seq pipeline**



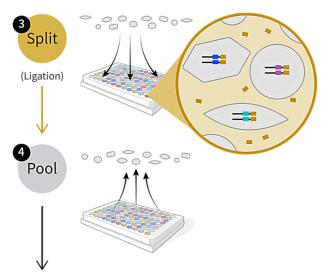
## 2.2 SCG Technology: Whole-Transcriptome **Amplification (WTA)**

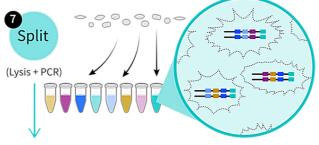
In the first round of barcoding, fixed cell samples are distributed into 48 wells, and cDNA is generated with an in-cell reverse transcription (RT) reaction using well-specific barcoded primers.



#### **SPLiT-seq pipeline**

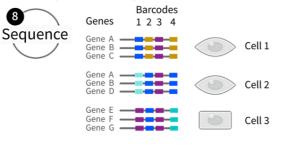
Cells are then distributed into 96 wells, and an in-cell ligation reaction appends a second well-specific barcode to the cDNA.





After sequencing, each single cell transcriptome is assembled by combining reads containing the same four-barcode combination.

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**Split Biosciences** 12

## 2.2 SCG Technology: Whole-Genome Amplification (WGA)

### **1. PCR amplification**

- First WGA method developed for SC
- Degenerate oligonucleotide primers (DOP- PCR) are used

#### **Disadvantages:**

- Amplification bias
- Variability in efficiency
- High error rate owing to the thermostable polymerase

#### 2. Isothermal Amplification (MDA)

- Most widely used protocol for WGA
- Has lower error rate
- Better genome coverage and reduced false-positive rates

### **3. Hybrid Methods**

 Combines advantages of the first 2 and reduces associated biases

Examples:

- PicoPLEX
- MALBAC

#### **Disadvantages:**

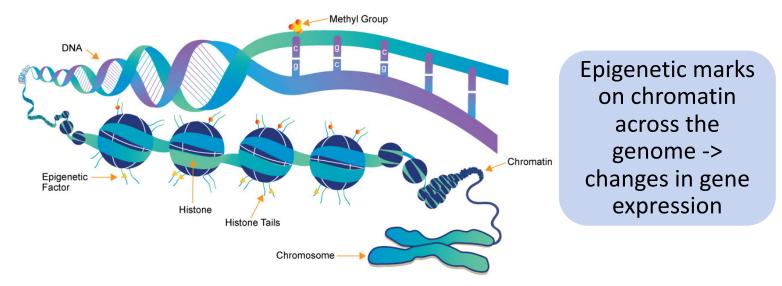
 Reduced uniformity and a high rate of allelic dropout

## 2.2 SCG Technology: WTA and WGA

Particular approach used should be based on

- specific application
- relative scarcity or abundance of the single cell to be isolated
- ease of DNA and/or RNA isolation from the target cell
- required depth of sequencing coverage

## 3. Applications of SCG: Epigenomics



**Epigenomics =** genome-wide studies of

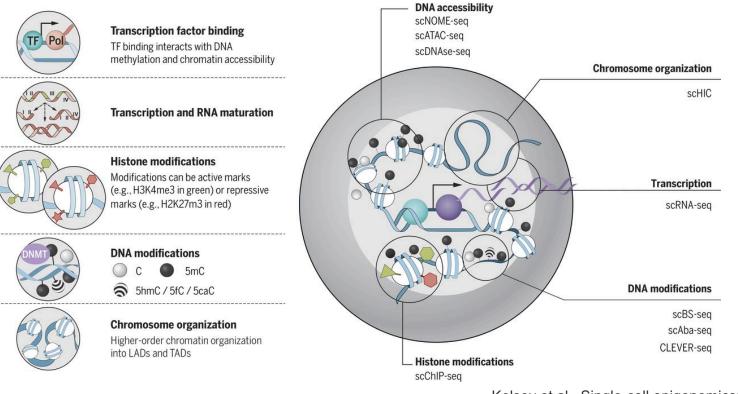
- DNA accessibility
- higher-order chromosome organization
  - DNA/protein modifications

https://www.whatisepigenetics.com/fundamentals/

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#### How can SCG help?

#### Information about chromatin modifications and their regulatory effects can be analyzed at single-cell resolution



Kelsey et al., Single-cell epigenomics: Recording the past and predicting the future 16

## 3. Applications of SGC: Cancer Genomics

#### **Cancer = high inter-tumor and intratumor variability**

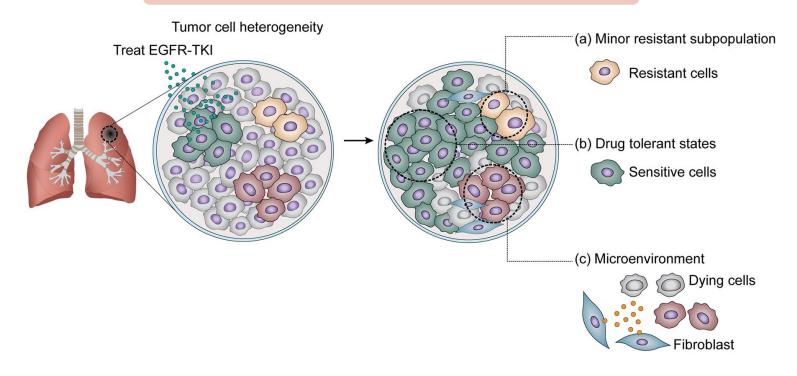
The ability to molecularly phenotype every clone or population within a tumor -> key to treatment

#### How can SCG help?

Monitoring disease progression and predict treatment resistance Isolation and characterization of CTCs Tumor heterogeneity, tumor microenvironment, and clonal evolution Identification of cancer cells of origin, rare cell subpopulations, tumor-tissue hierarchies

## 3. Applications of SGC: Cancer Genomics

#### **Drug resistant clone identification**



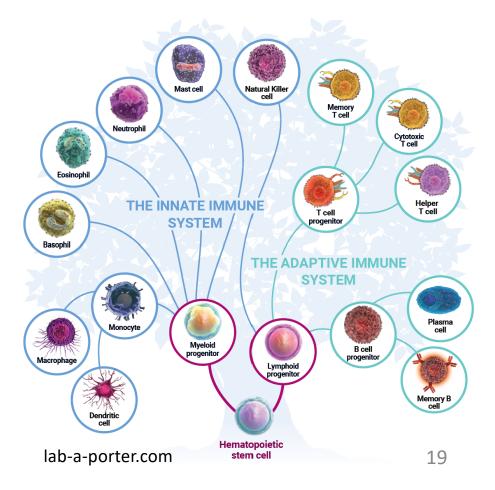
Hwang et al., Single-cell RNA sequencing technologies and bioinformatics pipelines

## 3. Applications of SGC: Immunology

Immune system = high degree
of genetic diversity representing
multiple different cell types

#### How can SCG help?

Cellular complexity of the immune system Characterize/classify the different cell types Immunotherapy Neurodegenerative disease



## 3. Applications of SGC

### Microbiology

• Evaluating microbiome biodiversity

### **Prenatal Screening**

• Detecting clinically significant genetic alterations with noninvasive prenatal diagnostic procedures

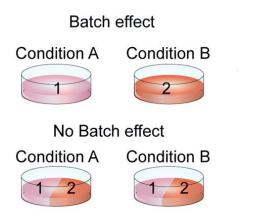
### **Cell Lineage and Differential Markers**

- Inferring lineage information from the early developmental stage
- Identifying novel differential markers

## 4. Bioinformatics for SGC

#### Challenges in organizing, archiving, and mining the data

Sparsity (dropouts) and high rates of technical artifacts (low-quality sequence data or batch effects)

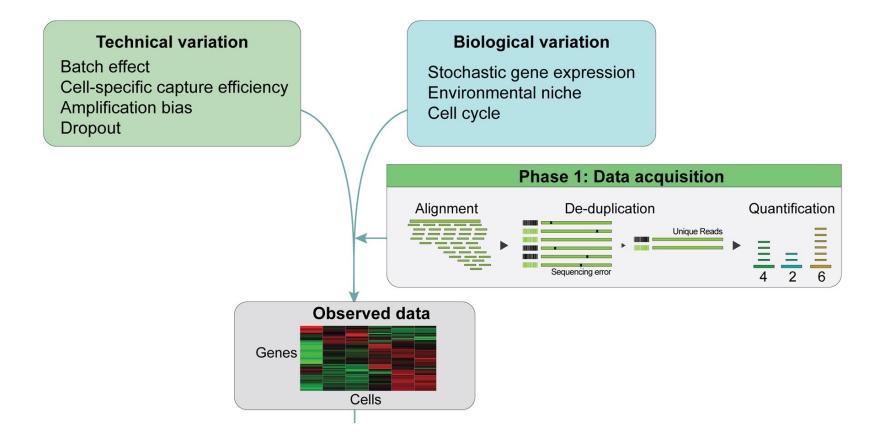


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## 4. Bioinformatics for SGC

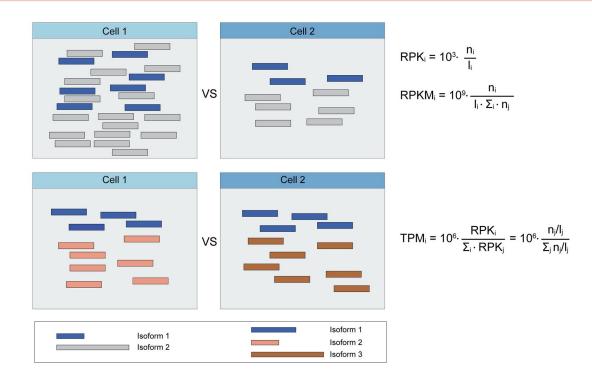
### **Basic approach for analyzing SCG data:**

- 1. Quality control and mapping of the sequence reads to the genome (as done with bulk sequencing)
- 2. Counting the number of genes
- 3. Normalization of the molecular counts
- 4. Clustering cells by the expressed genes

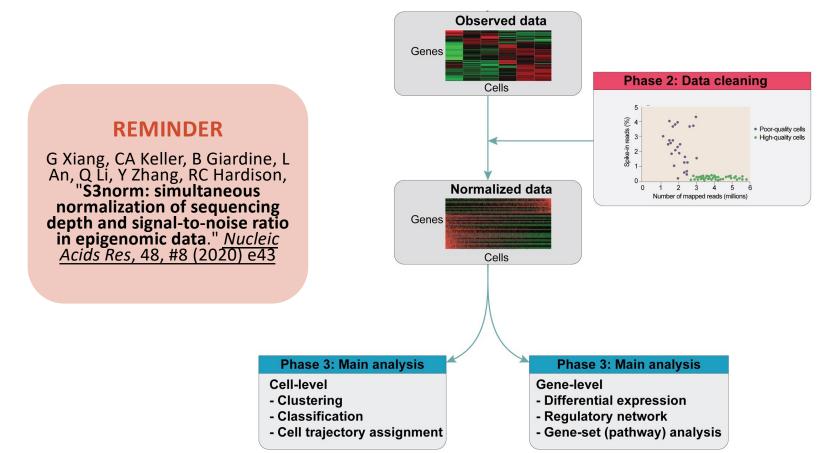


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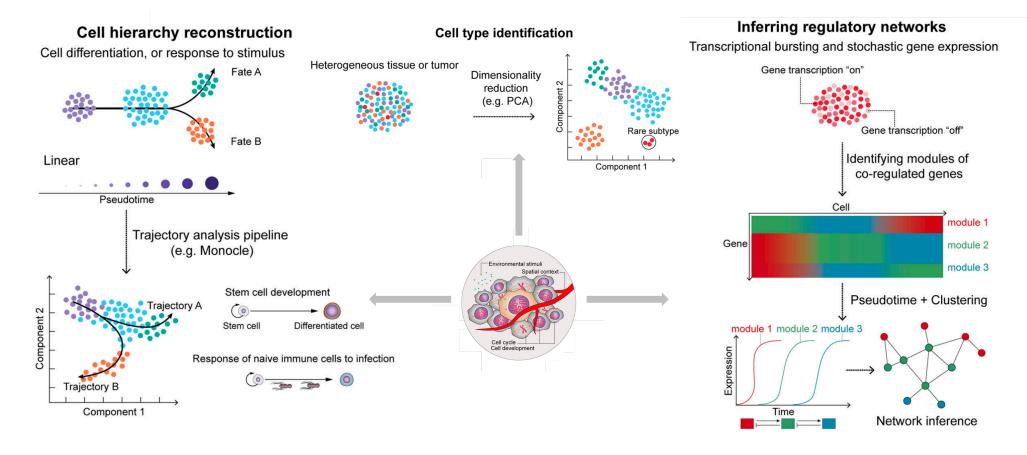
#### Problem of quantification of expression in scRNA-seq



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## **Discussion and Conclusions**

- Use of single-cell genomics technology has grown rapidly
  - Major advances in the SCG methodology
  - Applications to diverse fields of biological research
- SCG allows:
  - Analysis of single-cell variants in both normal and disease states
  - Identification of new biomarkers for disease diagnosis, staging of disease progression, and prediction of optimal treatment regimen
- Has potential in the future to generate a much more comprehensive understanding of hereditary and somatic genetics

## **Discussion and Conclusions**

### What is next?

- Standardized protocols for reproducibility of studies and comparison of data sets generated by different groups
- Improved sequencing technologies and better amplification methods that minimize errors
- Development of improved methods coupling single-cell and bulk NGS data
- SCG coupled with other multiomics technologies

## **Discussion and Conclusions**

#### Limitations:

- Various artifacts caused by suboptimal sample sizes, nonrepresentative starting samples, and other issues
- Difficult to interpret increasingly larger data sets

#### **Discussion Questions:**

What other limitations exist? What do you think is the next step? How do we select between bulk vs single-cell? Can we combine bulk and single-cell? Can we combine different omics data?