CSE 527

Markov Models and Hidden Markov Models



http://upload.wikimedia.org/wikipedia/commons/b/ba/Calico_cat

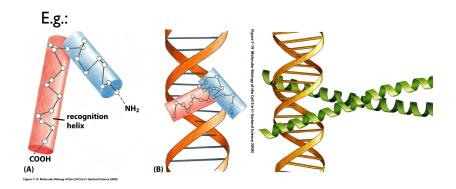
Dosage Compensation and X-Inactivation

2 copies (mom/dad) of each chromosome I-23
Mostly, both copies of each gene are expressed
E.g., A B O blood group defined by 2 alleles of I gene
Women (XX) get double dose of X genes (vs XY)?
So, early in embryogenesis:

- One X randomly inactivated in each cell | How?
- Choice maintained in daughter cells

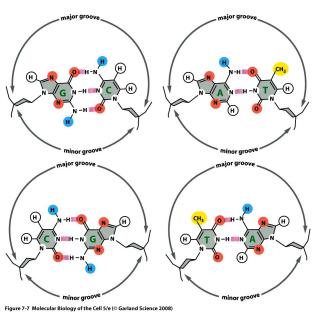
Calico: major coat color gene is on X

Reminder: Proteins "Read" DNA



Down in the Groove

Different patterns of hydrophobic methyls, potential H bonds, etc. at edges of different base pairs. They're accessible, esp. in major groove



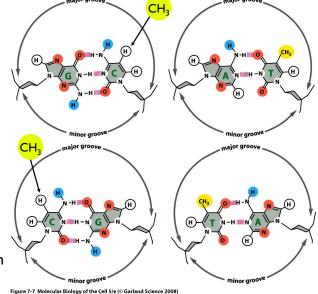
DNA Methylation

CpG - 2 adjacent nts, same strand (not Watson-Crick pair; "p" mnemonic for the phosphodiester bond of the DNA backbone)

C of CpG is often (70-80%) methylated in mammals i.e., CH₃ group added (both strands)

Same Pairing

Methyl-C alters major groove profile (:. TF binding), but not basepairing, transcription or replication



DNA Methylation–Why

In vertebrates, it generally silences transcription

(Epigenetics) X-inactivation, imprinting, repression of mobile elements, cancers, aging, and developmental differentiation



E.g., if a stem cell divides, one daughter fated to be liver, other kidney, need to

- (a) turn off liver genes in kidney & vice versa,
- (b) remember that through subsequent divisions

How?

- (a) Methylate genes, esp. promoters, to silence them
- (b) after ÷, DNA methyltransferases convert hemi- to fully-methylated (& deletion of methyltransferse is embrionic-lethal in mice)

Major exception: promoters of housekeeping genes

"CpG Islands"

Methyl-C mutates to T relatively easily

Net: CpG is less common than expected genome-wide: f(CpG) < f(C)*f(G)

BUT in some regions (e.g. active promoters), CpG remain unmethylated, so CpG → TpG less likely there: makes "CpG Islands"; often mark gene-rich regions

CpG Islands

CpG Islands

More CpG than elsewhere (say, CpG/GpC>50%) More C & G than elsewhere, too (say, C+G>50%) Typical length: few 100 to few 1000 bp

Ouestions

Is a short sequence (say, 200 bp) a CpG island or not? Given long sequence (say, 10-100kb), find CpG islands?

Markov & Hidden Markov Models

References (see also online reading page): Eddy, "What is a hidden Markov model?" Nature Biotechnology, 22, #10 (2004) 1315-6.

Durbin, Eddy, Krogh and Mitchison, "Biological Sequence Analysis", Cambridge, 1998 (esp. chs 3, 5)

Rabiner, "A Tutorial on Hidden Markov Models and Selected Application in Speech Recognition," Proceedings of the IEEE, v 77 #2,Feb 1989, 257-286

Independence

A key issue: Previous models we've talked about assume *independence* of nucleotides in different positions - definitely unrealistic.

Markov Chains

A sequence x_1, x_2, \ldots of random variables is a k-th order Markov chain if, for all i, ith value is independent of all but the previous k values:

$$P(x_i \mid x_1, x_2, \dots, x_{i-1}) = P(x_i \mid x_{i-k}, x_{i-k+1}, \dots, x_{i-1})$$

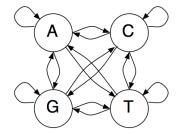
Example 1: Uniform random ACGT

Example 2: Weight matrix model

Example 3: ACGT, but \(\psi \) Pr(G following C)

Ist

A Markov Model (1st order)

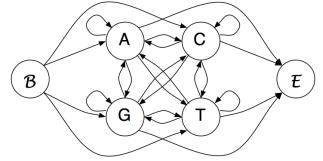


States: A,C,G,T

Emissions: corresponding letter

Transitions: $a_{st} = P(x_i = t \mid x_{i-1} = s)$ \leftarrow Ist order

A Markov Model (1st order)



States: A,C,G,T

Emissions: corresponding letter Transitions: $a_{st} = P(x_i = t \mid x_{i-1} = s)$

Begin/End states

Pr of emitting sequence x

$$x = x_1 x_2 \dots x_n$$
 $P(x) = P(x_1, x_2, \dots, x_n) > x_n$
 $= P(x_1) \cdot P(x_2 \mid x_1) \cdots P(x_n \mid x_{n-1}, \dots, x_1)$
 $= P(x_1) \cdot P(x_2 \mid x_1) \cdots P(x_n \mid x_{n-1})$
 $= P(x_1) \prod_{i=1}^{n-1} a_{x_i, x_{i+1}}$
 $= \prod_{i=0}^{n-1} a_{x_i, x_{i+1}}$ (with Begin state)

Training

Max likelihood estimates for transition probabilities are just the frequencies of transitions when emitting the training sequences

E.g., from 48 CpG islands in 60k bp:

+	A	C	G	T	-	A	C	G	T
A	0.180	0.274	0.426	0.120	A	0.300	0.205	0.285	0.210
C	0.171	0.368	0.274	0.188	C	0.322	0.298%	0.078	0.302
G	0.161	0.339	0.375	0.125	G	0.248	0.246	0.298	0.208
Т	0.079	0.355	0.384	0.182	T	0.177	0.239	0.292	0.292
								Fr	om DEKM

Discrimination/Classification

Log likelihood ratio of CpG model vs background model

$$S(x) = \log \frac{P(x|\text{model } +)}{P(x|\text{model } -)} = \sum_{i=1}^{L} \log \frac{a_{x_{i-1}x_i}^+}{a_{x_{i-1}x_i}^-} = \sum_{i=1}^{L} \beta_{x_{i-1}x_i}$$

β	A	С	G	T
A	-0.740	0.419	0.580	-0.803
C	-0.913	0.302	1.812	-0.685
G	-0.624	0.461	0.331	-0.730
T	-1.169	0.573	0.393	-0.679

From DEKM

CpG Island Scores

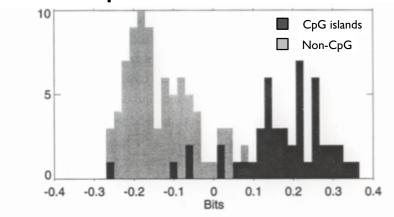


Figure 3.2 Histogram of length-normalized scores.

What does a 2nd order Markov Model look like?

3rd order?

From DEKM

Questions

Q1: Given a *short* sequence, is it more likely from feature model or background model? Above

Q2: Given a *long* sequence, where are the features in it (if any)

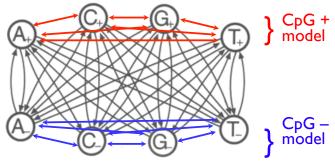
Approach I: score 100 bp (e.g.) windows

Pro: simple

Con: arbitrary, fixed length, inflexible

Approach 2: combine +/- models.

Combined Model



Emphasis is "Which (hidden) state?" not "Which model?"

Hidden Markov Models

(HMMs; Claude Shannon, 1948)

States: $1, 2, 3, \ldots$

Paths: sequences of states $\pi = (\pi_1, \pi_2, ...)$

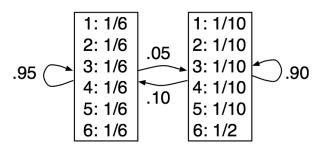
Transitions: $a_{k,l} = P(\pi_i = l \mid \pi_{i-1} = k)$ Emissions: $e_k(b) = P(x_i = b \mid \pi_i = k)$

Observed data: emission sequence

Hidden data: state/transition sequence

The Occasionally Dishonest Casino

1 fair die, 1 "loaded" die, occasionally swapped



Rolls Die Viterbi	315116246446644245311321631164152133625144543631656626566666 FFFFFFFFFFFFFFFFFFFFFFFFFFF
Rolls Die Viterbi	651166453132651245636664631636663162326455236266666625151631 LLLLLLFFFFFFFFFFFLLLLLLLLLLLLLLLLLLLL
Rolls Die Viterbi	$222555441666566563564324364131513465146353411126414626253356\\ FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF$
Rolls Die Viterbi	36616366466232534413661661163252562462255265252266435353336 LLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFF
Rolls Die Viterbi	233121625364414432335163243633665562466662632666612355245242 FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF

Figure 3.5

Rolls: Visible data—300 rolls of a die as described above. Die: Hidden data—which die was actually used for that roll (F = fair, L = loaded). Viterbi: the prediction by the Viterbi algorithm is shown.

From DEKM

The Viterbi Algorithm: The most probable path

Viterbi finds: $\pi^* = \arg \max_{\pi} P(x, \pi)$

Possibly there are 1099 paths of prob 10-99

More commonly, one path (+ slight variants) dominate others.

(If not, other approaches may be preferable.)

Key problem: exponentially many paths π

Inferring hidden stuff

Joint probability of a given path π & emission sequence x:

$$P(x,\pi) = a_{0,\pi_1} \prod_{i=1}^{n} e_{\pi_i}(x_i) \cdot a_{\pi_i,\pi_{i+1}}$$

But π is hidden; what to do? Some alternatives:

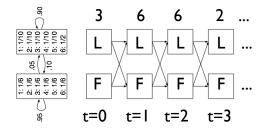
Most probable single path

$$\pi^* = \arg\max_{\pi} P(x, \pi)$$

Sequence of most probable states

$$\hat{\pi}_i = \arg\max_k P(\pi_i = k \mid x)$$

Unrolling an HMM



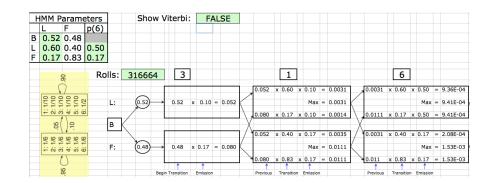
Conceptually, sometimes convenient Note exponentially many paths

Viterbi

 $v_l(i) = ext{probability of the most probable path}$ emitting x_1, x_2, \dots, x_i and ending in state l

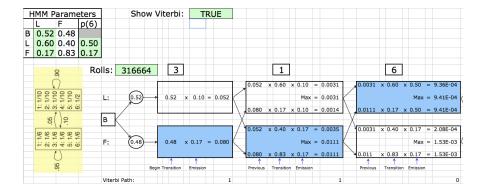
Initialize:

HMM Casino Example



(Excel spreadsheet on web; download & play...)

HMM Casino Example



(Excel spreadsheet on web; download & play...)

Viterbi Traceback

Above finds *probability* of best path

To find the path itself, trace *backward* to the state *k* attaining the max at each stage

$$v_l(i+1) = e_l(x_{i+1}) \cdot \max_k(v_k(i) a_{k,l})$$

Rolls Die Viterbi	315116246446644245311321631164152133625144543631656626566666666666666666666666666666
Rolls Die Viterbi	651166453132651245636664631636663162326455236266666625151631 LLLLLLFFFFFFFFFFLLLLLLLLLLLLLLLLLLLLL
Rolls Die Viterbi	$222555441666566563564324364131513465146353411126414626253356\\ FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF$
Rolls Die Viterbi	$36616366466232534413661661163252562462255265252266435353336\\ LLLLLLLLLFFFFFFFFFFFFFFFFFFFFFFFFFFFF$
Rolls Die Viterbi	$233121625364414432335163243633665562466662632666612355245242\\ FFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFFF$

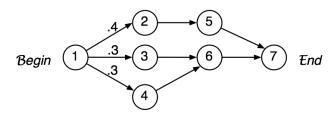
Figure 3.5

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From DEKM

Is Viterbi "best"?

Viterbi finds $\pi^* = \arg\max_{\pi} P(x,\pi)$



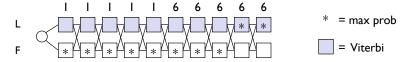
Most probable (Viterbi) path goes through 5, but most probable state at 2nd step is 6 (I.e., Viterbi is not the only interesting answer.)

Most probable path ≠ Sequence of most probable states

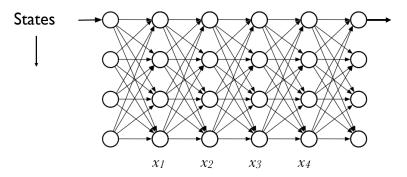
Another example, based on casino dice again

Suppose p(fair↔loaded) transitions are 10⁻⁹⁹ and

roll sequence is IIIII...66666; then fair state is more likely all through I's & well into the run of 6's, but eventually loaded wins, and the improbable $F \rightarrow L$ transitions make Viterbi = all L.

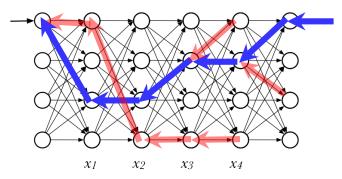


An HMM (unrolled)



Emissions/sequence positions ——

Viterbi: best path to each state



Viterbi score:

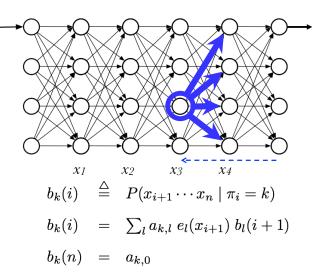
$$v_l(i+1) = e_l(x_{i+1}) \cdot \max_{k} (v_k(i) a_{k,l})$$

Viterbi path^R:

$$back_l(i+1) = \arg\max_{k} (v_k(i) a_{k,l})$$

The Backward Algorithm

Similar: for each state/time, want total probability of all paths from it, with given emissions, conditional on that state.



The Forward Algorithm

For each state/time, want total probability of all paths leading to it, with given emissions $f_k(i) \stackrel{\triangle}{=} P(x_1 \dots x_i, \ \pi_i = k)$ $f_l(i+1) = e_l(x_{i+1}) \sum_k f_k(i) a_{k,l}$ $P(x) = \sum_{x} P(x, \pi) = \sum_k f_k(n) a_{k,0}$

In state k at step i?

$$P(x, \pi_i = k)$$

$$= P(x_1, \dots, x_i, \pi_i = k) \cdot P(x_{i+1}, \dots, x_n \mid x_1, \dots, x_i, \pi_i = k)$$

$$= P(x_1, \dots, x_i, \pi_i = k) \cdot P(x_{i+1}, \dots, x_n \mid \pi_i = k)$$

$$= f_k(i) \cdot b_k(i)$$

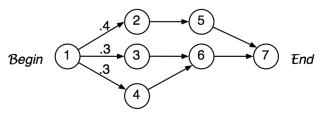
$$P(\pi_i = k \mid x) = \frac{P(x, \pi_i = k)}{P(x)} = \frac{f_k(i) \cdot b_k(i)}{P(x)}$$

Posterior Decoding, I

Alternative 1: what's the most likely state at step i?

$$\hat{\pi}_i = \arg\max_k P(\pi_i = k \mid x)$$

Note: the sequence of most likely states ≠ the most likely sequence of states. May not even be legal!



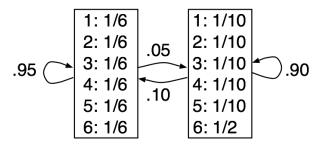
Rolls Die Viterbi	315116246446644245311321631164152133625144543631656626566666 FFFFFFFFFFFFFFFFFFFFFFFFFFF
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Figure 3.5

Rolls: Visible data—300 rolls of a die as described above. Die: Hidden data—which die was actually used for that roll (F = fair, L = loaded). Viterbi: the prediction by the Viterbi algorithm is shown.

The Occasionally Dishonest Casino

1 fair die, 1 "loaded" die, occasionally swapped



Posterior Decoding

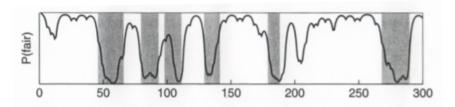


Figure 3.6 The posterior probability of being in the state corresponding to the fair die in the casino example. The x axis shows the number of the roll. The shaded areas show when the roll was generated by the loaded die.

From DEKM From DEKM

Posterior Decoding, II

Alternative 1: what's most likely state at step i?

$$\hat{\pi}_i = \arg\max_k P(\pi_i = k \mid x)$$

Alternative 2: given some function g(k) on states, what's its expectation. E.g., what's probability of "+" model in CpG HMM (g(k)=1) iff k is "+" state)?

$$G(i \mid x) = \sum_{k} P(\pi_i = k \mid x) \cdot g(k)$$

Training

Given model topology & training sequences, learn transition and emission probabilities

If π known, then MLE is just frequency observed in training data

$$a_{k,l} = \frac{\text{count of } k o l \text{ transitions}}{\text{count of } k o \text{ anywhere transitions}}$$
 $e_k(b) = \dots$

If π hidden, then use EM: given π , estimate θ ; given θ estimate π .

2 ways

CpG Islands again

Data: 41 human sequences, totaling 60kbp, including 48 CpG islands of about 1kbp each

Viterbi: Post-process:

Found 46 of 48 46/48

plus 121 "false positives" 67 false pos

Posterior Decoding:

same 2 false negatives 46/48

plus 236 false positives 83 false pos

Post-process: merge within 500; discard < 500

Viterbi Training

given π , estimate θ ; given θ estimate π

Make initial estimates of parameters θ Find Viterbi path π for each training sequence Count transitions/emissions on those paths, getting new θ Repeat

Not rigorously optimizing desired likelihood, but still useful & commonly used.

(Arguably good if you're doing Viterbi decoding.)

Baum-Welch Training

EM: given θ , estimate π ensemble; then re-estimate θ

$$P(\pi_{i} = k, \, \pi_{i+1} = l \mid x, \theta)$$

$$= \frac{f_{k}(i \mid \theta) \, a_{k,l} \, e_{l}(x_{i+1}) \, b_{l}(i+1 \mid \theta)}{P(x \mid \theta)}$$

Estimated # of k o l transitions $\hat{A}_{k,l}$

$$= \sum_{\mathsf{training seqs } x^j} \sum_i P(\pi_i = k, \, \pi_{i+1} = l \mid x^j, \theta)$$
 New estimate $\hat{a}_{k,l} = \frac{\hat{A}_{k,l}}{\sum_l \hat{A}_{k,l}}$

Emissions: similar

HMMs in Action: Pfam

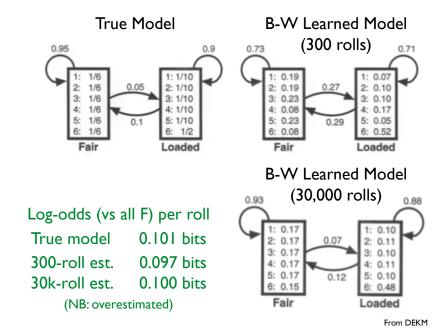
http://pfam.sanger.ac.uk/

Proteins fall into families, both across & within species

Ex: Globins, GPCRs, Zinc fingers, Leucine zippers,...

Identifying family very useful: suggests function, etc.

So, search & alignment are both important
One very successful approach: profile HMMs



Helix HBA_HUMAN -----VLSPADKTNVKAAWGKVGA--HAGEYGAEALERMFLSFPTTKTYFPHF HBB_HUMAN -----VHLTPEEKSAVTALWGKV----NVDEVGGEALGRLLVVYPWTQRFFESF MYG_PHYCA -----VLSEGEWQLVLHVWAKVEA--DVAGHGODILIRLFKSHPETLEKFDRF GLB3_CHITP -----LSADQISTVQASFDKVKG-----DPVGILYAVFKADPSIMAKFTOF GLB5_PETMA PIVDTGSVAPLSAAEKTKIRSAWAPVYS--TYETSGVDILVKFFTSTPAAOEFFPKF LGB2_LUPLU -----GALTESQAALVKSSWEEFNA--NIPKHTHRFFILVLEIAPAAKDLFS-F GLB1_GLYDI -----GLSAAQRQVIAATWKDIAGADNGAGVGKDCLIKFLSAHPQMAAVFG-F Consensus Ls.... va W kv. Helix DDDDDDDEEEEEEEEEEEEEEE HBA_HUMAN -DLS----HGSAQVKGHGKKVADALTNAVAHV---D--DMPNALSALSDLHAHKL-HBB_HUMAN GDLSTPDAVMGNPKVKAHGKKVLGAFSDGLAHL---D--NLKGTFATLSELHCDKL-MYG_PHYCA KHLKTEAEMKASEDLKKHGVTVLTALGAILKK----K-GHHEAELKPLAQSHATKH-GLB3_CHITP AG-KDLESIKGTAPFETHANRIVGFFSKIIGEL--P---NIEADVNTFVASHKPRG-GLB5_PETMA KGLTTADQLKKSADVRWHAERIINAVNDAVASM--DDTEKMSMKLRDLSGKHAKSF-LGB2_LUPLU LK-GTSEVPQNNPELQAHAGKVFKLVYEAAIQLQVTGVVVTDATLKNLGSVHVSKG-GLB1_GLYDI SG----AS---DPGVAALGAKVLAQIGVAVSHL--GDEGKMVAQMKAVGVRHKGYGN Consensus .. . v..Hg kv. a a...l d Helix FFGGGGGGGGGGGGGG ннинининининининининини HBA_HUMAN -RVDPVNFKLLSHCLLVTLAAHLPAEFTPAVHASLDKFLASVSTVLTSKYR---HBB HUMAN -HVDPENFRLLGNVLVCVLAHHFGKEFTPPVQAAYQKVVAGVANALAHKYH------KIPIKYLEFISEAIIHVLHSRHPGDFGADAQGAMNKALELFRKDIAAKYKELGYQG MYG_PHYCA GLB3 CHITP --VTHDQLNNFRAGFVSYMKAHT--DFA-GAEAAWGATLDTFFGMIFSKM-----

Alignment of 7 globins. A-H mark 8 alpha helices. Consensus line: upper case = 6/7, lower = 4/7, dot=3/7. Could we have a profile (aka weight matrix) w/ indels?

GLB5_PETMA -QVDPQYFKVLAAVIADTVAAG------DAGFEKLMSMICILLRSAY------LGB2_LUPLU --VADAHFPVVKEAILKTIKEVVGAKWSEELNSAWTIAYDELAIVIKKEMNDAA---

GLB1_GLYDI KHIKAQYFEPLGASLLSAMEHRIGGKMNAAAKDAWAAAYADISGALISGLOS--

f 1

Profile Hmm Structure

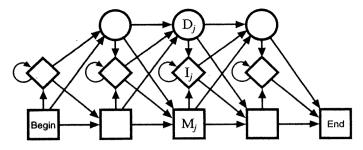


Figure 5.2 The transition structure of a profile HMM.

M_j: Match states (20 emission probabilities)

lj: Insert states (Background emission probabilities)

Dj: Delete states (silent - no emission)

From DEKM

Silent States

Example: chain of states, can skip some

Problem: many parameters.

A solution: chain of "silent" states; fewer parameters (but less detailed control)

Algorithms: basically the same.

Using Profile HMM's

Search

Forward or Viterbi

Scoring

Log likelihood (length adjusted)

Log odds vs background

Z scores from either

next slides

Alignment

Viterbi

Likelihood vs Odds Scores

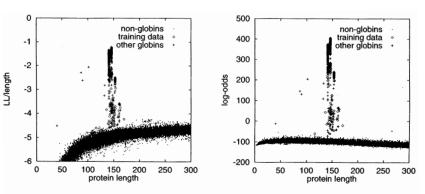


Figure 5.5 To the left the length-normalized LL score is shown as a function of sequence length. The right plot shows the same for the log-odds score.

Z-Scores

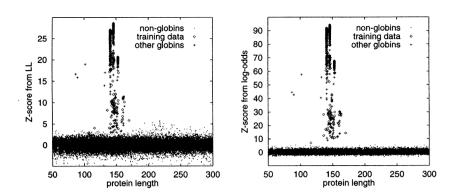


Figure 5.6 The Z-score calculated from the LL scores (left) and the log-odds (right).

From DEKM

Model-building refinements

Pseudocounts (count = 0 common when training with 20 aa's)

$$e_i(a)=rac{C_{i,a}+A\cdot q_a}{\sum_a C_{i,a}+A},~~A\sim 20,~q_a=~$$
 background (~50 training sequences)

Pseudocount "mixtures", e.g. separate pseudocount vectors for various contexts (hydrophobic regions, buried regions,...)

(~10-20 training sequences)

Pfam Model Building

Hand-curated "seed" multiple alignments
Train profile HMM from seed alignment
Hand-chosen score threshold(s)
Automatic classification/alignment of all other protein sequences
11912 families in Rfam 24.0, 10/200
(covers ~75% of proteins)

More refinements

Weighting: may need to down weight highly similar sequences to reflect phylogenetic or sampling biases, etc.

Match/insert assignment: Simple threshold, e.g. "> 50% gap ⇒ insert", may be suboptimal.

Can use forward-algorithm-like dynamic programming to compute max a posteriori assignment.

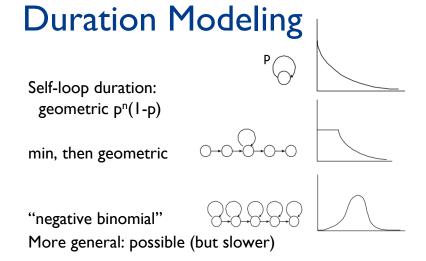
Numerical Issues

Products of many probabilities → 0
For Viterbi: just add logs
For forward/backward: also work with logs, but you need sums of products, so need "log-of-sum-of-product-of-exp-of-logs", e.g., by table/interpolation
Keep high precision and perhaps scale factor
Working with log-odds also helps.

Model structure

Define it as well as you can.

In principle, you can allow all transitions and hope to learn their probabilities from data, but it usually works poorly – too many local optima



HMM Summary

(max of products)

(sum of products)

joint vs onditional probs Inference

Viterbi – best single path

Forward – sum over all paths

Backward – similar

Posterior decoding

Model building

Semi-supervised – typically fix architecture (e.g. profile HMM), then learn parameters

Baum-Welch – training via EM and forward/backward (aka the forward/backward algorithm)

Viterbi training - also "EM", but Viterbi-based

HMM Summary (cont.)

```
Search:
Viterbi or forward

Scoring:
Odds ratio to background
Z-score
E-values, etc., too

Excellent tools available (SAM, HMMer, Pfam, ...)

A very widely used tool for biosequence analysis
```