Motif-Finding in Trypanosomatids

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Presentation Agenda

Introduction to Trypanosomatids and their genomes

 Algorithms to explore motifs in Trypanosomes

Results and conclusion

Trypanosomes in the world



- Family of parasites
- Human infective 12 million affected by *Leishmania* species of Trypanosomes alone
- Infection can be asymptomatic to deadly
- 2 million new cases every year, estimated by World Health Organization

Genome makeup

Leishmania major

○~33.6 megabase genome

- 36 chromosomes of sizes 300 to 2800 kilobaes
- Ochromosome 1: ~85 protein-coding ORFs

Trypanosoma brucei

- ○~35 megabase genome
- 11 chromosomes (larger than *L. major*)
- Ochromosome 1: ~145 protein-coding ORFs
- Overwhelming majority of genomes have been annotated in silico

Genome structure



- High conservation between related species
- Very syntenous despite divergence
- What else is shared?

Genome characteristics



- Gene organization follows a polycistronic structure
- Predictable via GC-content skew

Algorithms

Gibbs Sampling (Lawrence, 1993)
 Variations of Gibbs Sampling

 AlignACE
 GLAM (gapless local alignment of multiple sequences)

 Mismatch Tree Algorithm (MITRA)

Gibbs Sampling (review)

 Goal: locate the alignment that maximizes the ratio of the pattern probability to background probability

$$F = \sum_{i=1}^{W} \sum_{j=1}^{4} c_{i,j} \log \frac{q_{i,j}}{p_{j}}$$

Gibbs Sampling – basic algorithm

• Predictive update step:

- Choose one random sequence z, and random starting positions within the various sequences.
- Calculate pattern probability and background probability at current positions
- Sampling step:
 - Calculate probabilities of generating every possible segment of width W within sequence z according to the current pattern probability (Q), and the background probability (P).
 - The weight A = Q/P is assigned to each segment and a random one is selected for the next iteration.

AlignACE

Based on Gibbs sampling

• Differences:

- Both strands of the input sequences are considered
- Simultaneous vs. single motif searching: masking
- MAP score (maximum *a priori* log likelihood):
 - Degree of which a motif is over-represented relative to the expectation of the random occurrence in sequence
 - Drawbacks: ubiquitous but not relevant motifs

GLAM

Based on Gibbs sampling
 Bayesian scoring scheme
 Prior probability distribution
 Dirichlet function

$$prior\{q_i\} = \frac{1}{Z} \prod_i q_i^{\alpha_i - 1}$$

GLAM – alignment score

Scoring scheme:

$$S = \sum_{k=1}^{W} \ln \left[\frac{\prod_{i} \Gamma(\alpha_{i})}{\prod_{i} \Gamma(\alpha_{i})} \frac{\prod_{i} \Gamma(c_{ki} + \alpha_{i})}{\Gamma(N + A)} \right]$$

GLAM – resizing

 Automatic adjustment of width of alignment

- ○Fix left ends and right ends are varied
- OFix right ends and left ends are varied
- Over the problem of fixed width algorithm where end points are shifted left or right relative to the optimal

MITRA

Definitions:

- search of all L-mers (a continuous string of length L) that occur with up to d mismatches in at least k sequences in the sample S.
- Weak pattern: has less than k (L, d)-neighbors (all possible L-mers with up to d mismatches as compared to the canonical pattern) in the sample
- Weak subspace: all patterns are weak
- Data Structure:
 - Rooted tree where each node has 4 branches {A, C, G, T}
 - OMaximum depth: L

MITRA – algorithm: 1

Search for (8, 1) motif for a sequence AGTATCAGTT



Initial State

MITRA – algorithm: 2



MITRA – algorithm:3



MITRA-graph

Pairwise similarity match

- OGraph G(P, S)
- OVertex: L-mer in the sample
- OEdge: if two L-mers are similar
- Subspace is empty if clique of size k does not exist
- More efficient pruning of mismatch tree

In search of common motifs

- Ran GLAM, AlignACE and MITRA motif-finding programs on upstream non-coding regions of annotated genes
 - OGLAM: http://zlab.bu.edu/glam/ (binary)
 - OAlignACE: <u>http://atlas.med.harvard.edu/</u> (binary)
 - OMITRA:

http://fluff.cs.columbia.edu:8080/domain/mitra.html (webpage)

Parsed out sequences, generated WMMs

 Hypothesis: outstanding motifs will appear in 2-3 of the algorithms (detection via consensus/overlap)

AlignACE results

Data profile:

- O Max = 5272.33 (LmjF)
- 21 motifs in LmjF, 100 motifs in Tb
- Repetitious motifs frequent in short windows
- Large number of simple repeating sequences (e.g. ACACAC.., AGAGAG..)

AlignACE WMM

MOLII	#0:								
	A	С	Т	G	N				
1	0.03		0.38	0.03		0.57	0.00	G	
2	0.28		0.20	0.19		0.33	0.00	*	
3	0.22		0.26	0.25		0.27	0.00	*	
4	0.01		0.18	0.01		0.80	0.00	G	
5	0.06		0.38	0.16		0.41	0.00	*	
6	0.09		0.37	0.00		0.54	0.00	G	
7	0.25		0.21	0.29		0.24	0.00	*	
8	0.04		0.14	0.03		0.80	0.00	G	
9	0.00		0.77	0.00		0.23	0.00	С	
10	0.20		0.13	0.00		0.66	0.00	G	
11	0.00		0.52	0.12		0.35	0.00	С	
12	0.11		0.36	0.00		0.53	0.00	G	
13	0.08		0.41	0.09		0.42	0.00	C/G	

Trypanosome motifs in AlignACE

Commonalities

- OG*G*G.. repeating pattern common to both L. major and T. brucei
- Generally of type GAGA or GCGC

Differences

 T. brucei possessed high-scoring relatively complex repeating sequences, while L. major did not

Complicated reoccurring motifs in *T.* brucei

PIOLLI #1:

	A	с т	G	N		
1	1.00	0.00	0.00	0.00	0.00	А
2	0.00	0.00	0.00	1.00	0.00	G
3	0.00	0.00	1.00	0.00	0.00	Т
4	1.00	0.00	0.00	0.00	0.00	А
5	0.00	1.00	0.00	0.00	0.00	С
6	1.00	0.00	0.00	0.00	0.00	А
7	0.45	0.48	0.00	0.07	0.00	C/A
8	0.00	0.00	0.00	1.00	0.00	G
9	0.00	1.00	0.00	0.00	0.00	С
10	0.44	0.00	0.05	0.51	0.00	G/A
11	0.51	0.04	0.01	0.44	0.00	A/G
12	0.01	0.00	0.48	0.51	0.00	G/T
13	0.48	0.00	0.51	0.00	0.00	T/A
14	0.51	0.01	0.48	0.00	0.00	G/A
15	0.48	0.51	0.00	0.00	0.00	C/A
16	0.51	0.40	0.00	0.09	0.00	A/C
17	0.00	0.52	0.48	0.00	0.00	T/G
18	0.06	0.00	0.00	0.93	0.00	G
19	0.00	0.99	0.00	0.00	0.00	С

GLAM results



 Much less variability found than from AlignACE

Trypanosome motifs in GLAM

Commonalities

 Very few at the sequence level

 Differences

 L. major dominated by alternating bases
 T. brucei dominated by repeating adenine sequences (possibly poly-A tails?)

GLAM: L. major vs. T. brucei

AGAGAGAGAGAGAGAGAGAG GGAGAGAAAGACAAGCGGAG GCAGAGAGAGAGAGAGAGAGA AGAACGAGAGACGGACACAG ACGACGAGAGAGAGAGAGAA AGAAAGAGAGACGGAGAGAG GCGGAGAAAGGGAAAAGGAG GCAGAGCGAGAGAGCGAGAG ACAGAGAGAGAGAGAGAGAG ACACAGAGAGAGAGAGAGAG ACAGCGCGAGACAGAGGGAG AGAAAGAGAGAGAGAGGCA AGAGAGAGAGAGAGAGAGAG AGAGAGAGAGAGAGCGGGGG AGAAGAAGAAGAAGGGGGAG AGAAAGAGAGAGAGAGAGAG AGGAAGAGAGAAAGAGAGAA AGAGAGAGAGAGAGAGAGAG AGAGAGAGAGAGAGAGAGAG

L. major

AAAAAAGAAAAGAAAAAACAA AAAAAAAGAAAGGAAAAGAA AAGAAAAGAAAAGAAAAGAA GAAAGAAAAAAAAAGACAG AAGAAAAAAAAGGAAAAAGGA GAAAGAAATAATGAAGAGAA AAAAAAAAAAAAGAGAAAGAA AAAGGAGGAAAAAACAAAAG GAAAAAACAAAAGAAAACAT AAAAGATATAAAAGAGGAAA CAAAAAAAAAAAAGTAGAGGC GAAAAAGAAAAAGAAGGGGA CAAAAAATAAATGGCAACAA GAAAAAAACAAAAGAAAACA GAAAGATATAAAAGAGGAAA

T. brucei

MITRA results

Data profile:

OWeb interface returned A LOT of data

OMax = 35.0

Motifs mostly ATAT variants

- Some ACAC, AGAG seen as in AlignACE and GLAM
- Notable limitation web interface had sequence size limitation

Trypanosome motifs in MITRA

Commonalities

 NTNT, NTTNTT patterns

Differences

T. brucei included results with repeating adenine's (AAAANA, etc)

	А	С	Т	G	Ν			
1	0.25		0.25	0.25		0.25	0.00	*
2	0.50		0.25	0.25		0.00	0.00	A
3	0.00		0.00	0.75		0.25	0.00	Т
4	0.25		0.00	0.25		0.25	0.25	*
5	0.25		0.25	0.25		0.00	0.25	*
6	0.25		0.00	0.75		0.00	0.00	Т
7	0.25		0.25	0.25		0.25	0.00	*

Characterizing possible motifs across two genomes

 CACA, GAGA patterns very common in both genomes from all algorithms
 GAGA.. possible true motif within both

genomes

- **CACA..** perhaps maybe altered poly-A tail?
- Extremely high scores for possible motifs OAlignACE, GLAM had scores upwards of 2000+

Altering the experiment

Stuff to try in the future

- Use more of the genome, once its completely annotated officially
- Inclusion of other highly-conserved species in the Trypanosoma family
 - L. infantum, T. cruzi, etc.
- OPrune out possible poly-A regions to refine searching
- Apply or alter other algorithms to increase breadth of search
- Connect found motifs to gene function is there a relation?



Questions?