

# CSE 428

Spring 2021

<https://courses.cs.washington.edu/courses/cse428/21sp/>

### Course Web Pages:

<https://courses.cs.washington.edu/courses/cse428/21sp/>

### TA:

Alyssa La Fleur

### Group-Project-oriented:

Typically teams of ~2-4 students

We will offer some projects ideas

We are open to student-generated ideas

“computers” + “biology”

(+ reasonable scope + something we can facilitate)

Organization & Scheduling

Bio Jargon

Tools from elsewhere

Did I mention Organization & Scheduling?

*The #1 challenge identified by 99% of former students*

See previous slide!

You'll see real DNA/RNA seq data in all of them, plus

Some mixture of:

- data structures,

- algorithms,

- data analytics,

- statistics,

- biology,

- HCI,

- ML, ...

Weekly Goals + Progress reports

Some midcourse checkpoint

Final written reports + oral presentations

Including evaluation of code, test results, etc.

Peer comments

# Project Idea: Next Few Slides

Open-ended, underspecified; as you think about them, both let your imagination run free, *and* think carefully about how to scale and stage your project so you can collect low-hanging fruit before potentially getting lost in the open-ended weeds. (Fortunately, I don't think mixing metaphors is a crime in this state—yet.)

# Misc. Projects From 428's Past

Just to give you some idea of scope, here are some projects from previous iterations of 428:

- Convenient web interface for "phylogenetic footprinting" in prokaryotes
- Build a genome assembler
- Machine learning applied to cancer genomics
- Convenient web interface for exploring "Foldit" results
- Visualization of technical biases in RNAseq
- Downstream impact of technical biases in RNAseq
- Crossover detection in DNAseq data

### Discovery of regulatory non-coding RNA ("ribosomal leaders") in lower Eukaryotes

- \* You might remember my "L19" example from 427 – in some bacteria, excess L19 down-regulates itself by binding to its own mRNA
- \* This kind of thing is widespread in Prokaryotes
- \* Few if any examples are known in Eukaryotes
- \* But I speculate that one group of single-celled Euks is a strong contender to show this behavior
- \* Chances are I'm wrong! But should be "fun" to try; you'll see real data, a variety of state-of-the-art algorithms, and methods to evaluate your results

Deep learning for non-coding RNA discovery and classification.

- \* ncRNA is an immense landscape that is radically changing our understanding of molecular biology, esp. regulation
- \* But really hard problems
- \* Deep learning (DL) is sweeping the world, on hard problems
- \* Initial results suggest DL can be faster and better at ncRNA discovery/classification tasks than classical statistical methods
- \* Do you believe it?
- \* Initial DL architectures seem pretty naive: can you improve them?
- \* Again, look at real algorithms, real data, cutting-edge problems

## 2021 Project Idea #3

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*You* may have an idea that you want to pursue, and preferably recruit a partner or three to help

Again, we're open to this, provided its got a reasonable blend of Comp + Bio, reasonably scoped, and something that we can facilitate.

**More Details**

**Idea #1:**

**Ribosomal Leaders**

High copy-number, multi-protein complexes may benefit from stoichiometric control of their components

Ribosomes are high copy-number, multi-protein complexes

Widespread use of "ribosomal leader" auto regulation in prokaryotes exemplifies this (but details vary)

Eukaryotes may accomplish this by other means, although a few putative examples are known

My suggestion: Unique biology of ciliates probably greatly exacerbates the stoichiometry problem, making them a prime target for discovery of Eukaryotic ribosomal leader auto-regulation.

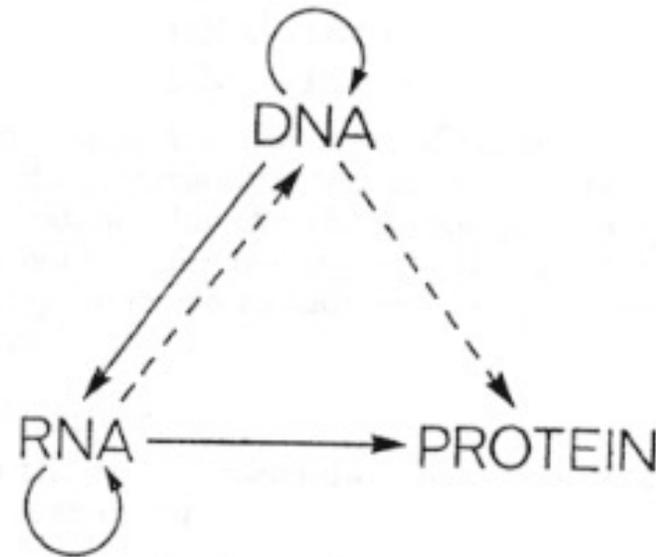
# Central Dogma of Molecular Biology

by  
FRANCIS CRICK  
MRC Laboratory  
Hills Road,  
Cambridge CB2 2QH

The central dogma of molecular biology deals with the detailed residue-by-residue transfer of sequential information. It states that such information cannot be transferred from protein to either protein or nucleic acid.

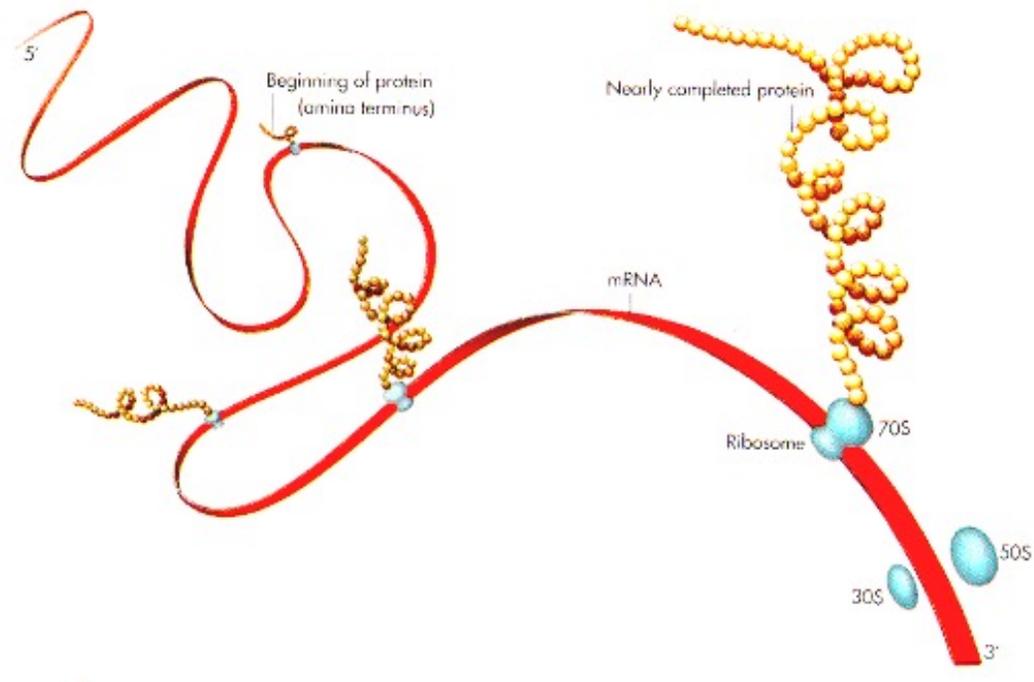
“The central dogma, enunciated by Crick in 1958 and the keystone of molecular biology ever since, is likely to prove a considerable over-simplification.”

Fig. 2. The arrows show the situation as it seemed in 1958. Solid arrows represent probable transfers, dotted arrows possible transfers. The absent arrows (compare Fig. 1) represent the impossible transfers postulated by the central dogma. They are the three possible arrows starting from protein.



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# Translation: mRNA → Protein



Watson, Gilman, Witkowski, & Zoller, 1992

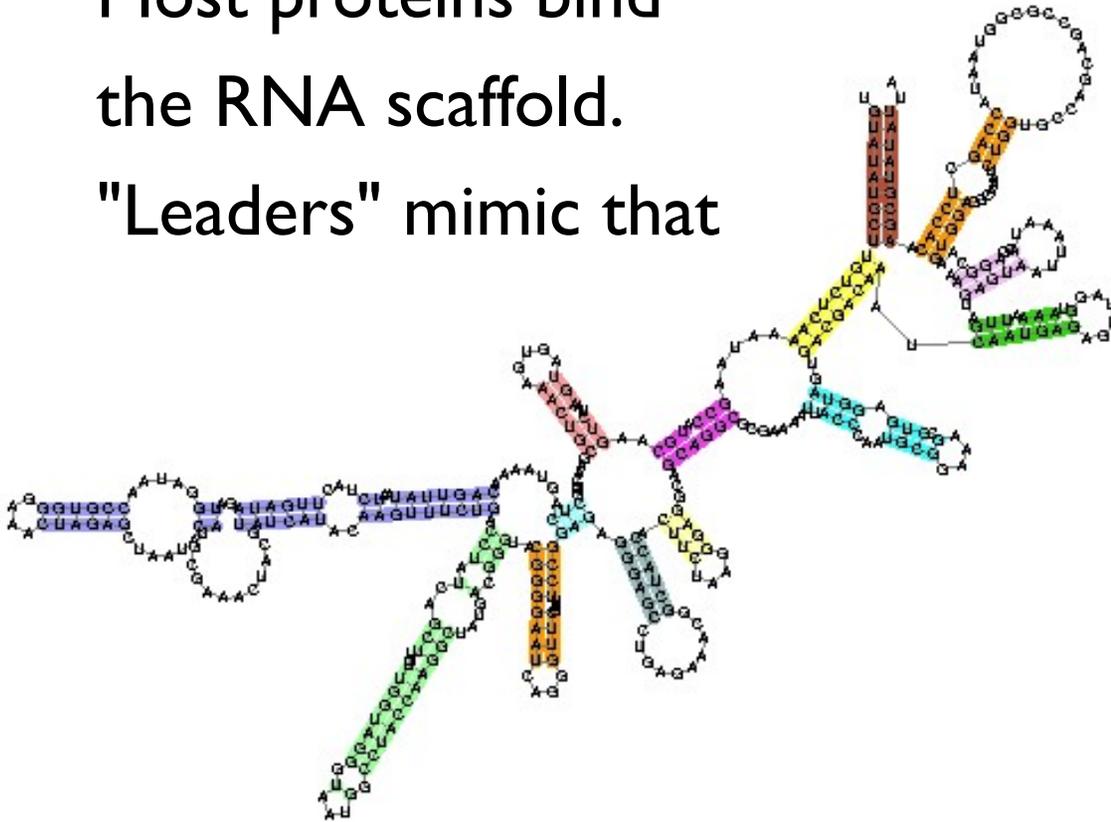
Ribosome is a large complex,

~ 1/2 RNA, 1/2 protein

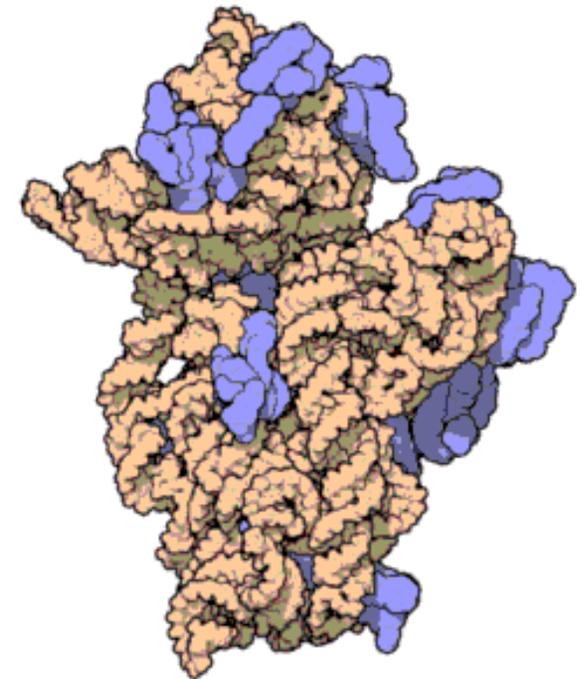
Most proteins bind

the RNA scaffold.

"Leaders" mimic that

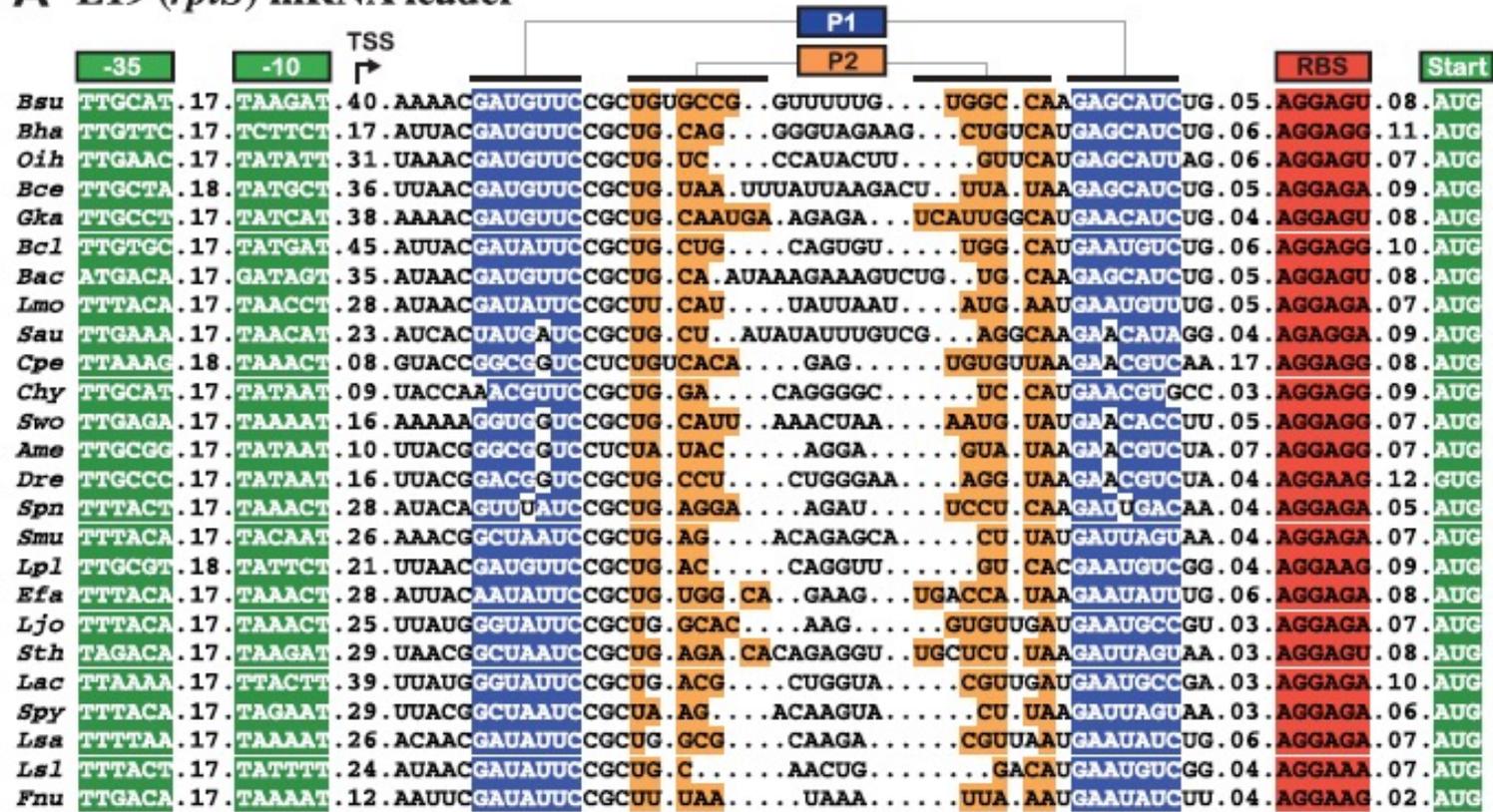


Small subunit ribosomal RNA, 5' domain taken from the [Rfam](#) database. This example is [RF00177](#), a fragment from an uncultured bacterium.

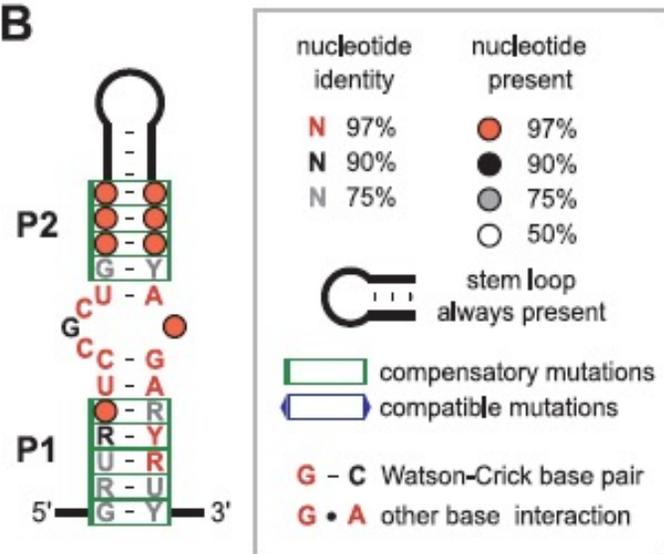


An example of a fully-assembled small subunit of ribosomal RNA in prokaryotes, specifically *Thermus thermophilus*. The actual ribosomal RNA (16S) is shown coiled in orange with ribosomal proteins attaching in blue.

# A L19 (*rplS*) mRNA leader



## B



## C

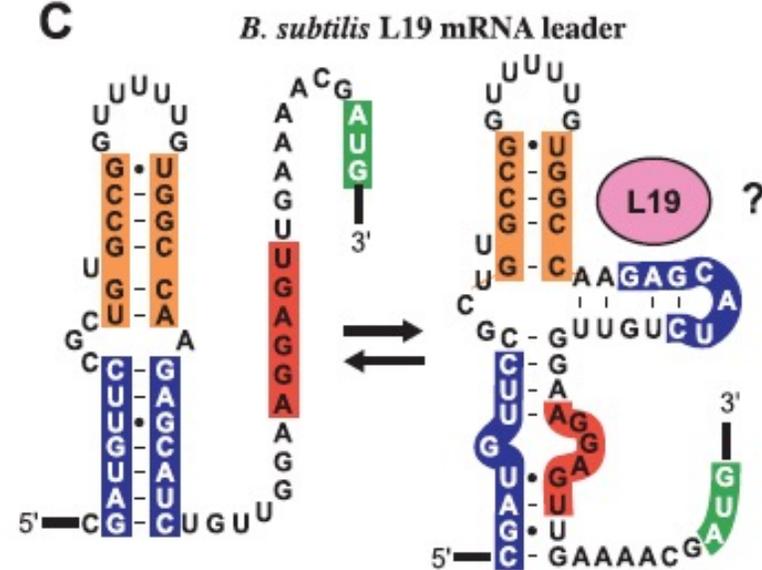
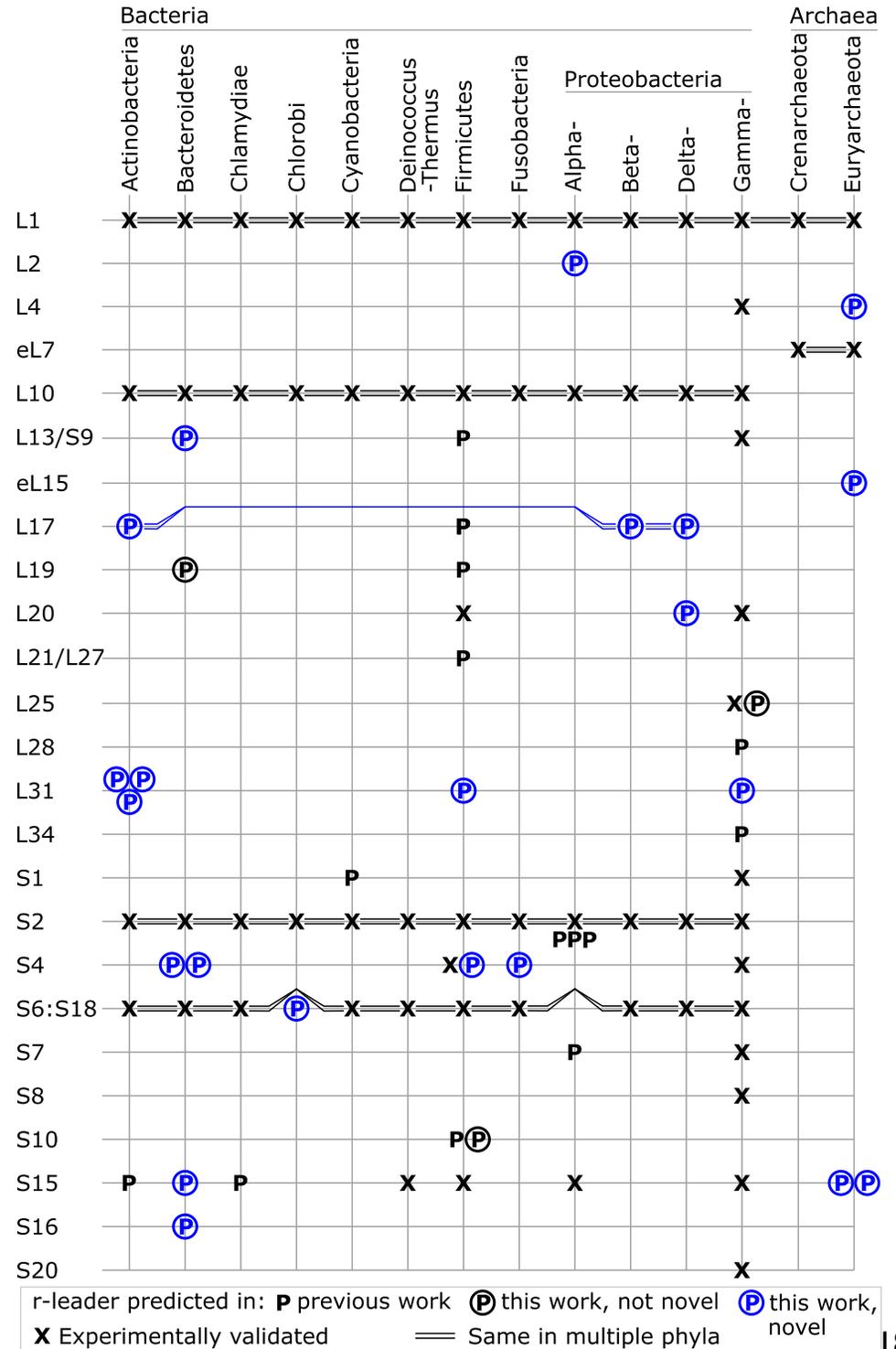
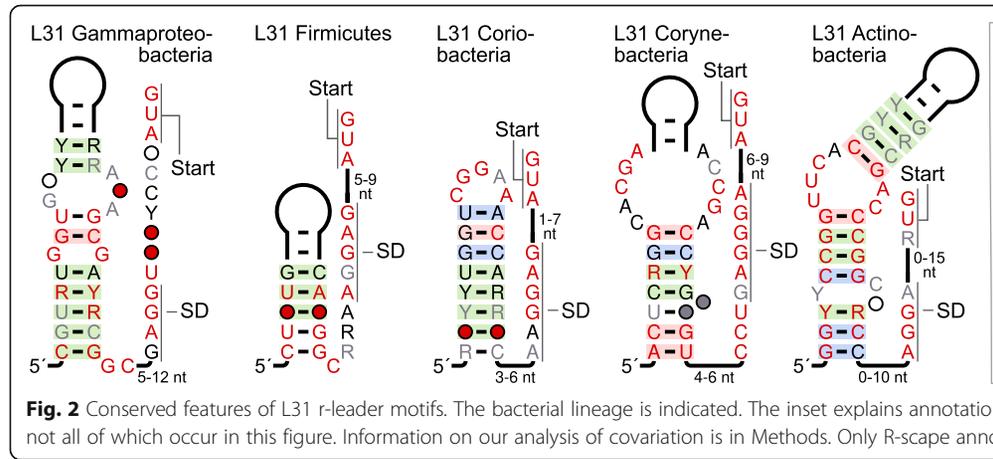


Figure 3. Putative Autoregulatory Structure in L19 mRNA Leaders

RESEARCH ARTICLE

# Discovery of 20 novel ribosomal leader candidates in bacteria and archaea

Iris Eckert and Zasha Weinberg\* 



Large, single-celled Eukaryotes

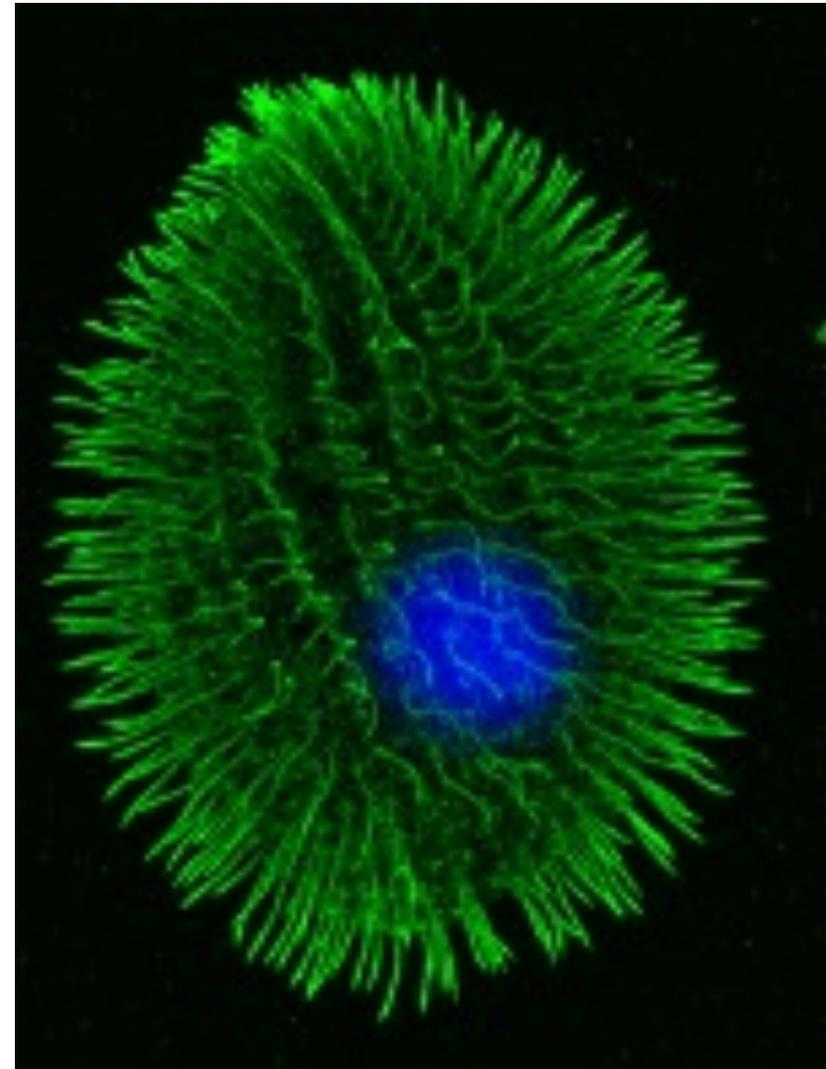
Reproduce by fission, with occasional sexual cycles ("conjugation")

Two nuclei:

MIC: typical diploid nuc with 5-10 pairs of large chromosomes, but *transcriptionally silent*

MAC: copy of MIC, but processed to have many copies of many mini-chromosomes (e.g. in *Tt*, several hundred, w/ ~45 copies of each, and  $>10^3$  copies of rDNA) *transcriptionally active*

In fission, MIC is usual mitotic div, but MAC is amitotic—pinches in half w/ semi-random segregation of chromosome copies to each daughter cell. (Restored by conjugation.)



*Tetrahymena thermophila*

Pick genes; for each:

Find genomes

Find chosen gene in multiple genomes

Download sequences (5' UTR, especially)

Validate them?

Prep them for ncRNA "discovery"

Predict:

CMfinder

Rscape, etc.

Evaluation!!

Multiperm, SSIz, control genes, outgroup, paralogs, visualize,  
Rscape, ...

Other directions: introns, 3' UTR, CDS, Other complexes  
(spliceosome, maybe? RNA or DNA Pol, telomerase?)

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Chances of success:

???

... But as Dr. Laughlin said, nature might have the last laugh. “Given the rule of thumb that 99 percent of one’s own cool ideas tend not to work out,” he said, “I think the smart money [is against us].” ...

<https://www.nytimes.com/2021/03/23/science/astronomy-oumuamua-comet.html?referringSource=articleShare>

**More Details**

**Idea #2:**

**Deep Learning for  
ncRNA Classification &  
Discovery**

See separate .pdf file linked from  
course home page as "Idea #2"

**Idea #3:  
Yours?**

# Next steps

review slides

which (if any) appeals?

form groups

skim references on web

talk to/email me/Alyssa

we may have fragments of code for parts of this  
(may or may not be useful...)

**Form a Group/Form a Plan!**

# Next Steps

Set up CSE GITLab Repo

Share with ruzzo@cs and lafleur1@cs (and teammates)

create a "group-info.txt" doc with

team members names & emails, other contact info  
members roles as they become defined

Create a "Progress" doc for *weekly reports*:

goals for next week

review of what did/didn't get accomplished last week

Bibliography

Make a "Plan" (what needs to be done, in what order);

revise it as you go

New Results

 [Comment on this paper](#)

 Previous

## Fifty generations of amitosis: tracing asymmetric allele segregation in polyploid cells with single-cell DNA sequencing

Posted March 30, 2021.

 Valerio Vitali,  Rebecca Rothering,  Francesco Catania

**doi:** <https://doi.org/10.1101/2021.03.29.437473>

This article is a preprint and has not been certified by peer review [what does this mean?].

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